



Review

Diaphragm Dysfunction in Mechanically Ventilated Patients[☆]



Irene Dot,^{a,b} Purificación Pérez-Teran,^{a,b} Manuel-Andrés Samper,^{a,b} Joan-Ramon Masclans^{a,b,c,d,*}

^a Servicio de Medicina Intensiva, Hospital del Mar-Parc de Salut Mar de Barcelona, Barcelona, Spain

^b Institut Hospital del Mar d'Investigacions Mèdiques (IMIM)-GREPAC, Barcelona, Spain

^c Universitat Pompeu Fabra, Barcelona, Spain

^d CBERES, Spain

ARTICLE INFO

Article history:

Received 17 December 2015

Accepted 17 July 2016

Available online 12 January 2017

Keywords:

Diaphragm

Diaphragmatic dysfunction

Mechanical ventilation

Ultrasound

ABSTRACT

Muscle involvement is found in most critical patients admitted to the intensive care unit (ICU). Diaphragmatic muscle alteration, initially included in this category, has been differentiated in recent years, and a specific type of muscular dysfunction has been shown to occur in patients undergoing mechanical ventilation. We found this muscle dysfunction to appear in this subgroup of patients shortly after the start of mechanical ventilation, observing it to be mainly associated with certain control modes, and also with sepsis and/or multi-organ failure. Although the specific etiology of process is unknown, the muscle presents oxidative stress and mitochondrial changes. These cause changes in protein turnover, resulting in atrophy and impaired contractility, and leading to impaired functionality. The term 'ventilator-induced diaphragm dysfunction' was first coined by Vassilakopoulos et al. in 2004, and this phenomenon, along with injury cause by over-distention of the lung and barotrauma, represents a challenge in the daily life of ventilated patients.

Diaphragmatic dysfunction affects prognosis by delaying extubation, prolonging hospital stay, and impairing the quality of life of these patients in the years following hospital discharge. Ultrasound, a non-invasive technique that is readily available in most ICUs, could be used to diagnose this condition promptly, thus preventing delays in starting rehabilitation and positively influencing prognosis in these patients.

© 2016 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

Disfunción diafragmática: una realidad en el paciente ventilado mecánicamente

RESUMEN

La afectación muscular del paciente crítico está presente en la mayoría de pacientes que ingresan en el servicio de medicina intensiva (SMI). La alteración, en particular, del músculo diafragmático, inicialmente englobada en esta categoría, se ha diferenciado en los últimos años y se ha demostrado la existencia de una disfunción muscular propia de los pacientes sometidos a ventilación mecánica. En este subgrupo de pacientes encontramos una disfunción muscular que aparece de manera precoz después del inicio de la ventilación mecánica y que se relaciona principalmente con el uso de modalidades control, la presencia de sepsis y/o de fracaso multiorgánico. Aunque se desconoce la etiología concreta que desencadena el proceso, el músculo presenta procesos de estrés oxidativo y alteración mitocondrial que provocan un desequilibrio en la síntesis proteica, con el resultado de atrofia y alteración de la contractilidad y, como consecuencia, una menor funcionalidad. No fue, de hecho, hasta 2004 cuando Vassilakopoulos et al.

Palabras clave:

Diáfragma

Disfunción diafragmática

Ventilación mecánica

Ecografía

☆ Please cite this article as: Dot I, Pérez-Teran P, Samper M-A, Masclans J-R. Disfunción diafragmática: una realidad en el paciente ventilado mecánicamente. Arch Bronconeumol. 2017;53:150–156.

* Corresponding author.

E-mail address: jrmascians@parcdesalutmar.cat (J.-R. Masclans).

describieron el término «disfunción diafragmática asociada a ventilación mecánica», que, junto a la lesión por sobredistensión pulmonar y por barotrauma, representan un reto en el día a día de los pacientes ventilados.

La disfunción diafragmática tiene influencia en el pronóstico, retardando la extubación, aumentando la estancia hospitalaria y afectando la calidad de vida de estos pacientes en los años siguientes al alta hospitalaria. La ecografía, como técnica no invasiva y accesible en la mayoría de unidades, podría ser de utilidad en el diagnóstico precoz para iniciar, de forma avanzada, la rehabilitación e influir positivamente en el pronóstico de estos enfermos.

© 2016 SEPAR. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Introduction

Mechanical ventilation (MV) has been used throughout history as a basic tool in the treatment of patients with respiratory failure, and as a means of improving prognosis. Almost 40% of mechanically ventilated patients have difficulties during MV weaning, due to multiple factors. A delay in weaning can prolong the stay in the intensive care unit (ICU) and lead to a poorer prognosis^{1–3} and a 12% increase in mortality compared to patients with no weaning problems.⁴

In the last 20 years or so, attention has focused on the study of ventilation-induced diaphragm dysfunction (VIDD)⁵ as one of the complications associated with MV. This term refers to diaphragm dysfunction that occurs soon after initiating MV.⁶ VIDD worsens prognosis and is associated with extubation failure, which in turn prolongs MV^{7–12} and increases the risk of mortality.^{13–16} At present, however, diaphragm function is not routinely monitored in many units, suggesting that this entity may be systematically underdiagnosed.¹⁷

Ventilation-induced Diaphragm Dysfunction

Although respiratory muscle weakness could be considered part of the overall muscle involvement in a critical patient, the last 10 years have seen the emergence of the concept of VIDD.¹⁸ This term refers to diaphragm muscle dysfunction caused by the negative effect of the MV itself, that can occur along with or independently of involvement of the rest of the musculature.

