



Original Article

Increased Urinary Erythropoietin Excretion in Severe Sleep Apnea–Hypoapnea Syndrome: The Effect of CPAP[☆]



Miquel Félez,^{a,b,*} Nuria Grau,^{a,b} Antonia Ruiz,^a Encarna Guardiola,^a Carles Sanjuas,^{a,b} Cristina Estirado,^b Maribel Navarro-Muñoz,^c Antoni Pascual,^{c,d} Mauricio Orozco-Levi,^e Joaquim Gea^b

^a Unidad Multidisciplinar de Trastornos del Sueño, Hospital del Mar, Parc de Salut Mar, UAB-UPF, Barcelona, Spain

^b Servicio de Neumología, Hospital del Mar, Parc de Salut Mar, IMIM, UAB-UPF, CIBERES, ISC III, Barcelona, Spain

^c Group of Integrative Pharmacology and Systems Neuroscience, Neurosciences Programme, IMIM (Hospital del Mar Medical Research Institute), Parc de Recerca Biomèdica de Barcelona, Barcelona, Spain

^d Department of Experimental and Health Sciences, Universitat Pompeu Fabra, Parc de Recerca Biomèdica de Barcelona, Barcelona, Spain

^e Servicio de Neumología, Fundación Cardiovascular de Colombia, Hospital Internacional de Colombia, Bucaramanga, Colombia

ARTICLE INFO

Article history:

Received 15 May 2017

Accepted 21 October 2017

Available online 31 March 2018

Keywords:

Erythropoietin in urine

Sleep apnea–hypopnea syndrome

Continuous positive airway pressure

ABSTRACT

Introduction: Tissue hypoxia stimulates the production of erythropoietin (EPO), the main effect of which is, in turn, to stimulate erythropoiesis. Sleep apnea–hypopnea syndrome (SAHS) is an entity characterized by repeated episodes of hypoxemia during sleep.

Objective: To analyze whether hypoxemia stimulated increased urinary excretion of EPO, and if so, to evaluate if treatment with continuous positive airway pressure (CPAP) can inhibit this phenomenon.

Methods: We studied 25 subjects with suspected SAHS who underwent a polysomnography study (PSG). EPO levels in first morning urine (uEPO) and blood creatinine and hemoglobin were determined in all patients. Patients with severe SAHS repeated the same determinations after CPAP treatment.

Results: Twelve subjects were diagnosed with severe SAHS (mean±SD, AHI 53.1±22.7). Creatinine and hemoglobin levels were normal in all subjects. uEPO was 4 times higher in the SAHS group than in the control group (1.32±0.83 vs 0.32±0.35 UI/l, $P<.002$). CPAP treatment reduced uEPO to 0.61±0.9 UI/l ($P<.02$), levels close to those observed in healthy subjects. No dose–response relationship was observed between severity of PSG changes and uEPO values.

Conclusions: Patients with severe SAHS show increased uEPO excretion, but this normalizes after treatment with CPAP.

© 2017 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

El síndrome de apneas-hipoapneas del sueño (SAHS) grave incrementa la excreción urinaria de eritropoyetina. Efecto del tratamiento con CPAP

RESUMEN

Introducción: La hipoxia tisular estimula la producción de eritropoyetina (EPO) que tiene como principal función estimular la eritropoyesis. El SAHS es una entidad caracterizada por la presencia de episodios repetidos de hipoxemia durante el sueño.

Objetivo: Analizar si dicha hipoxemia es un estímulo suficiente para incrementar la excreción urinaria de EPO. Si la respuesta fuera positiva, valorar si el tratamiento con presión continua positiva de la vía aérea (CPAP) la inhibiría.

Métodos: Se han estudiado 25 sujetos con sospecha de SAHS, a los que se les realizó un estudio polisomnográfico. En todos ellos se determinaron los niveles de EPO en la primera orina de la mañana (uEPO), así como los niveles de creatinina y hemoglobina en sangre. En los pacientes con SAHS grave se repitieron las mismas determinaciones tras el tratamiento con CPAP.

