

disease, and plays a significant role in the control of both gastrointestinal and pulmonary manifestations, as reported by Hayek.<sup>10</sup> Treatment duration has not been established and varies greatly: from 2 weeks in the shortest schedules, up to 3 months in the longest. In our case, treatment continued at full dose for 1 month, and was then tapered over the following 2 months until discontinuation.

The mechanism of how inflammatory bowel disease can lead to pulmonary manifestations is still unknown. In embryonic development, the formation of the gastrointestinal tract and respiratory system originate in the same part of the embryonic structure and have a similar epithelial structure, so this might explain why the lung may become involved in this entity. However, many other alternative mechanisms have been proposed, such as bacterial dysbiosis, environmental pollution, or even genetic factors.<sup>1</sup> For this reason, when performing a differential diagnosis in any patient with inflammatory bowel disease, the presence of manifestations of their underlying disease should be taken into account, especially in patients with cough and fever and no obvious infection. Although the incidence of these manifestations is still believed to be low, underdiagnosis seems likely, given the similarity of diagnostic test results and the fact that many patients who respond well to inhaled corticosteroids may be classified as asthmatics.<sup>6,7</sup> We must, therefore, take into account this diagnostic possibility when we encounter patients with these characteristics.

## References

1. Vutcovici M, Brassard P, Bitton A. Inflammatory bowel disease and airway diseases. *World J Gastroenterol*. 2016;22:7735–41.
2. Majewski S, Piotrowski W. Pulmonary manifestations of inflammatory bowel disease. *Arch Med Sci*. 2015;11:1179–82.
3. Larsen S, Bendtzen K, Nielsen OH. Extraintestinal manifestations of inflammatory bowel disease: Epidemiology, diagnosis, and management. *Ann Med*. 2010;42:97–114. <http://dx.doi.org/10.3109/07853890903559724>.

4. Asami T, Koyama S, Watanabe Y, Miwa C, Ushimaru S, Nakashima Y, et al. Tracheobronchitis in a Patient with Crohn's Disease. *Intern Med*. 2009;48:1475–8.
5. Yeung V, Govind AG, Arastu S, Henry CH. Tracheobronchitis in a patient with Crohn's disease. *ACG Case Rep J*. 2016;3:181–3.
6. Javia S, Agrawal A, Patell R, Jasdandwala S. Tracheobronchitis as an extraintestinal manifestation of ulcerative colitis. *BMJ Case Rep*. 2014;16 <https://doi.org/10.1136/bcr-2014-205328>, e2014205328.
7. Kar S, Thomas S. A case of tracheobronchitis in ulcerative colitis: a review of literature. *Clin Respir J*. 2009;31:51–4. <http://dx.doi.org/10.1111/j.1752-699X.2008.00053.x>.
8. Hamada S, Ito Y, Imai S, Oguma T, Niimi A, Mishima M. Effect of inhaled corticosteroid therapy on CT scan-estimated airway dimensions in a patient with chronic bronchitis related to ulcerative colitis. *Chest*. 2011;139:930–2. <http://dx.doi.org/10.1378/chest.10-1105>.
9. Kinebuchi S, Oohashi K, Takada T, Moriyoama H, Yoshizawa H, Kobayashi O, et al. Tracheo-bronchitis associated with Crohn's disease improved on inhaled corticotherapy. *Intern Med*. 2004;43:829–34.
10. Hayek A, Pfanner T, White H. Inflammatory bowel disease of the lung: The role of infliximab? *Respir Med Case Reports*. 2015;15:85–8. <http://dx.doi.org/10.1016/j.rmcr.2015.05.012>.

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## Diagnostic Performance of a Lateral Flow Assay for the Detection of Alpha-1-Antitrypsin Deficiency



### Rendimiento diagnóstico de un ensayo de flujo lateral para la detección de la deficiencia de alfa-1 antitripsina

Dear Editor:

Despite of the demonstrated benefits of an early diagnosis and treatment on disease progression,<sup>1</sup> today the diagnosis of alpha1-antitrypsin deficiency (AATD) remains a challenge in daily clinical practice. Different barriers have been identified during the past years including a low suspicion level mainly based on a misconception of the disease among clinicians, who only suspect the disease in a selected and infrequent group of patients.<sup>2</sup> As a consequence, the degree of alpha1-antitrypsin (AAT) serum determination is frequently insufficient.<sup>3,4</sup> Accordingly, different initiatives have been developed to improve diagnosis over the past years. Based on lateral-flow paper-based technologies,<sup>5</sup> the Alphakit Quickscreen (Grifols, Barcelona, Spain) has been marketed in Europe for the identification of the Z protein (PiZ) in serum. Therefore, the test allows the identification of the Z allele homozygous, heterozygous and carriers