Although earlier studies described how complete diaphragm inactivity induced in patients receiving controlled modality ventilation leads to a rapid, progressive loss of diaphragm function,^{19,20} it was not until 2004 that Vassilakopoulos first coined the term VIDD,¹⁸ defined as a progressive loss of diaphragm muscle strength that occurs soon after starting MV.^{8,21} The condition affects up to 65% of ventilated patients,⁷ and is clinically significant due to its early onset.

Pathophysiology of Ventilator-induced Diaphragm Dysfunction

Several studies have shown that the use of controlled MV (CMV), in which the patient makes no inspiratory effort and the diaphragm is not actively contracted, can lead to contractile dysfunction and diaphragm atrophy within 24 h in both laboratory animals and humans.^{20,22,23} Atrophied muscles lose strength and the diaphragm excursion is diminished due to the reduction in the cross-sectional area of the muscle fibers, manifesting as a loss of inspiratory capacity.^{7,24} Below we discuss the physiopathological principles associated with VIDD.

Diaphragmatic Atrophy

CMV-induced diaphragm atrophy occurs extremely rapidly.²² Significant diaphragm atrophy can be observed in laboratory animals within the first 12–18 h of starting CMV, with no sign of peripheral atrophy.²⁵ Thus, MV-induced diaphragm atrophy is significantly greater than the atrophy caused by skeletal muscle disuse.²² Levine et al. encountered similar findings in patients receiving CMV for 18–69 h, and showed a significant reduction of around 53%–57% compared to healthy volunteers in both type 1 and type 2 fibers in cross-sectional diaphragm biopsies.²³

Changes in Muscle Fiber Ultrastructure

CMV causes time-dependent changes in the ultrastructure of the diaphragm muscle fibers.^{19,26,27} First, areas containing abnormal myofibrils due to myofibrillar disorganization and Z-band changes appear.¹⁹ This is followed by the appearance of areas of muscle fiber regeneration, with no signs of inflammation.²⁷ Finally, if MV is prolonged (over 3 days), cytoplasmic lipid vacuoles increase, probably due to an autophagic process.^{27–29}

Contractile Dysfunction

In 1994, Le Bourdelles et al. were the first to describe, in an animal model, the appearance of contractile dysfunction 48 h after starting CMV.³⁰ Prolonged MV causes progressive, time-dependent loss of diaphragm strength.²⁰ As with atrophy, this reduction in diaphragm strength can be seen within 12 h of CMV.^{31,32} Several studies have shown that the maximum inspiratory pressure peak is lower in patients receiving prolonged MV compared to controls.¹⁸

Changes in Protein Synthesis

Diaphragmatic atrophy and dysfunction associated with the use of CMV occur primarily due to a reduction in protein synthesis and an increase in proteolysis.^{23,33} Diaphragmatic protein synthesis can fall by 30% after only 6 h of MV.³³ Increased proteolysis is associated with the activation of protease pathways (calpain, caspase-3, and ubiquitin-proteasome system), apoptosis pathways, and activation of autophagy.^{34,35} Hooijman et al. analyzed the activity of the ubiquitin-proteasome pathway in diaphragm biopsy samples from patients ventilated during thoracic surgery. Patients with a significant increase in this pathway showed a loss of approximately 25% of both slow and fast twitch diaphragm muscle fibers on cross-sectional biopsy, and reduced contractile strength.^{36,37}

Increased Oxidative Stress

During CMV, changes occur in mitochondrial morphology causing altered functioning and excessive creation of reactive oxygen species (ROS). Increased oxidative stress triggers the activation of calpain and caspase-3 pathways in the diaphragm.^{34,38,39} The very early appearance (at 18 h) of reduced diaphragm muscle fiber diameter and evidence of oxidative stress phenomena and activation of proteases has been confirmed in brain-dead donor patients.²³ Prolonged MV also leads to diminished antioxidant capacity in the diaphragm, reflected by reduced glutathione levels, CuZn superoxide dismutase, and peroxidase glutathione activity.⁴⁰

Effect of Critical Disease on Diaphragm Function

Although the consequences of CMV-induced diaphragm dysfunction are known, the pathways leading to these changes have yet to be described. Diaphragm inactivity induces progressive, time-dependent diaphragm atrophy.^{6,25} However, other factors also play a part in increasing the risk of VIDD and greater diaphragmatic involvement. Sepsis and multiorgan failure are the factors most robustly associated with diaphragm dysfunction, which is more severe in septic patients than in patients with shock due to other causes.⁷ The proinflammatory situation that occurs in these diseases may reduce protein synthesis and increase proteolysis, which itself may trigger an acute loss of muscle mass.^{9,41}

Concomitant factors and pharmacological treatment (corticosteroids or muscle relaxants, for example) may exacerbate VIDD. Two studies examining muscle relaxants in mice confirmed that the use of rocuronium exacerbates MV-induced contractile dysfunction.^{42,43} Data on the effect of corticosteroids on diaphragm function are contradictory. Sasoon et al. found an association between the administration of high-dose methylprednisolone and reduced inspiratory force in an experimental model.⁴⁴ However, the group led by Maes showed that administration of a single dose of corticosteroids reduced the deleterious effect of MV.⁴² Experimental data have associated high blood glucose levels with loss of diaphragm strength, related with an increase in oxidative stress processes.⁴⁵