Palabras clave:

Eritropoyetina en orina

Síndrome de apneas-hipoapneas del sueño

Presión positiva continua de la vía aérea

[☆] Please cite this article as: Félez M, Grau N, Ruiz A, Guardiola E, Sanjuas C, Estirado C, et al. El síndrome de apneas-hipoapneas del sueño (SAHS) grave incrementa la excreción urinaria de eritropoyetina. Efecto del tratamiento con CPAP. Arch Bronconeumol. 2018;54:255–259.

* Corresponding author.

E-mail address: 91435@hospitaldelmar.cat (M. Félez).

Resultados: Doce sujetos fueron diagnosticados de SAHS grave (media \pm SD, IAH de $53,1 \pm 22,7$). La creatinina y la hemoglobina fueron normales en todos los sujetos. La uEPO fue cuatro veces superior en el grupo SAHS respecto a los controles ($1,32 \pm 0,83$ vs $0,32 \pm 0,35$ IU/l, $p < 0,002$). El tratamiento con CPAP descendió la uEPO hasta $0,61 \pm 0,49$ IU/l ($p < 0,02$), acercándose al valor de los sujetos sanos. No se observó una relación dosis-respuesta entre la gravedad de las alteraciones de la PSG y los valores de uEPO.

Conclusiones: Los pacientes con SAHS grave muestran un incremento en su excreción de uEPO, que se normaliza tras el tratamiento con CPAP.

© 2017 SEPAR. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Introduction

Sleep apnea–hypopnea syndrome (SAHS) is a very common disease among adults.¹ It is characterized by multiple and repeated episodes of upper airway obstruction that cause sleep disruption, microarousals, hypoxemia, and hypercapnia of variable duration and intensity during sleep.² Its impact on sleep quality, health-related quality of life, daytime sleepiness, and cardiovascular morbidity and mortality have been widely studied.² However, few attempts have been made to understand the impact of intermittent hypoxemia associated with SAHS on erythropoiesis, and results in general have been divergent.

Human erythropoietin (EPO) is a 30.4 kDa glycoprotein hormone composed of a single 165 amino acid residues chain to which four glycans are attached. The kidneys are the main source of EPO, but its messenger RNA (mRNA) has also been detected in the liver, spleen, bone marrow, lung and brain. Circulating levels of EPO increase exponentially with the decline of partial pressure and the levels of oxygen in tissue (PtiO₂ and CtiO₂, respectively), which in turn depend on local blood flow, hemoglobin (Hb) concentration, partial pressure of oxygen in arterial blood (PaO₂), and Hb–O₂ affinity. However, PtiO₂ and CtiO₂ in the renal cortex are less affected by changes in blood flow than other organs; the renal cortex then is very appropriate for regulating EPO production.

In the bone marrow, EPO promotes the survival, proliferation and differentiation of nucleated red blood cells, particularly colony-forming units. Following a rise in plasma EPO it takes around 4 days before reticulocytosis becomes apparent. The hormone is metabolized in the liver and eliminated via the urine.³

Several studies have attempted to show that the intermittent episodes of hypoxia typical of SAHS stimulate EPO production, but results have been inconsistent. Cahan et al. showed that EPO levels rise in plasma during the day⁴ in patients with SAHS and hypoxemia. They subsequently found that treatment with CPAP could normalize this effect.⁵ Other authors have reported similar results, but only in severe SAHS, as opposed to mild–moderate disease,⁶ while others reported minor increases in EPO.⁷ These results, however, could not be reproduced by authors such as Mckeon et al.,⁸ Pokala et al.,⁹ Ryan et al.,¹⁰ and Ciftici et al.¹¹

Recently, Zhang et al.¹² attempted to clarify these partial and contradictory results with a meta-analysis of 9 studies and 407 patients in whom plasma EPO had been evaluated. They concluded that plasma EPO is significantly higher in patients with SAHS than in normal individuals. They also determined that among the SAHS group, plasma EPO is higher in patients with cardiovascular complications, and, surprisingly, in patients with a body mass index <30 .

