



## Special Article

Spanish Consensus on the Prevention and Treatment of *Pseudomonas aeruginosa* Bronchial Infections in Cystic Fibrosis Patients<sup>☆</sup>

Rafael Cantón,<sup>a,b,\*</sup> Luis Máiz,<sup>c</sup> Amparo Escribano,<sup>d</sup> Casilda Olveira,<sup>e</sup> Antonio Oliver,<sup>b,f</sup> Oscar Asensio,<sup>g</sup> Silvia Gartner,<sup>h</sup> Eva Roma,<sup>i</sup> Esther Quintana-Gallego,<sup>j</sup> Antonio Salcedo,<sup>k</sup> Rosa Girón,<sup>l</sup> María Isabel Barrio,<sup>m</sup> María Dolores Pastor,<sup>n</sup> Concepción Prados,<sup>o</sup> María Teresa Martínez-Martínez,<sup>p</sup> José Barberán,<sup>q</sup> Juan José Castón,<sup>r</sup> Luis Martínez-Martínez,<sup>b,s</sup> José Luis Poveda,<sup>i</sup> Carlos Vázquez,<sup>t</sup> Javier de Gracia,<sup>u</sup> Amparo Solé<sup>v</sup>, representing the Grupo Español de Consenso del Tratamiento Antimicrobiano en el Paciente con Fibrosis Quística<sup>◇</sup>

<sup>a</sup> Servicio de Microbiología, Hospital Universitario Ramón y Cajal e Instituto Ramón y Cajal de Investigaciones Sanitarias (IRYCIS), Madrid, Spain

<sup>b</sup> Red Española de Investigación en Patología Infecciosa (REIPI), Instituto de Salud Carlos III, Madrid, Spain

<sup>c</sup> Unidad de Bronquiectasias y Fibrosis Quística, Servicio de Neumología, Hospital Universitario Ramón y Cajal e Instituto Ramón y Cajal de Investigaciones Sanitarias (IRYCIS), Madrid, Spain

<sup>d</sup> Unidad de Neumología Pediátrica y Fibrosis Quística, Servicio de Pediatría, Hospital Clínico Universitario, Universidad de Valencia, Valencia, Spain

<sup>e</sup> Unidad de Gestión Clínica de Enfermedades Respiratorias, Hospital Regional Universitario de Málaga, Instituto de Investigación Biomédica de Málaga (IBIMA), Universidad de Málaga, Málaga, Spain

<sup>f</sup> Servicio de Microbiología y Unidad de Investigación, Hospital Universitario Son Espases, Instituto de Investigación Sanitaria de Palma (IdISPa), Palma de Mallorca, Spain

<sup>g</sup> Unidad de Neumología y Alergia Pediátrica, Hospital Universitario de Sabadell. Corporació Sanitària Parc Taulí, Sabadell, Barcelona, Spain

<sup>h</sup> Unidad de Neumología Pediátrica y Fibrosis Quística, Hospital Universitario Vall d'Hebron, Barcelona, Spain

<sup>i</sup> Servicio de Farmacia, Hospital Universitario y Politécnico La Fe, Valencia, Spain

<sup>j</sup> Unidad de Fibrosis Quística, Servicio de Neumología, Hospital Universitario Virgen del Rocío, Sevilla, Spain

<sup>k</sup> Unidad de Fibrosis Quística Interhospitalaria Niño Jesús-Gregorio Marañón, Madrid, Spain

<sup>l</sup> Unidad de Bronquiectasias y Fibrosis Quística, Hospital Universitario La Princesa, Instituto La Princesa de Investigación Sanitaria, Madrid, Spain

<sup>m</sup> Sección de Neumología Pediátrica y Unidad de Fibrosis Quística, Hospital Universitario La Paz, Madrid, Spain

<sup>n</sup> Unidad de Neumología Pediátrica y Fibrosis Quística, Hospital Universitario Virgen de la Arrixaca, Murcia, Spain