Invasive Mechanical Ventilation Modalities and Ventilation-induced Diaphragm Dysfunction

The diaphragmatic changes characteristic of VIDD caused by controlled modality ventilation have been demonstrated in animal models and humans, and onset is early.^{8,23,2633,46,47} This modality, in which the diaphragm remains totally inactive, can cause greater atrophy and contractile dysfunction, leading to loss of diaphragm strength. Unlike CMV modalities, uncontrolled MV requires the patient to make the inspiratory effort. Several authors have suggested that this action may prevent the deleterious effects of CMV. Doering et al. showed that the prolonged use of high levels of pressure in support pressure modalities caused atrophy and contractile dysfunction.⁴⁸ In contrast, Futier et al. suggested that assisted MV and pressure support would reduce proteolysis and increase protein synthesis in the diaphragm.⁴⁹ Gayan-Ramírez et al. demonstrated that short periods of spontaneous breathing during CMV delays the effects of this modality on the diaphragm, adding credence to the idea that diaphragm disuse is the main causative factor of VIDD.⁵⁰ Changes in the configuration of the diaphragm may be prevented by maintaining inspiratory effort levels provided by pressure support.⁵¹

Non-invasive Mechanical Ventilation and High-flow Oxygen Therapy

Only 1 study has evaluated diaphragm functionality after starting non-invasive MV (NIMV). The authors conclude that the force generated by the diaphragm falls after initiating ventilation, and that this is related with high levels of pressure support.⁵² NIMV applies positive pressure, with a certain degree of flattening and muscle stretching, which may be the origin of the dysfunction.

High-flow oxygen therapy provides gas at up to 60 l/min with the corresponding FIO₂, generating low levels of positive pressure and ventilatory pattern changes, with increased circulating volume,⁵³ but to date, no studies have analyzed its impact on diaphragm function. However, as this respiratory support system ensures the patient's diaphragm is fully activated at all times, it probably has little impact on muscle strength.

Diagnosis and Monitoring

Very few UCIs conduct routine monitoring of respiratory muscle function, either because its importance is underestimated or because the technical difficulties are considerable.¹⁷ VIDD diagnosis is initially based on suspicion in mechanically ventilated patients whose intercurrent difficulties have been solved but who present problems on MV weaning. In the case of VIDD, no relationship has been established between clinical examination and the degree of diaphragm dysfunction,⁵⁴ so additional examinations are needed for diagnosis (Table 1). Below is a list of some of the best tests for evaluating diaphragm function. Some are only useful in research; others can be used in clinical practice.

Transdiaphragmatic and Esophageal Pressure

Esophageal pressure is a good indicator of pleural pressure^{55,56} and is used together with gastric pressure to calculate transdiaphragmatic pressure (Pdi). Pdi is simply the difference between pleural pressure and abdominal pressure, and is the basis for calculating diaphragm strength.^{7,13,57,58} The gold standard in patients who cannot collaborate in the test is to calculate Pdi via cervical magnetic stimulation of the phrenic nerve.^{8,54,59} However, this is an invasive technique that is not available in most ICUs.

A reduction in Pdi can be detected soon after starting MV,⁵⁹ and continues logarithmically as the duration of MV increases.⁸ Demoule et al. published a study⁷ that showed how Pdi values lower than 11 cm H₂O on the first day of ICU admission predicted a poor prognosis, high mortality (49%), and longer MV.^{7,14}

Gilbert's index, calculated using Pdi (Δ Gastric pressure/ Δ Pdi) during a normal inhalation, evaluates the contribution of the diaphragm to respiratory pressure during quiet breathing.⁶⁰ Watson et al., using diaphragm ultrasonography and Pdi in patients undergoing cardiothoracic surgery, found a good correlation between low diaphragm excursion and low Gilbert indices.⁶¹

P_{0.1}

P_{0.1} measures the negative pressure generated by the patient against an artificial airway in the first 100 ms of an inspiratory effort, reflecting the neuromuscular impulse transmitted by the phrenic nerve.⁶² Though not without limitations, it is a

Table 1
Tests for the Evaluation of Diaphragm Function in Mechanically Ventilated Patients.

	Requires Collaboration	Comment	Values	Useful in the Clinic	Useful in Research
Invasive					
<i>Measures of pressure</i>					
P _{max}	Yes	Evaluation of overall respiratory muscle strength	-30 (F)/-45 cm H ₂ O (M) ^a	+	+++
P _{Emax}		Influence: communication, sedation, anxiety	>60 (F)/80 cm H ₂ O (M) ^a	+	+++
P _{0.1}	No	Not useful as isolated parameter			
		Intensity of respiratory stimulus	0 to -2 cm H ₂ O ^a	+	+++
		Conditioned by respiratory center			
		Not useful as isolated parameter			
Pdimax.	Yes	Complex interpretation and difficult to position	>15 cm H ₂ O ^a	-	+++
PdiMS	No	Patients without ventilatory support	>10 cm H ₂ O ^b	+	+++
		Useful in sedated and ventilated patients			
		Gold standard			
EMG	No	Technically difficult	No standard values	+	+
Gilbert index	No	Contribution of the diaphragm to respiratory pressure	>0.3 ^b	+	+++
Fluoroscopy	Yes	Not useful in mechanically ventilated patients		+	+
Non-invasive					
<i>Diaphragm ultrasonography</i>					
Thickness	No	Useful in ventilated patients reproducible	>2 mm ^b	+++	+++
Shortening fraction	No	Useful in ventilated patients reproducible	>20%	+++	+++
Excursion	No	Measurement of functionality in patients with spontaneous breathing	>10 mm ^b	+++	+++

PdiMS: transdiaphragmatic pressure by magnetic stimulation of phrenic nerve; Pdimax: maximum transdiaphragmatic pressure; PEmax: maximum expiratory pressure; P_{max}: maximum inspiratory pressure.