Given the confusion surrounding the effect of SAHS on plasma EPO levels, we hypothesized that EPO levels in urine (uEPO) may be a more sensitive marker than EPO in plasma, particularly if the analysis is performed on first morning urine, in which at least part of the night's excretion is concentrated. This hypothesis is based on three considerations: (1) EPO is excreted in urine,³ (2) peak

production occurs in the early hours of the morning,⁴ and (3) the hypoxic stimulus of SAHS occurs during the night.

The main objective of this study was to determine if SAHS is a sufficient stimulus to increase uEPO concentrations in first morning urine. Our secondary objective, provided the first objective was met, was to determine if eliminating the hypoxic stimulus of SAHS with CPAP therapy could reduce the morning concentration of uEPO.

Materials and Methods

Patients referred to the Multidisciplinary Sleep Disorder Unit of our hospital with suspected SAHS were recruited consecutively. The study was approved by the local Clinical Research Ethics Committee, and all subjects signed informed consent. Clinical, demographic and anthropometric measurements were obtained at the time of the polysomnography (PSG) study, which was carried out in all study participants. Patients with low oxygen saturation at rest (SatO₂ $\leq 92\%$) due to any underlying pathology, SAHS patients who were not candidates for CPAP, and patients with kidney or liver diseases were excluded from the study.

Design

The study involved an initial observational phase, followed by a second interventional phase. In the first phase, participants were divided into two groups depending on PSG results: controls ($n=13$) and patients with severe SAHS ($n=12$). In accordance with our secondary objective, all individuals with severe SAHS were candidates for CPAP. After 3–6 months of treatment, they were re-evaluated for the second study phase.

Sleep Study

SAHS was evaluated by medical staff specializing in sleep disorders, using the Epworth sleepiness test¹³ and results of a PSG. The PSG included electroencephalogram, electro-oculogram, electromyogram, electrocardiogram, chest and abdominal movements, oronasal flow, pulse oximetry, leg movements, and body position (eXea Series 5, Bitmed, Zaragoza, Spain).

Mean obstructive apneas and hypopneas per hour were defined according to the Apnea–Hypopnea Index (AHI), and mean desaturations (reduction $\geq 3\%$) per hour were defined according to the desaturation index (DI). The percentage of time with SatO₂ below 90% was defined using CT_{90%}. SAHS diagnosis was established in patients with AHI >10 , and classified as severe in those with AHI >30 . The SAHS patient group received auto-titrating CPAP (Autoset S9 with ResScan software, ResMed, San Diego, USA) to determine optimal pressure. Residual AHI was determined during auto-titration.

Determination of Urinary Erythropoietin

uEPO was quantified using a commercially available EPO ELISA kit (STEMCELL Technologies, Vancouver, Canada). The kit, initially

developed for serum samples, was adapted according to the manufacturer's instructions for use with urine samples. Fifteen ml samples were concentrated 10-fold before quantification by ultrafiltration (molecular weight cut-off [MWCO] 30 kDa). The calibration curve was also appropriately diluted in phosphate buffered saline (PBS) and processed using the same ultrafiltration procedure.

Ultrafiltration was performed using the Amicon Ultra-15 (Merck Millipore, Darmstadt, Germany), with a 30 kDa MWCO. The sample was activated with 15 ml Milli-Q water and centrifuged at $4000\times g$ for 1–2 min at 20°C . Tris–HCl 3.75 M at pH 7.4 (1.5 ml) and Complete Protease Inhibitor Cocktail (300 μl) (Sigma–Aldrich, St. Louis, USA) were added to all samples. These were then gently shaken and subjected to ultrasound for 5 min to facilitate passage through the filtration device, and centrifuged at $4000\times g$ for 15 min at 20°C . The supernatant was filtered under vacuum through a $0.22\ \mu\text{m}$ Steriflip device (Merck Millipore, Darmstadt, Germany).