The opportunities for the diagnosis of AATD of such a test are clear, since the on-site rapid and accurate identification of PiZ carriers may help identify AATD cases at an early stage with implications

for management and family screening. However, the diagnostic accuracy of this test has been scarcely studied. In the one real world evaluation of its performance available,<sup>6</sup> the evaluation of the test's ability to detect the PiZ protein showed a specificity of 97.8%, sensitivity of 73.8%, negative predictive value of 98.9%, and positive predictive value of 58.5%. Of note, after exploring the test-performance with different prevalence pre-test probability, the authors found lower negative predictive values in a population with a very high pre-test probability. Therefore, the authors concluded that the device could be used as an appropriate tool to exclude AATD in primary care and in the overall COPD population, except in patients with a high a priori-probability of AATD. Additionally, an added finding was that all false negatives ( $n = 11$ ) were heterozygote Pi\**MZ* samples, with a correct identification of ZZ and SZ genotypes. In this context, an unexplored population of special interest is that with a low a priori pre-test probability of severe AATD defined as those with AAT serum concentration > 50 mg/dl which are associated with increased risk for COPD<sup>7</sup>. The identification of the Z allele in this population may help identifying Z carriers and advance in the early identification of treatable cases.

We performed a prospective, single-center, observational, cross-sectional, real-world analysis on the performance of Alphakit Quickscreen. Following the Spanish recommendations,<sup>8</sup> all patients referred to our COPD-dedicated outpatient clinic from January 2016 to July 2018 for a diagnosis of COPD had a serum AAT determination by nephelometry and those between 50 and

**Table 1**  
Results of the Alphascreen test and the genotyping.

Genotype	Complete cohort (n = 16)		Alphascreen negative (n = 7)		Alphascreen positive (n = 9)	
	N	AAT <sup>a</sup>	N	AAT <sup>a</sup>	N	AAT <sup>a</sup>
MM	2	116.1 ± 7.6	1	110.7	1	120.0
MZ	4	83.4 ± 12.5	0	–	4	83.4 ± 12.5
SM	4	104.5 ± 9.8	4	104.5 ± 9.8	0	–
SS	1	87.0	1	87.0	0	–
SZ	5	79.3 ± 25.9	1	56.5	4	85.0 ± 26.0

<sup>a</sup> Alpha1 antitrypsin serum concentration expressed as mg/dL.

Data are expressed as absolute frequencies and mean ± standard deviation according to the variables' nature.

120 mg/dL, had an Alphascreen determination done. The test was performed complying with all the requirements of the manufacturer in the conservation and performance of the test. The study was completed by allele-specific genotyping. The diagnostic profile including the sensitivity, specificity, and the positive and negative predicted values for the identification of the Z allele was calculated together with the 95% confidence interval (95%CI). Data are expressed as absolute (relative) frequencies and mean (standard deviation) according to the variables' nature.

The sample was composed of 16 subjects: 11 (68.8%) males, 58.4 (15.3) years of age, 6 (37.5%) current smokers, tobacco history 52.7 (31.7) pack-year, body mass index 27.7 (6.8) kg/m<sup>2</sup>, of which 12 (75.0%) fulfilled COPD diagnostic criteria with a post-bronchodilator FEV<sub>1</sub> of 64.3 (18.0) %. Serum AAT concentrations varied from 56.5 to 120.0 mg/dL, with a mean value of 91.7 (20.7) mg/dL. The distribution of the Z alleles according to the Alphascreen result is presented in Table 1. The prevalence of the Z allele in this highly selected low-risk population of cases was 56.2% (95%CI 31.9–80.5). The diagnostic profile of the test resulted in a specificity of 85.71 (95%CI 68.5–100), sensitivity 88.9 (95%CI 73.5–100), negative predictive value 85.7 (95%CI 68.5–100), and positive predictive value 88.9 (95%CI 73.5–100). We detected one false negative (a SZ case with AAT serum concentration of 56.5 mg/dL) and one false positive (a MM case with AAT serum concentration of 120 mg/dL).

The Alphascreen represents an attractive diagnostic method since it explores the presence of the Z protein using a non-invasive method and is freely distributed in the country by Grifols (Grifols, Barcelona, Spain). Our data show the diagnostic profile of Alphascreen device in an a priori low probability of AATD cohort and additionally describing one case of a false negative in a SZ patient and a false positive in a MM case. The main strength of our approach is the exploration of a specific subgroup of patients in which the test may be useful. The limitation is the small sample size which is intrinsically associated with the population selection in a rare disease like AATD. Therefore, these findings should be confirmed in an independent cohort that includes these types of patients.