^a Values from healthy individuals.

^b Values from ventilated patients.

measurement of central stimulus and an indicator of diaphragm function.^{63,64} This measurement has been included in artificial respirators, and, as such, it has become a clinical variable accessible at the bedside.⁶⁵

Ultrasonography

The need for early diagnosis with an easily available, non-invasive technique explains the growing use of ultrasonography in the evaluation and follow-up of diaphragm function and inspiratory effort.⁶⁶ Its limitations include inter- and intra-observer variability and the fact that the dynamic study (diaphragm excursion) can only be conducted in patients without MV.^{67–70} However, several studies using ultrasonography have shown good accuracy and reproducibility for evaluating diaphragm function, both in healthy volunteers and in critical patients.^{52,69}

Diaphragm thickness (as a sign of atrophy), shortening fraction, and diaphragm mobility studies (as a sign of diaphragm activity) are the parameters most commonly evaluated by ultrasonography.

Diaphragm Excursion

To evaluate diaphragm motion, a 3.5–5 MHz probe must be positioned below the rib cage at the level of the mid-clavicular line, directing the ultrasound beam perpendicularly toward the posterior third of the hemidiaphragm. During inhalation, the normal diaphragm moves down, and thus approaches the transducer (Fig. 1).

Normal values in healthy non-ventilated patients differ between men and women (18 ± 3 and 16 ± 3 mm, respectively),⁶⁹ similarly to baseline values in mechanically ventilated patients.⁷¹ Diaphragm dysfunction is defined as an excursion of less than 10 mm or a negative excursion (or paradoxical movement). These values are also good predictors of failure to wean.⁷²

Diaphragm Thickness

Diaphragm thickness depends on muscle mass, and is correlated with forced vital capacity.⁷³ Reduced diaphragm thickness is associated with low amplitudes on electromyography, and may indicate the presence of atrophy.^{23,74}

Diaphragm thickness can be quantified simply using ultrasonography, with the right hemidiaphragm, via the hepatic window, being more accessible than the left.⁵¹ To determine thickness, a 10–12 MHz probe, positioned perpendicularly over the 9th–10th intercostal space in the anterior axillary line, is used to observe part of the zone of apposition of diaphragm to rib cage. In this area, the diaphragm is seen as 3 parallel layers of differing density

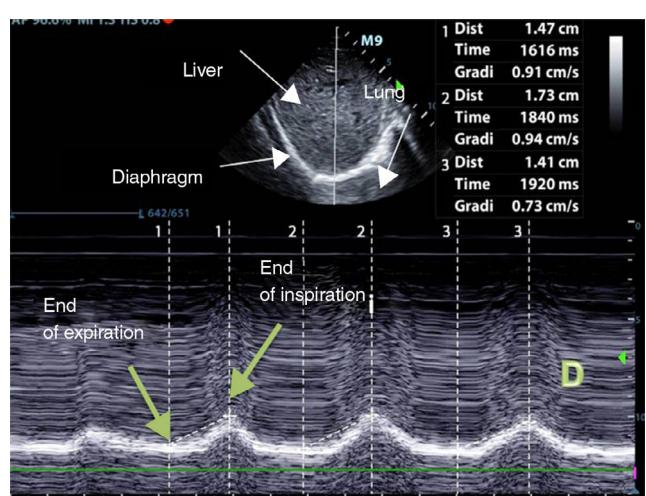


Fig. 1. M-mode measurement of diaphragm excursion during a normal inspiration. The diaphragm can be seen as a hyperechogenic line indicated with an arrow. The excursion is measured as the difference between the end of the inspiration and the end of expiration (cm).

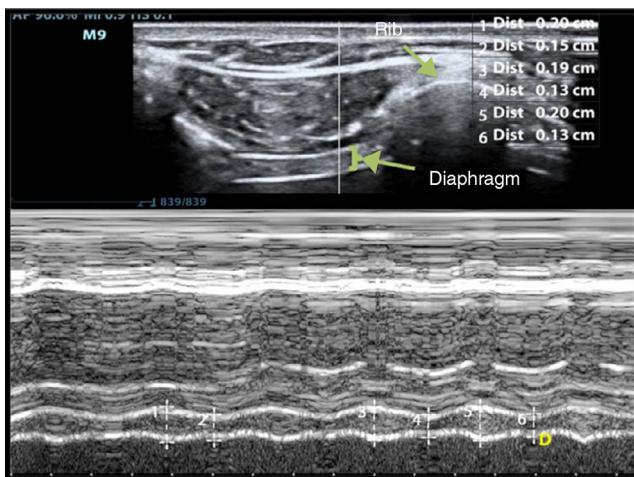


Fig. 2. M-mode measurement of diaphragm thickness variation of the right hemidiaphragm. The diaphragm is seen as 3 parallel lines: 2 hyperechogenic layers and a thicker, central hypoechoic layer. Number 1 is the inspiratory thickness and number 2 is the expiratory thickness. Diaphragm shortening fraction is the difference between the inspiratory thickness and the expiratory thickness.