The filtrate from each sample was transferred to the above-mentioned activated device. The resulting filtrate was washed twice with 15 ml of Tris–HCl 50 mM buffer at pH 7.4 supplemented with 300 μl Complete™ and centrifuged at $4000\times g$ for 25 min at 20°C until approximately 200 μl were obtained. This was then transferred to a new Eppendorf tube and stored at -20°C until the time of EPO analysis.

Following the instructions of the manufacturer of the ELISA kit, samples were diluted 10-fold with the supplied buffer B before they were placed on the anti-EPO microwell plate. Assay samples that showed out-of-range absorbance values underwent an additional dilution with buffer B, and were reanalyzed along with the reference samples. Intra- and inter-assay coefficients of variation were $<15\%$. Data were analyzed automatically using MyAssays software (MyAssays Ltd., Brighton, East Sussex, United Kingdom).

Statistical Analysis

Values are expressed as mean \pm standard deviation. Clinical differences between the 3 groups were analyzed using the Student's *t*-test, Mann–Whitney *U* test, Chi-square test, or Fisher's exact test, as appropriate. Correlations between AHI, DI, $\text{CT}_{90\%}$, mean SatO_2 , minimum SatO_2 , and uEPO levels were measured by the Spearman correlation test. All statistical tests were considered significant at a *P* value of $<.05$. We calculated the sample size for a universe size of 1000 patients, a confidence level of 90%, 10% accuracy, and a proportion of 50%. This gave a total of 60 participants, divided into 2 groups.

Data were analyzed using SPSS version 22 (IBM Corp., Armonk, USA).

Results

Population

A total of 31 subjects were initially considered for inclusion. Of these, 25 were included in the study. The remaining 6 were excluded due to refusal to participate, low baseline SatO_2 , or high serum creatinine. Clinical characteristics of the study population are shown in Table 1. Of the 25 patients included, 12 were diagnosed with SAHS, with a mean AHI of 56.1 ± 22.7 , DI 53.6 ± 24.3 , $\text{CT}_{90\%}$ $27.3\pm 22.7\%$, mean SatO_2 $90.6\pm 3.9\%$ and minimum SatO_2 $67.2\pm 12.4\%$. As expected, these variables were higher in SAHS patients than in the controls. No significant differences were observed in age, body mass index, serum creatinine, and hemoglobin levels between the two groups. No statistically significant differences in sleepiness according to the Epworth scale

Table 1
Study Population Clinical Data and Polysomnography Results.

	Control (n=13)	SAHS Patients (n=12)	SAHS Post-CPAP (n=12)
Age (years)	50.6 \pm 15.8	55.8 \pm 13.9	–
Sex (σ / φ)	6/7	9/3	–
BMI (kg/m ²)	29 \pm 5	32.9 \pm 9	32.9 \pm 9
Creatinine (mg/dl)	0.73 \pm 0.20	0.84 \pm 0.2	0.84 \pm 0.2
Hb (g/l)	13.7 \pm 1.3	14.4 \pm 1.3	14.2 \pm 0.8
Epworth scale	9.9 \pm 5	10.8 \pm 4.5	7.5 \pm 2 ^{***}
AHI	5.8 \pm 2.5	56.1 \pm 22.7 ^{**}	7.2 \pm 4.7 ^{****}
$\text{CT}_{90\%}$	0.3 \pm 0.4	27.3 \pm 22.7 [*]	–
DI	5.5 \pm 3.7	55.6 \pm 24.73 [*]	–
Mean SatO_2	95.6 \pm 1.4%	90.6 \pm 3.9 ^{**}	–
Minimum SatO_2	81.0 \pm 12.6%	67.2 \pm 12.5 ^{**}	–
CPAP pressure (cmH ₂ O)	–	–	10.8 \pm 1.1

Values are expressed as mean \pm standard deviation.