The evaluation of the diagnostic profile of new diagnostic techniques is a necessary step before wide clinical implementation. The one previous real-world available study evaluated 1019 samples in test-naïve COPD patients from 9 centers in Spain and 10 centers in Germany. Patients included were ≥30 years of age and with a confirmed diagnosis of COPD, with no exclusion criteria regarding the AAT serum levels. Our study continues the evaluation of the Alphascreen in a specific population which may be of interest for the identification of Z-allele carriers. Our area has a lower prevalence of Pi\*Z as compared to the rest of the country.<sup>9</sup> As a quick, easy, point-of-care test, this may have implications in the use of this test in future diagnostic algorithms. Although this device allows for rapid analysis at the patient's point of care, its actual position within a diagnostic algorithm as a screening tool needs to be assessed.

Another interesting finding is the identification of one case of SZ which was a false negative, as compared to the previous analysis in which all false negatives were MZ.<sup>6</sup> False negatives in a lateral-flow analysis may be related to preservation, technical performance or correct interpretation, rather than the specific genotype. Therefore, it is expected that false negatives may affect different Z-allele carriers irrespective of the final genotype, as the study of the test expands. Moreover, it would be interesting to confirm the genotype in all false negative and false positive cases by complete sequencing of coding exons of AAT gene.

In summary, the evaluation of the performance of Alphascreen in an a priori low risk of AATD population shows a diagnostic profile that allows it to be considered as a potential step in the diagnostic algorithm of AATD diagnosis. Additionally, this device represents a great opportunity to increase the awareness of AATD.

### Conflicts of interest

JLLC has received during the last 3 years personal fees or non-financial support from Grifols and CSL Behring. The rest of the authors declare no conflicts of interest.

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### References

- Chapman KR, Burdon JG, Piitulainen E, Sandhaus RA, Seersholm N, Stocks JM, et al. Intravenous augmentation treatment and lung density in severe alpha1 antitrypsin deficiency (RAPID): a randomised, double-blind, placebo-controlled trial. *Lancet*. 2015;386:360–8.
- Torres-Duran M, Lopez-Campos JL, Barrecheguren M, Miravittles M, Martinez-Delgado B, Castillo S, et al. Alpha-1 antitrypsin deficiency: outstanding questions and future directions. *Orphanet J Rare Dis*. 2018; 13:114.
- Soriano JB, Lucas SJ, Jones R, Miravittles M, Carter V, Small I, et al. Trends of testing for and diagnosis of alpha1-antitrypsin deficiency in the UK: more testing is needed. *Eur Respir J*. 2018;52.
- Calle Rubio M, Soriano JB, Lopez-Campos JL, Soler-Cataluna JJ, Alcazar Navarrete B, Rodriguez Gonzalez-Moro JM, et al. Testing for alpha-1 antitrypsin in COPD in outpatient respiratory clinics in Spain: a multilevel, cross-sectional analysis of the EPOCONSUL study. *PLOS ONE*. 2018;13:e0198777.
- Yetisen AK, Akram MS, Lowe CR. Paper-based microfluidic point-of-care diagnostic devices. *Lab Chip*. 2013;13:2210–51.
- Greulich T, Rodriguez-Frias F, Belmonte I, Klemmer A, Vogelmeier CF, Miravittles M. Real world evaluation of a novel lateral flow assay (AlphaKit(R) QuickScreen) for the detection of alpha-1-antitrypsin deficiency. *Respir Res*. 2018; 19:151.
- Molloy K, Hersh CP, Morris VB, Carroll TP, O'Connor CA, Lasky-Su JA, et al. Clarification of the risk of chronic obstructive pulmonary disease in alpha1-antitrypsin deficiency PiMZ heterozygotes. *Am J Respir Crit Care Med*. 2014;189: 419–27.
- Miravittles M, Soler-Cataluna JJ, Calle M, Molina J, Almagro P, Quintano JA, et al. Spanish Guidelines for Management of Chronic Obstructive Pulmonary Disease (GesEPOC) 2017. Pharmacological treatment of stable phase. *Arch Bronconeumol*. 2017;53:324–35.

9. Blanco I, Fernandez E. Alpha-1-antitrypsin Pi phenotypes S and Z in Spain: an analysis of the published surveys. *Respir Med.* 2001;95:109–14.