(pleura, diaphragm, and peritoneum).^{51,70} Diaphragm thickness is measured by M-mode or with a 2D image during non-forced expiration (Fig. 2). Normal diaphragm thickness in ventilated patients is 2.4 ± 0.8 mm, with atrophy defined as values below 2 mm.⁵¹ Ratios of 1.8 are considered normal, with a lower accepted limit of 1.2.⁶⁸

Variation in Diaphragm Thickness

Variation in diaphragm thickness can be calculated using M-mode with the formula indicated in Fig. 3. Diaphragm thickness variation can be used as an indicator of diaphragm capacity to generate pressure.^{52,75} A variation of <20% may be considered as a predictor for failure to wean⁵⁴ and is a better prediction than the Tobin index.⁷⁶

Grosu et al. found a daily reduction of 6% in diaphragm thickness in the first 48 h after starting ventilation with assisted modalities.⁶⁷ In the study of Schepens et al., in which most patients were receiving CMV, a 10.9% loss of thickness was reported.⁷¹ Although no direct association was found between diaphragm thickness and CMV, the data show a greater loss of thickness in patients primarily ventilated using the controlled modality.

In MV patients, changes in diaphragm thickness, unlike diaphragm excursion, may really reflect active diaphragm contraction. This was demonstrated by Goligher et al., who did not observe variations in relaxed patients but did see changes in healthy volunteers and patients undergoing different MV methods.⁷⁴ In non-ventilated patients, thickness variation has been shown to be directly related with lung volumes^{70,77,78} and with maximum inspiratory pressure⁷⁹ and, as such, could be an indirect measurement of contractile activity and the diaphragm workload.

Laboratory Markers

No relationship between the appearance of respiratory muscle atrophy and increased muscle enzymes, such as creatine kinase

$$\text{Diaphragm thickness variation} = \frac{\text{Thickness at end of inspiration} - \text{Thickness at end of expiration}}{\text{Thickness at end of expiration}}$$

Fig. 3. Formula for the calculation of diaphragm thickness variation.

and myoglobin, has been observed in blood tests.⁹ An increase in skeletal protein troponin I may, in the future, be used as a marker of diaphragm muscle damage.^{80–82}

Conclusions

Diaphragm dysfunction in the mechanically ventilated patient is a disorder that is still undefined and underdiagnosed. It appears early after the initiation of MV and is associated with risk factors such as sepsis and multiorgan failure. It affects a high percentage of patients and leads to extubation failure.

Future studies are needed to determine the initial processes that trigger protein changes and to develop non-invasive techniques, accessible at the bedside, in order to establish an early diagnosis: in this respect, ultrasonography is playing an increasingly important role.

Conflict of Interests

The authors state that they have no conflict of interests.

Acknowledgement

We thank Mrs Marta Gas, from the Intensive Medicine Department of the Hospital del Mar, Barcelona, for her willingness and kind collaboration.

References

- Hermans G, van den Berghe G. Clinical review: intensive care unit acquired weakness. *Crit Care*. 2015;19:274.
- Tobin MJ. Advances in mechanical ventilation. *N Engl J Med*. 2001;344:1986–96.
- Dreyfuss D, Saumon G. State of the art: ventilator-induced lung injury. *Am J Respir Crit Care Med*. 1998;157:294–323.
- Peñuelas O, Frutos-Vivar F, Fernández C, Anzueto A, Epstein SK, Apezteguia C, et al. Characteristics and outcomes of ventilated patients according to time to liberation from mechanical ventilation. *Am J Respir Crit Care Med*. 2011;184:430–7.
- Jubran A. Critical illness and mechanical ventilation: effects on the diaphragm. *Respir Care*. 2006;51:1054–61.
- Jaber S, Jung B, Matecki S, Petrof BJ. Clinical review: ventilator-induced diaphragmatic dysfunction—human studies confirm animal model findings! *Crit Care*. 2011;15:206.
- Demoule A, Jung B, Prodanovic H, Molinari N, Chanques G, Coirault C, et al. Diaphragm dysfunction on admission to the intensive care unit. Prevalence, risk factors, and prognostic impact—a prospective study. *Am J Respir Crit Care Med*. 2013;188:213–9.
- Hermans G, Agten A, Testelmans D, Decramer M, Gayan-Ramirez G. Increased duration of mechanical ventilation is associated with decreased diaphragmatic force: a prospective observational study. *Crit Care*. 2010;14:R127.
- Puthucheary ZA, Rawal J, McPhail M, Connolly B, Ratnayake G, Chan P, et al. Acute skeletal muscle wasting in critical illness. *JAMA*. 2013;310:1591–600.
- Hermans G, Van Mechelen H, Clerckx B, Vanhullebusch T, Mesotten D, Wilmer A, et al. Acute outcomes and 1-year mortality of intensive care unit-acquired weakness. A cohort study and propensity-matched analysis. *Am J Respir Crit Care Med*. 2014;190:410–20.
- Sharshar T, Bastuji-Garin S, Stevens RD, Durand MC, Malissin I, Rodriguez P, et al. Presence and severity of intensive care unit-acquired paresis at time of awakening are associated with increased intensive care unit and hospital mortality. *Crit Care Med*. 2009;37:3047–53.
- Latronico N, Bolton CF. Critical illness polyneuropathy and myopathy: a major cause of muscle weakness and paralysis. *Lancet Neurol*. 2011;10:931–41.
- Jaber S, Petrof BJ, Jung B, Chanques G, Berthet J, Rabuel C, et al. Rapidly progressive diaphragmatic weakness and injury during mechanical ventilation in humans. *Am J Respir Crit Care Med*. 2011;183:364–71.
- Supinski GS, Ann Callahan L. Diaphragm weakness in mechanically ventilated critically ill patients. *Crit Care*. 2013;17:R120.
- Dasta JF, McLaughlin TP, Mody SH, Piech CT. Daily cost of an intensive care unit day: the contribution of mechanical ventilation. *Crit Care Med*. 2005;33:1266–71.
- Esteban A, Ferguson ND, Meade MO, Frutos-Vivar F, Apezteguia C, Brochard L, et al. Evolution of mechanical ventilation in response to clinical research. *Am J Respir Crit Care Med*. 2008;177:170–7.
- Heunks LMA, Doorduin J, van der Hoeven JG. Monitoring and preventing diaphragm injury. *Curr Opin Crit Care*. 2015;21:34–41.