$\text{CT}_{90\%}$: percentage of time with oxygen saturation $<90\%$; Hb: serum hemoglobin; AHI: sleep apnea-hypopnea index per hour; DI: index of desaturations (3%) per hour; BMI, body mass index; mean SatO_2 : mean value of oxygen saturation during the night; minimum SatO_2 : minimum oxygen saturation during the night.

^{*} $P<.05$ comparing controls with SAHS patients.

^{**} $P<.01$ comparing controls with SAHS patients.

^{***} $P>.05$ comparing patients SAHS before and after CPAP.

^{****} $P<.01$ comparing SAHS patients before and after CPAP.

were observed between the control group (9.9 \pm 5) and the SAHS group (10.8 \pm 4.5). However, this variable declined significantly in the SAHS group after treatment with CPAP (7.5 \pm 2) (Table 1).

Mean CPAP pressure administered in the treatment of SAHS patients was 10.8 ± 1.1 cmH₂O, and residual AHI after 3–6 months of treatment was 7.2 ± 4.7 cmH₂O.

Two patients in the control group had associated cardiovascular alterations, involving supraventricular arrhythmias in both cases. In the SAHS group, 3 patients were hypertensive and 1 had moderate chronic obstructive pulmonary disease. None of them were active smokers.

UEPO Levels

UEPO levels were higher in the SAHS group than in the control group: 1.32 ± 0.83 versus 0.32 ± 0.35 IU/l, respectively ($P<.002$). CPAP treatment reduced uEPO levels to 0.61 ± 0.49 IU/l ($P<.02$), a value not statistically different from the baseline value in the control group ($P>.1$) (Fig. 1).

No significant correlation was observed between the PSG variables that define disease severity (AHI, DI, $\text{CT}_{90\%}$, mean SatO_2 , and minimum SatO_2) and uEPO levels.

Discussion

This study demonstrates that SAHS patients have higher levels of uEPO in first morning urine compared with control subjects. Moreover, CPAP administered to treat SAHS decreases uEPO to levels similar to those of healthy subjects.

These results are in line with those of Cahan et al.⁴ and Winnicki et al.,⁶ who reported elevated plasma levels of EPO in patients with severe SAHS. Cahan et al.⁵ subsequently showed that treatment with CPAP could normalize these increased concentrations. The most significant contribution of this study is that this behavior, already established in plasma,¹² is also observed in urine, a specimen that is easier and less invasive to obtain.

In the study population, increased uEPO levels did not lead to a significant increase in the concentration of hemoglobin in blood, although mean values in the SAHS group were slightly higher than in controls (Table 1). This result could be expected, since secondary

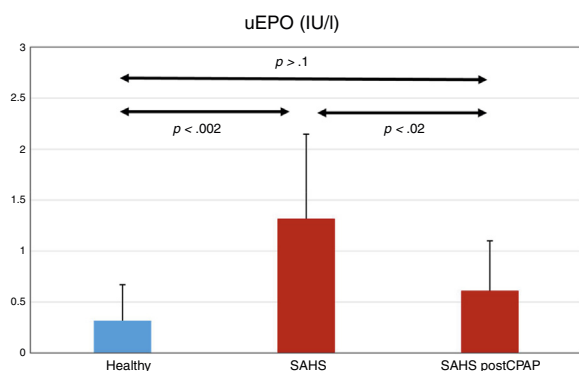


Fig. 1. EPO levels in urine (uEPO) in the group of healthy subjects ($n=13$), and in the group of SAHS subjects ($n=12$) before and after CPAP.

polycythemia is a rare finding in SAHS, observed in approximately 1% of cases.¹⁴ Very minor but significant differences with healthy subjects have only been achieved in large patient series, such as that published by Hoffstein et al.¹⁵ (624 patients).

The effect of CPAP treatment in reducing uEPO excretion is consistent with that observed with CPAP in polycythemia,¹⁴ which also normalizes. Recently, Song et al.,¹⁶ in an article pending publication, showed that CPAP corrects intermittent hypoxia caused by SAHS, inhibits EPO production, and also triggers neocytolysis. This is the main mechanism responsible for the rapid normalization of the number of red blood cells after the stimulus of intermittent hypoxia is eliminated.

