

20. Brand P, Schulte M, Wencker M, Herpich CH, Klein G, Hanna K, et al. Lung deposition of inhaled alpha1-proteinase inhibitor in cystic fibrosis and alpha1-antitrypsin deficiency. *Eur Respir J*. 2009;34:354–60.

Gregory Reyhler,^{a,b,c,*} Teresinha Leal,^d Anne-Sophie Aubriot,^{b,e} Giuseppe Liistro^{a,c}

^a Institut de Recherche Expérimentale et Clinique (IREC), Pôle de Pneumologie, ORL & Dermatologie, Université Catholique de Louvain, Belgium

^b Service de Médecine Physique et Réadaptation, Cliniques universitaires Saint-Luc, Belgium

^c Service de Pneumologie, Cliniques universitaires Saint-Luc, Belgium

^d Louvain Centre for Toxicology and Applied Pharmacology, Institut de Recherche Expérimentale et Clinique, Secteur des Sciences de la Santé, Université Catholique de Louvain, Belgium

^e Centre de Référence pour la Mucoviscidose, Cliniques universitaires Saint-Luc, Belgium

* Corresponding author.

E-mail address: gregory.reyhler@uclouvain.be (G. Reyhler).

1579-2129/

© 2017 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

In Vitro and In Vivo Comparison of Two Nebulizers Used for Inhaled Pentamidine Delivery



Comparación in vitro e in vivo de dos nebulizadores utilizados para administrar pentamidina inhalada

Dear Editor,

Pneumocystis jirovecii pneumonia (PJP) represents a significant cause of morbidity and mortality in immunosuppressed patients.^{1,2} Pentamidine, used in secondary prevention of PJP, is administered via inhalation and requires a specific nebulizer.^{3,4} In the recent ERS/ISAM Task Force Consensus Statement, the RespigardII® (Vital Signs) was considered as the reference nebulizer to deliver pentamidine.⁵ Nebulizers with comparable properties are required because RespigardII® is no longer available since its recent withdrawal from the market in some countries.³ Most of the nebulizers compared previously to the RespigardII were ultrasonic nebulizers.⁶ In this study, we compared a jet nebulizer (ISO-NEB®, Teleflex) to RespigardII® for pentamidine delivery. Both nebulizers possess one-way valves on the inspiratory and on the expiratory way and an expiratory filter as recommended.^{3,4} Both were driven by a similar air flow (8 L/min) and deliver particles with similar size (MMAD: 1–2 µm).^{3,7,8} In vitro, nebulizers were connected to a dual chamber lung model (5600i Dual Adult Test Lung®, Michigan Instrument Inc.) driven by a ventilator (SERVO-1®, Maquet) in volume-controlled mode simulating an adult breathing pattern (Vt=500 mL; RF=15 breaths/min; I/E ratio=1:2; no end-inspiratory pause) (Fig. 1). Artificial lung compliance and resistance were set to 70 mL/cmH₂O and 5 cmH₂O, respectively. Nebulizations of a pentamidine solution (300 mg/6 mL sterile water) were performed in triplicate for each model in accordance

with manufacturer's and guidelines recommendations until one minute after the appearance of the sputtering point.^{1,2,7,9} Inhaled dose, expressed in percentage of the nominal dose (ND), corresponding to the nebulized doses deposited on the filter interposed between nebulizer and lung model (weighed before the nebulization and after drying for 24 h) multiplied by the relative mass of pentamidine. The residual volume was also quantified.

In the in vivo part, after ethical approval (2013/27JUI/375) and registration of the trial (NCT02277808), five non-smoker healthy male volunteers were recruited and signed a written informed consent. Each subject performed a spirometry according to the ATS/ERS guidelines.¹⁰ This was a randomized cross-over study based on CONSORT statement for clinical trials. Nebulizations of amikacin sulfate (Amukin®, Bristol-Myers Squibb) dissolved in 4 mL of normal saline (125 mg/mL) were made during 10 min with both devices. During nebulization, tidal volume (Vt; L), respiratory frequency (RF; min⁻¹) and minute ventilation (VE; Lmin⁻¹) were monitored by inductance plethysmography (Respiraces®, Ambulatory Monitoring Inc.). Participants were requested (1) to empty their bladder before nebulization, (2) to inhale spontaneously through the mouthpiece with a nose clip in a sitting position, (3) to collect their urine for 24 h following nebulization and (4) to observe a wash-out period of one week between both nebulizations. Then comparison was performed by sampling the daily urinary excretion of nebulized amikacin following the technique previously described by Dequin et al.¹¹ and analyzed by High Performance Liquid Chromatography. The total daily amount of amikacin excreted in the urine (Cu max) was calculated from cumulating amikacin amount measured at each micturition (Cu) and represents the lung dose. The elimination constant (K_e) was calculated from the fitted curve of the cumulated amount of amikacin excreted in the urine plotted versus the time. The equation is $Cu = Cu_{max}(1 - e^{-K_e t})$.