18. Vassilakopoulos T, Petrof BJ. Ventilator-induced diaphragmatic dysfunction. *Am J Respir Crit Care Med.* 2004;169:336–41.
19. Sassoone CSH, Caiozzo VJ, Manka A, Sieck GC. Altered diaphragm contractile properties with controlled mechanical ventilation. *J Appl Physiol.* 2002;92:2585–95.
20. Powers SK, Shanely RA, Coombes JS, Koesterer TJ, McKenzie M, Van Gammeren D, et al. Mechanical ventilation results in progressive contractile dysfunction in the diaphragm. *J Appl Physiol.* 2002;92:1851–8.
21. Petrof BJ, Jaber S, Matecki S. Ventilator-induced diaphragmatic dysfunction. *Curr Opin Crit Care.* 2016;16:19–25.
22. Powers SK, Kavazis AN, Levine S. Prolonged mechanical ventilation alters diaphragmatic structure and function. *Crit Care Med.* 2009;37 Suppl.: S347–53.
23. Levine S, Nguyen T, Taylor N, Friscia M, Budak M, Powers SK, et al. Rapid disuse atrophy of diaphragm fibers in mechanically ventilated humans. *N Engl J Med.* 2008;358:1327–35.
24. Powers SK, Wiggs MP, Sollanek KJ, Smuder AJ. Ventilator-induced diaphragm dysfunction: cause and effect. *AJP Regul Integr Comp Physiol.* 2013;305:464–77.
25. Shanely A, Zergeroglu MA, Lennon SL, Sugiura T, Yilmalai T, Enns D, et al. Mechanical ventilation-induced diaphragmatic atrophy is associated with oxidative injury and increased proteolytic activity. *Am J Respir Crit Care Med.* 2002;166:1369–74.
26. Capdevila X, Lopez S, Bernard N, Sugiura T, Yilmalai T, Enns D, et al. Effects of controlled mechanical ventilation on respiratory muscle contractile properties in rabbits. *Intensive Care Med.* 2003;29:103–10.
27. Radell P, Edström L, Stibler H, Eriksson LI, Ansved T. Changes in diaphragm structure following prolonged mechanical ventilation in piglets. *Acta Anaesthesiol Scand.* 2004;48:430–7.
28. Bernard N, Matecki S, Py G, Lopez S, Mercier J, Capdevila X. Effects of prolonged mechanical ventilation on respiratory muscle ultrastructure and mitochondrial respiration in rabbits. *Intensive Care Med.* 2003;29:111–8.
29. Malicdan MC, Noguchi S, Nonaka I, Saftig P, Nishino I. Lysosomal myopathies: an excessive build-up in autophagosomes is too much to handle. *Neuromuscul Disord.* 2008;18:521–9.
30. Le Bourdelles G, Viires N, Boczkowski J, Seta N, Pavlovic D, Aubier M. Effects of mechanical ventilation on diaphragmatic contractile properties in rats. *Am J Respir Crit Care Med.* 1994;149:1539–44.
31. Criswell DS, Shanely RA, Betters JJ, McKenzie MJ, Sellman JE, Van Gammeren D, et al. Cumulative effects of aging and mechanical ventilation on *in vitro* diaphragm function. *Chest.* 2003;124:2302–8.
32. Davis RT, Bruells CS, Stabley JN, McCullough DJ, Powers SK, Behnke BJ. Mechanical ventilation reduces rat diaphragm blood flow and impairs oxygen delivery and uptake. *Crit Care Med.* 2012;40:2858–66.
33. Shanely RA, Van Gammeren D, Deruisseau KC, Zergeroglu M, McKenzie MJ, Yarasheski K, et al. Mechanical ventilation depresses protein synthesis in the rat diaphragm. *Am J Respir Crit Care Med.* 2004;170:994–9.
34. Hudson MB, Smuder AJ, Nelson WB, Bruells CS, Levine S, Powers SK. Both high level pressure support ventilation and controlled mechanical ventilation induce diaphragm dysfunction and atrophy. *Crit Care Med.* 2013;40:1254–60.
35. Hussain SNA, Mofarrahi M, Sigala I, Kim HC, Vassilakopoulos T, Maltais F, et al. Mechanical ventilation-induced diaphragm disuse in humans triggers autophagy. *Am J Respir Crit Care Med.* 2010;182:1377–86.
36. Hooijman PE, Beishuizen A, Witt CC, de Waard MC, Girbes ARJ, Spoelstra-de Man A, et al. Diaphragm muscle fiber weakness and ubiquitin-proteasome activation in critically ill patients. *Am J Respir Crit Care Med.* 2015;191:1126–38.
37. Barreiro E, Sznajder JJ, Nader GA, Budinger GRS. Muscle dysfunction in patients with lung diseases. A growing epidemic. *Am J Respir Crit Care Med.* 2015;191:616–9.
38. Picard M, Azuelos I, Jung B, Giordano C, Matecki S, Hussain S, et al. Mechanical ventilation triggers abnormal mitochondrial dynamics and morphology in the diaphragm. *J Appl Physiol.* 2015;118:1161–71.