One rather surprising finding in our study is that no correlation was found between the variables commonly used to quantify SAHS severity (AHI, DI, CT_{90%}, mean SatO₂, and minimum SatO₂) and uEPO concentrations in the SAHS group. This may be because these variables are not a good indicator of PtiO₂ in the renal cortex, which is the stimulus required to increase EPO production. SAHS is widely known to be associated with advanced age and hypertension, which may cause endothelial dysfunction¹⁰ and may, over time, affect circulation, particularly in the brain.¹⁷ These changes in local circulation may explain why systemic variables, such as AHI, DI, CT_{90%}, and mean and minimum SatO₂, are not a good reflection of PtiO₂ changes in the renal cortex.

However, chronic structural changes in local circulation are not a prerequisite for PtiO₂ to behave differently in different tissues in the presence of apneas. Almendros et al. showed in a rat model of obstructive apnea that PtiO₂ in the brain increases with apneas,¹⁸ but this phenomenon was not observed in muscle tissue or visceral fat.¹⁹ These authors found that this distinct response in the brain is dependent on hypercapnia that occurs during apneas, and is not seen in a model of intermittent hypoxia without hypercapnia.¹⁹ All these factors – the structural status of local circulation, presence or absence of hypercapnia, and degree of response by local circulation to hypercapnia – appear to be important for determining PtiO₂ in the renal cortex that cannot be measured simply with AHI, DI, CT_{90%}, mean SatO₂, and minimum SatO₂, and may explain the lack of correlation with uEPO. It is not unreasonable to speculate that other variables such as transcutaneous PO₂ or near-infrared spectroscopy²⁰ could provide more accurate data on real PtiO₂ in patients of this kind.²¹

Aside from this, we should point out that our study sample size was calculated to demonstrate our 2 main objectives, and proved sufficient for this purpose, but it is insufficient for identifying correlations between PSG variables and uEPO. If this objective is to be explored, a larger sample and different techniques will be necessary.

It should be noted that the SAHS patient group had severe disease, and this characteristic probably helped to highlight differences in uEPO compared to the control subjects. However, our data cannot be used to predict if the same would occur in patients with mild-to-moderate SAHS. A specific study to quantify uEPO in this population is necessary.

In short, this study shows that uEPO is clearly elevated in a group of patients with severe SAHS compared to the control group. The determination of this hormone in first morning urine is proposed as a simple approach for future studies. Moreover, we were able to reduce these elevated levels to normal with the use of CPAP. Further studies in larger populations are probably required, and other physiological and biological variables should be included for a better characterization of the mechanisms and clinical implications of these findings.

Conflict of Interests

The authors state that they have no conflict of interests.