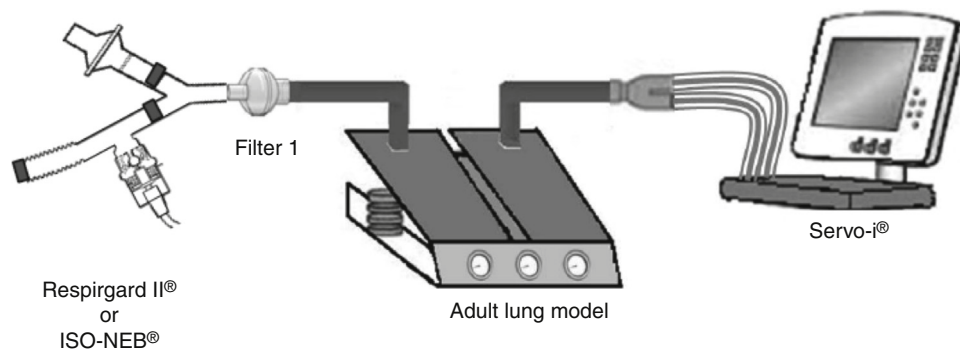


Fig. 1. Schematic diagram of experimental devices used in the in vitro study.

Inhaled dose was similar between devices, 28.7% (22.7; 33.5) vs 29.3% of ND (26.3; 33.1) for ISO-NEB[®] and for RespigardII[®] ($p=0.792$). Residual volume was 0.9313 g (0.9270; 0.9382) and 1.4087 g (1.3845; 1.4416) for ISO-NEB[®] and for RespigardII[®].

In vivo, all volunteers (23.5 ± 1.3 years) had spirometric values in the normal range. Cu_{max} was similar between devices with 3.5% (3.1; 3.6) and 3.6% of ND (2.2; 4.2), ($p=0.893$) for ISO-NEB[®] and RespigardII[®], respectively. Urine volume was 1.37 L (0.80; 1.72) and 1.30 L (0.75; 2.10) for ISO-NEB[®] and RespigardII[®], respectively ($p=0.686$). Elimination constant (K_e) of the drug following nebulization was similar for both devices (0.159 (0.078; 0.208) vs 0.130 (0.085; 0.162) for ISO-NEB[®] and for RespigardII[®], respectively ($p=0.225$)). There was no significant difference in RF, Vt and VE. Our results were in the range of previous studies even if the comparison is difficult because there were no data for ISO-NEB[®] and the previous studies on RespigardII[®] presented many differences in protocols and measurements techniques.^{6,9,12,13} We used the most frequent dosage reported in the previous studies and in the manufacturer's recommendations (300 mg pentamidine in 6 mL sterile water).^{1,2,7,9} The airflow rate (8 L min⁻¹) to produce aerosolized pentamidine was in the range of the rate described in previous studies about RespigardII[®] (about 6–10 L min⁻¹ for RespigardII[®]) or recommended by the manufacturer for ISO-NEB[®] (from 5 to 9 L min⁻¹). The total amount of drug reaching the lungs was similar and lower than 5% of ND for both devices. Our results are in line with those obtained with pentamidine in prior studies (2–6.74% of ND).^{4,6,9,12} Some methodological conditions need to be discussed. In vitro, we used the residual gravimetric technique even though it was not validated for pentamidine. However previous studies validated this process for different drugs.¹⁴ In vivo, we used amikacin sulfate because pentamidine has not been previously considered as a valid pharmacological marker of pulmonary deposition.^{11,15} For a methodological consideration, we recruited only male subjects because it is easier for a man to collect his urine without loss and to prevent potential fetal risk ototoxicity in pregnant female subjects. Finally, we did not measure the particle sizes for both nebulizers, but according to previous studies and manufacturer's data we can consider that they were similar (1–2 μm). It is important to notice that the two nebulizers are in the same price range. In conclusion, this in vitro and in vivo study demonstrated that ISO-NEB[®] and RespigardII[®] have similar properties in the conditions study. Further clinical studies are needed to confirm that ISO-NEB[®] is a valuable alternative to the reference nebulizer recommended by guidelines for pentamidine delivery. Altogether these data suggest that the performance of both devices is similar in the conditions of this in vitro and in vivo study.