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## Cystic Metastasis of a Giant Cell Tumor Causing Recurrent Spontaneous Pneumothorax



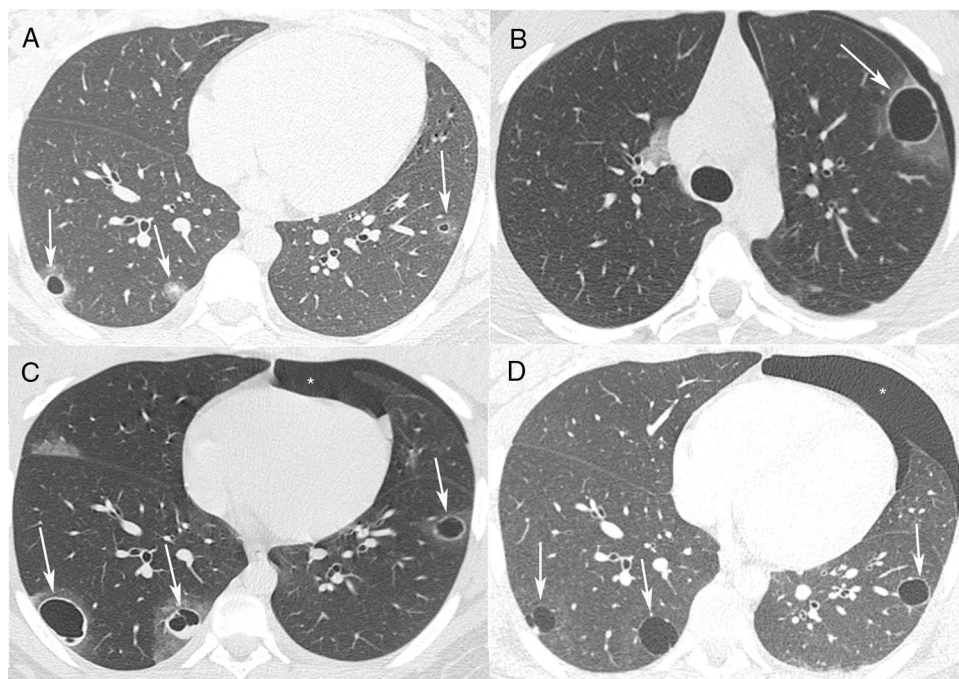
### Metástasis quística de un tumor de células gigantes causante de neumotórax espontáneo recurrente

Dear Editor:

A 17-year-old male was admitted to the Emergency Department with cough and episodes of hemoptysis. The patient had a history of a giant cell tumor (GCT) in the left tibia, resected 6 months previously. Chest computed tomography (CT) revealed pulmonary nodules, some of which were cavitated (Fig. 1A). Laboratory test findings were unremarkable. The patient's sputum was negative for acid-fast bacilli. He was referred for fiberoptic bronchoscopy with bronchoalveolar lavage. The bronchoalveolar lavage fluid contained a small amount of blood, and was negative for neoplastic cells. Cultures were negative for fungus and bacteria. Video-assisted thoracoscopy was performed, and the biopsy findings from one of the nodules were compatible with GCT metastasis. The patient started a new chemotherapy cycle. Four months later, he had an episode of chest pain associated with hemoptysis. A new CT examination showed a left pneumothorax, and cavitated

thick-walled nodules with ground-glass halos (Fig. 1B and C). The pneumothorax was drained. The patient evolved well, with pulmonary re-expansion. Eight months later, he had a new episode of chest pain and dyspnea. CT showed a spontaneous left pneumothorax, and evolution of the cavitated nodules into thin-walled cysts (Fig. 1D). In this phase, the patient presented metastasis to intraabdominal lymph nodes in addition to the pulmonary metastases. He underwent new chest drainage and pleuroscopy with bilateral pleurodesis through the intrapleural instillation of talc. During pleuroscopy, pleural metastases were detected. The patient underwent a chemotherapy regimen with six cycles of doxorubicin and cisplatin, which resulted in regression of some of the lung lesions. He remains in outpatient follow-up with no new complication 1 year after the last pneumothorax.

GCT in the bone is a primary intramedullary tumor; it is generally benign, but can be locally aggressive and even metastatic. Malignant transformation and distant metastasis are extremely uncommon. Malignant transformation may occur as a result of dedifferentiation of the primary tumor or secondary to previous radiation therapy. Metastasis of GCTs most commonly arises in the lungs. Pulmonary metastases are more likely to appear in patients with recurrent GCTs, and often have an indolent course; they are rarely fatal.<sup>1,2</sup> Cavitation of metastases is extremely rare.



**Fig. 1.** (A) Axial chest CT with pulmonary window settings shows bilateral small pulmonary nodules, two of which are cavitated (arrows). (B, C) CT performed 4 months later demonstrates a left spontaneous pneumothorax (asterisk) and growth of the nodules, which now present with relatively thick walls and ground-glass halos (arrows). (D) CT performed 1 year after A shows a new left pneumothorax (asterisk) and evolution of the cavitated nodules into thin-walled cystic lesions (arrows).