References

- Durán J, Esnaola S, Rubio R, Iztueta A. Obstructive sleep apnea–hypopnea and related clinical features in a population-based sample of subjects aged 30 to 70 Yr. *Am J Respir Crit Care Med.* 2001;163:685–9.
- Durán-Cantolla J, Puertas-Cuesta FJ, Pin-Arboledas G, Santa María-Cano J, el Grupo Español de Sueño (GES). Documento de consenso nacional sobre el síndrome de apneas-hipopneas del sueño. *Arch Bronconeumol.* 2005;41:1–110.
- Jelkmann W. Physiology and pharmacology of erythropoietin. *Tranfus Med Hemother.* 2013;40:302–9.
- Cahan C, Decker MJ, Arnold JL, Washington LH, Veldhuis JD, Goldwasser E, et al. Diurnal variations in serum erythropoietin levels in healthy subjects and sleep apnea patients. *J Appl Physiol.* 1992;2112–7.
- Cahan C, Decker MJ, Arnold JL, Goldwasser E, Strohl P. Erythropoietin levels with treatment of obstructive sleep apnea. *J Appl Physiol.* 1995;76:1278–85.
- Winnicki P, Shamsuzzaman A, Lan Franchi P, Accurso V, Olson E, Davison D, et al. Erythropoietin and obstructive sleep apnea. *Am J Hypertens.* 2004;17:783–6.
- Imagawa S, Yamaguchi Y, Higuchi M, Neichi T, Hasegawa Y, Mukai HY, et al. Levels of vascular endothelial growth factor are elevated in patients with obstructive sleep apnea–hypopnea syndrome. *Blood.* 2001;98:1255–7.
- McKeon JM, Saunders NA, Murree-Allen K, Olson LG, Gyulay S, Dickeson J, et al. Urinary uric acid:creatinine ratio, serum erythropoietin, and blood 2,3-diphosphoglycerate in patients with obstructive sleep apnea. *Am Rev Respir Dis.* 1990;142:8–13.
- Pokala P, Llanera M, Sherwood J, Scharf S, Steinberg H. Erythropoietin response in subjects with obstructive sleep apnea. *Am J Respir Crit Care Med.* 1995;151:1862–5.
- Ryan S, Taylor CT, McNicholas WT. Selective activation of inflammatory pathways by intermittent hypoxia in obstructive sleep apnea syndrome. *Circulation.* 2005;112:2660–7.
- Ciftici TU, Jokturk O, Demirtas S, Gülbahar O, Bukan N. Consequences of hypoxia-reoxygenation phenomena in patients with obstructive sleep apnea syndrome. *Ann Saudi Med.* 2011;31:14–8.
- Zhang XB, Zeng YL, Zeng HQ, Zhang HP. Erythropoietin levels in patients with sleep apnea: a meta-analysis. *Eur Arch Otorhinolaryngol.* 2017;274:2505–12.
- Chiner E, Arriero JM, Signes-Costa J, Marco J, Fuentes I. Validación de la versión española del test de somnolencia Epworth en pacientes con síndrome de apnea de sueño. *Arch Bronconeumol.* 1999;35:422–7.
- Gangaraju R, Sundar KM, Song J, Prchal JT. Polycythemia is rarely caused by obstructive sleep apnea. *Blood.* 2016;128:2444.
- Hoffstein V, Herridge M, Mateika S, Redline S, Strohl KP. Hematocrit levels in sleep apnea. *Chest.* 1994;106:787–91.
- Song J, Sundar K, Gangaraju R, Prchal JT. Regulation of erythropoiesis after normoxic return from chronic sustained and intermittent hypoxia. *J Appl Physiol.* (1985). 2017. jap.00119.2017.
- Arzt M, Yount T, Finn L, Skatrud JB, Bradley TD. Association of sleep-disordered breathing and the occurrence of stroke. *Am J Respir Crit Care Med.* 2005;172:1447–51.
- Almendros I, Montserrat JM, Torres M, Gonzalez C, Navajas D, Farre R. Changes in oxygen partial pressure of brain tissue in an animal model of obstructive apnea. *Respir Res.* 2010;11:3.
- Almendros I, Farré R, Planas AM, Torres M, Bonsignore MR, Navajas D, et al. Tissue oxygenation in brain, muscle, and fat in a rat model of sleep apnea: differential

- effect of obstructive apneas and intermittent hypoxia. *Sleep*. 2011;34:1127–33.
20. Scheeren TWL, Schober P, Schwarte LA. Monitoring tissue oxygenation by near infrared spectroscopy (NIRS): background and current applications. *J Clin Monit Comput*. 2012;26:279–87.
21. Marín-Ceballos AJ, Murillo-Cabezas F, Domínguez-Roldan JM, Leal-Noval SR, Rincón-Ferrari MD, Muñoz-Sánchez MA. Monitorización de la presión tisular de oxígeno (PtiO₂) en la hipoxia cerebral: aproximación diagnóstica y terapéutica. *Med Intensiva*. 2008;32:81–90.