References

- Pyrgos V, Shoham S, Roilides E, Walsh TJ. Pneumocystis pneumonia in children. *Paediatr Respir Rev*. 2009;10:192–8.
- Thomas CF Jr, Limper AH. Pneumocystis pneumonia. *N Engl J Med*. 2004;350:2487–98.
- Oudry M, Chaumazeau JP, Diot P, Dubus JC. [Use of pentamidine nebulization in children]. *Rev Malad Respir*. 2012;29:656–63.
- O'Doherty M, Thomas S, Page C, Bradbeer C, Nunan T, Bateman N. Pulmonary deposition of nebulised pentamidine isethionate: effect of nebuliser type, dose, and volume of fill. *Thorax*. 1990;45:460–4.
- Boe J, Dennis JH, O'Driscoll BR, Bauer TT, Carone M, Dautzenberg B, et al. European Respiratory Society Guidelines on the use of nebulizers. *Eur Respir J*. 2001;18:228–42.
- Ilowite JS, Baskin MI, Sheetz MS, Abd AG. Delivered dose and regional distribution of aerosolized pentamidine using different delivery systems. *Chest*. 1991;99:1139–44.
- Peron N, Le Guen P, Andrieu V, Bardot S, Ravilly S, Oudry M, et al. [Inhalation therapy: inhaled generics, inhaled antidotes, the future of anti-infectives and the indications of inhaled pentamidine. GAT aerosolstorming, Paris 2012]. *Rev Malad Respir*. 2013;30:832–42.
- Hess DR. Nebulizers: principles and performance. *Respir Care*. 2000;45:609–22.
- Kim CS, Garcia L, Wanner A. Actual pentamidine dose delivered by Respigard II nebulizer. *Eur Respir J*. 1995;8:2178–81.
- Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, et al. Standardisation of spirometry. *Eur Respir J*. 2005;26:319–38.
- Dequin PF, Faurisson F, Lemarie E, Delatour F, Marchand S, Valat C, et al. Urinary excretion reflects lung deposition of aminoglycoside aerosols in cystic fibrosis. *Eur Respir J*. 2001;18:316–22.
- Ferretti PP, Versari A, Gafa SI, Becquemin MH, Barchi E, Serafini D, et al. Pulmonary deposition of aerosolized pentamidine using a new nebuliser: efficiency measurements in vitro and in vivo. *Eur J Nuclear Med*. 1994;21:399–406.
- Smaldone GC, Fuhrer J, Steigbigel RT, McPeck M. Factors determining pulmonary deposition of aerosolized pentamidine in patients with human immunodeficiency virus infection. *Am Rev Respir Dis*. 1991;143:727–37.
- Vecellio None L, Grimbert D, Bordenave J, Benoit G, Furet Y, Fauroux B, et al. Residual gravimetric method to measure nebulizer output. *J Aerosol Med*. 2004;17:63–71.
- Reychler G, Leal T, Roeseler J, Thys F, Delvau N, Liistro G. Effect of continuous positive airway pressure combined to nebulization on lung deposition measured by urinary excretion of amikacin. *Respir Med*. 2007;101:2051–5.

Nicolas Audag^{a,b,*}, Giuseppe Liistro^{b,c}, Dimitri Van der Linden^d, Françoise Smets^e, Teresinha Leal^f, Gregory Reychler^{a,b,c}

^a Service de Médecine Physique et Réadaptation, Cliniques universitaires Saint-Luc, Brussels, Belgium

^b Institut de Recherche Expérimentale et Clinique (IREC), Pôle de Pneumologie, ORL & Dermatologie, Université Catholique de Louvain, Brussels, Belgium

^c Service de Pneumologie, Cliniques universitaires Saint-Luc, Brussels, Belgium

^d Pediatric Infectious Diseases, Cliniques universitaires Saint-Luc, Brussels, Belgium

^e Pediatric Gastroenterology and Hepatology Department, Cliniques universitaires Saint-Luc, Brussels, Belgium

^f Louvain Centre for Toxicology and Applied Pharmacology (LTAP), Institut de Recherche Expérimentale et Clinique (IREC), Brussels, Belgium

* Corresponding author.

E-mail address: nicolas.audag@uclouvain.be (N. Audag).

1579-2129/

© 2017 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

Mycobacterium xenopi and Squamous Cell Carcinoma of the Lung[☆]



Mycobacterium xenopi y carcinoma pulmonar de células escamosas

[☆] Please cite this article as: Martín Asenjo M, Martín Guerra JM, López Pedreira MR, Prieto de Paula JM. *Mycobacterium xenopi* y carcinoma pulmonar de células escamosas. *Arch Bronconeumol*. 2017;53:698–700.

Dear Editor,

Non-tuberculous mycobacteria (NTM) or atypical mycobacteria are aerobic bacteria of the genus *Mycobacterium*, the pathogenic potential of which has been known since the 1950s.¹ The AIDS pandemic, the progressive increase in immunosuppressive states, and the improvement of microbiological techniques have made isolation of these microorganisms more common nowadays.²

Mycobacterium xenopi (*M. xenopi*) is a NTM associated with water systems that is found primarily in North America, the south