39. Whidden MA, Smuder AJ, Wu M, Hudson MB, Nelson WB, Powers SK. Oxidative stress is required for mechanical ventilation-induced protease activation in the diaphragm. *J Appl Physiol.* 2010;108:1376–82.
40. Falk DJ, Deruisseau KC, van Gammeren DL, Deering MA, Kavazis AN, Powers SK. Mechanical ventilation promotes redox status alterations in the diaphragm. *J Appl Physiol.* 2006;101:1017–24.
41. Vesali RF, Cibicek N, Jakobsson T, Klaude M, Werner J, Rooyackers O. Protein metabolism in leg muscle following an endotoxin injection in healthy volunteers. *Clin Sci (Lond).* 2010;118:421–7.
42. Maes K, Testelmans D, Cadot P, DeRuisseau K, Powers SK, Decramer MA, et al. Effects of acute administration of corticosteroids during mechanical ventilation on rat diaphragm. *Am J Respir Crit Care Med.* 2008;178:1219–26.
43. Testelmans D, Maes K, Wouters P, Gosselin N, Deruisseau K, Powers SK, et al. Rocuronium exacerbates mechanical ventilation-induced diaphragm dysfunction in rats. *Crit Care Med.* 2006;34:3018–23.
44. Sassoone CSH, Zhu E, Pham HT, Nelson RS, Fang L, Baker M, et al. Acute effects of high-dose methylprednisolone on diaphragm muscle function. *Muscle Nerve.* 2008;38:1161–72.
45. Callahan LA, Supinski GS. Hyperglycemia-induced diaphragm weakness is mediated by oxidative stress. *Crit Care.* 2014;18:R88.
46. Tang H, Lee M, Khuong A, Wright E, Shrager JB. Diaphragm muscle atrophy in the mouse after long-term mechanical ventilation. *Muscle Nerve.* 2013;48:272–8.
47. Welvaart WN, Paul M, Stienen GJM, van Hees HWH, Loer S, Bouwman R, et al. Selective diaphragm muscle weakness after contractile inactivity during thoracic surgery. *Ann Surg.* 2011;254:1044–9.
48. Doering LV, Imperial-Perez F, Monsein S, Esmailian F. Preoperative and postoperative predictors of early and delayed extubation after coronary artery bypass surgery. *Am J Crit Care.* 1998;7:37–44.
49. Futier E, Constantin J-M, Combaret L, Mosoni L, Roszyk L, Sapin V, et al. Pressure support ventilation attenuates ventilator-induced protein modifications in the diaphragm. *Crit Care.* 2008;12:R116.
50. Gayan-Ramirez G, Testelmans D, Maes K, Rácz GZ, Cadot P, Zádor E, et al. Intermittent spontaneous breathing protects the rat diaphragm from mechanical ventilation effects. *Crit Care Med.* 2005;33:2804–9.
51. Goligher EC, Laghi F, Detsky ME, Farias P, Murray A, Brace D, et al. Measuring diaphragm thickness with ultrasound in mechanically ventilated patients: feasibility, reproducibility and validity. *Intensive Care Med.* 2015;41:642–9.
52. Vivier E, Dessap AM, Dimassi S, Vargas F, Lyazidi A, Thile A, et al. Diaphragm ultrasonography to estimate the work of breathing during non-invasive ventilation. *Intensive Care Med.* 2012;38:796–803.
53. Masclans JR, Pérez-Terán P, Roca O. Papel de la oxigenoterapia de alto flujo en la insuficiencia respiratoria aguda. *Med Intensiva.* 2015;39:505–15.
54. Jung B, Moury PH, Mahul M, de Jong A, Galia F, Prades A, et al. Diaphragmatic dysfunction in patients with ICU-acquired weakness and its impact on extubation failure. *Intensive Care Med.* 2016;42:853–61.
55. Akoumianaki E, Maggiore SM, Valenza F, Bellani G, Jubran A, Loring S, et al. The application of esophageal pressure measurement in patients with respiratory failure. *Am J Respir Crit Care Med.* 2014;189:520–31.
56. Talmon D, Sarge T, Malhotra A, O'Donnell CR, Ritz R, Lisbon A, et al. Mechanical ventilation guided by esophageal pressure in acute lung injury. *N Engl J Med.* 2008;359:2095–104.
57. Hamnegård CH, Wragg S, Mills G, Kyroussis D, Road J, Daskos G, et al. The effect of lung volume on transdiaphragmatic pressure. *Eur Respir J.* 1995;8:1532–6.
58. Mier A, Brophy C, Moxham J, Green M. Twitch pressures in the assessment of diaphragm weakness. *Thorax.* 1989;44:990–6.
59. Watson AC, Hughes PD, Louise Harris M, Hart N, Ware RJ, Wendon J, et al. Measurement of twitch transdiaphragmatic, esophageal, and endotracheal tube pressure with bilateral anterolateral magnetic phrenic nerve stimulation in patients in the intensive care unit. *Crit Care Med.* 2001;29:1325–31.
60. Gilbert R, Auschincloss JH, Peppi D. Relationship of rib cage and abdomen motion to diaphragm function during quiet breathing. *Chest.* 1981;80:607–12.
61. Watson AC, Hughes PD, Louise Harris M, Hart N, Ware RJ, Wendon J, et al. Ultrasonographic diagnostic criterion for severe diaphragmatic dysfunction after cardiac surgery. *Chest.* 2009;135:401–7.
62. Kera T, Aihara A, Inomata T. Reliability of airway occlusion pressure as an index of respiratory motor output. *Respir Care.* 2013;58:845–9.
63. Umbrello M, Formenti P, Longhi D, Galimberti A, Piva I, Pezzi A, et al. Diaphragm ultrasound as indicator of respiratory effort in critically ill patients undergoing assisted mechanical ventilation: a pilot clinical study. *Crit Care.* 2015;19:1–10.
64. Ko R, Ramos L, Chalela JA. Conventional weaning parameters do not predict extubation failure in neurocritical care patients. *Neurocrit Care.* 2009;10:269–73.
65. Fernandez R, Raurich JM, Mut T, Blanco J, Santos A, Villagra A. Extubation failure: diagnostic value of occlusion pressure (P_O.1) and P_O.1-derived parameters. *Intensive Care Med.* 2004;30:234–40.
66. Baldwin CE, Paratz JD, Bersten AD. Diaphragm and peripheral muscle thickness on ultrasound: intra-rater reliability and variability of a methodology using non-standard recumbent positions. *Respirology.* 2011;16:1136–43.
67. Grossi HB, Lee YI, Lee J, Eden E, Eikermann M, Rose KM. Diaphragm muscle thinning in patients who are mechanically ventilated. *Chest.* 2012;142:1455–60.
68. Boon AJ, Harper CJ, Ghahfarokhi LS, Strommen JA, Watson JC, Sorenson EJ. Two-dimensional ultrasound imaging of the diaphragm: quantitative values in normal subjects. *Muscle Nerve.* 2013;47:884–9.
69. Boussuges A, Gole Y, Blanc P. Diaphragmatic motion studied by M-mode ultrasonography. *Chest.* 2009;135:391–400.
70. Cohn D, Benditt JO, Eveloff S, McCool FD. Diaphragm thickening during inspiration. *J Appl Physiol.* 1997;83:291–6.
71. Schepens T, Verbrugge W, Dams K, Corthouts B, Parizel PM, Jorens PG. The course of diaphragm atrophy in ventilated patients assessed with ultrasound: a longitudinal cohort study. *Crit Care.* 2015;19:422.
72. Kim WY, Suh HJ, Hong S-B, Koh Y, Lim C-M. Diaphragm dysfunction assessed by ultrasonography: influence on weaning from mechanical ventilation. *Crit Care Med.* 2011;39:2627–30.
73. Noda Y, Sekiguchi K, Kohara N, Kanda F, Toda T. Ultrasonographic diaphragm thickness correlates with compound muscle action potential amplitude and forced vital capacity. *Muscle Nerve.* 2015;53:522–7.
74. Goligher EC, Fan E, Herridge MS, Murray A, Vorona S, Brace D, et al. Evolution of diaphragm thickness during mechanical ventilation: impact of inspiratory effort. *Am J Respir Crit Care Med.* 2015;192:1–56.
75. Matamis D, Soilemezi E, Tsagourias M, Akoumianaki E, Dimassi S, Boroli F, et al. Sonographic evaluation of the diaphragm in critically ill patients. Technique and clinical applications. *Intensive Care Med.* 2013;39:801–10.
76. DiNino E, Gartman EJ, Sethi JM, McCool FD. Diaphragm ultrasound as a predictor of successful extubation from mechanical ventilation. *Thorax.* 2014;69:423–7.
77. Summerhill EM, el-Sameed YA, Glidden TJ, McCool FD. Monitoring recovery from diaphragm paralysis with ultrasound. *Chest.* 2008;133:737–43.
78. Polkey MI, Hamnegård C, Hughes PD, Rafferty GF, Green M, Moxham J. Influence of acute lung volume change on contractile properties of human diaphragm. *J Appl Physiol.* 1998;85:1322–8.

79. Ueki J, De Bruin PF, Pride NB. In vivo assessment of diaphragm contraction by ultrasound in normal subjects. *Thorax*. 1995;50:1157–61.
80. Foster GE, Nakano J, Sheel AW, Simpson JA, Road JD, Reid WD. Serum skeletal troponin I following inspiratory threshold loading in healthy young and middle-aged men. *Eur J Appl Physiol*. 2012;112:3547–58.
81. Nedergaard A, Karsdal MA, Sun S, Henriksen K. Serological muscle loss biomarkers: an overview of current concepts and future possibilities. *J Cachexia Sarcopenia Muscle*. 2013;4:1–17.
82. Simpson JA, van Eyk J, Iscoe S. Respiratory muscle injury, fatigue and serum skeletal troponin I in rat. *J Physiol*. 2004;554:891–903.