<sup>o</sup> Unidad de Fibrosis Quística y Bronquiectasias, Servicio de Neumología, Hospital Universitario La Paz, Madrid, Spain

<sup>p</sup> Servicio de Neumología, Unidad de Fibrosis Quística, Hospital Universitario 12 de Octubre, Madrid, Spain

<sup>q</sup> Departamento de Medicina Interna, Hospital Montepíncipe, Universidad CEU San Pablo, Madrid, Spain

<sup>r</sup> Unidad de Enfermedades Infecciosas, Hospital General Universitario de Ciudad Real, Ciudad Real, Spain

<sup>s</sup> Servicio de Microbiología, Hospital Universitario Marqués de Valdecilla-IDIVAL y Departamento de Biología Molecular, Universidad de Cantabria, Santander, Spain

<sup>t</sup> Unidad de Neumología Pediátrica y Fibrosis Quística, Hospital Universitario de Cruces, Baracaldo, Vizcaya, Spain

<sup>u</sup> Servicio de Neumología y CIBER en Enfermedades Respiratorias (CibeRES), Hospital Universitari Vall d'Hebron, Universidad Autònoma, Barcelona, Spain

<sup>v</sup> Unidad de Trasplante Pulmonar y Fibrosis Quística, Hospital Universitario y Politécnico la Fe, Valencia, Spain

## ARTICLE INFO

## Article history:

Received 5 April 2014

Accepted 22 September 2014

Available online 13 February 2015

## Keywords:

Cystic fibrosis

*Pseudomonas aeruginosa*

Bronchial infection

Antibiotic treatment

## ABSTRACT

*Pseudomonas aeruginosa* is the main pathogen in bronchopulmonary infections in cystic fibrosis (CF) patients. It can only be eradicated at early infection stages while reduction of its bacterial load is the therapeutic goal during chronic infection or exacerbations. Neonatal screening and pharmacokinetic/pharmacodynamic knowledge have modified the management of CF-patient. A culture based microbiological follow-up should be performed in patients with no infection with *P. aeruginosa*. At initial infection, inhaled colistin (0.5–2 MU/tid), tobramycin (300 mg/bid) or aztreonam (75 mg/tid) with or without oral ciprofloxacin (15–20 mg/kg/bid, 2–3 weeks) are recommended. In chronic infections, treatment is based on continuous administration of colistin or with a 28-day on-off regimen with tobramycin or aztreonam. During mild-moderate exacerbations oral ciprofloxacin (2–3 weeks) can be administered while serious exacerbations must be treated with intravenous combination therapy (beta-lactam with an aminoglycoside or a fluoroquinolone). Future studies will support rotation and/or new combination therapies. Epidemiological measures are also recommended to avoid new *P. aeruginosa* infections and cross transmission of this pathogen.

© 2014 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

<sup>☆</sup> Please cite this article as: Cantón R, Máiz L, Escribano A, Olveira C, Oliver A, Asensio O, et al. Consenso español para la prevención y el tratamiento de la infección bronquial por *Pseudomonas aeruginosa* en el paciente con fibrosis quística. Arch Bronconeumol. 2015;51:140–150.

\* Corresponding author.

E-mail address: rafael.canton@salud.madrid.org (R. Cantón).

◇ Spanish Consensus Group for Antimicrobial Treatment in the Cystic Fibrosis Patient (SEFQ), Infectious Diseases and Clinical Microbiology (SEIMC), Pediatric Pulmonology (SENP), Pulmonology and Thoracic Surgery (SEPAR), Chemotherapy (SEQ) and Hospital Pharmacy (SEFH).

## Consenso español para la prevención y el tratamiento de la infección bronquial por *Pseudomonas aeruginosa* en el paciente con fibrosis quística

**Palabras clave:**  
Fibrosis quística  
*Pseudomonas aeruginosa*  
Infección bronquial  
Tratamiento antibiótico

**RESUMEN:** *Pseudomonas aeruginosa* es el patógeno más importante en la infección broncopulmonar en fibrosis quística (FQ). Solo se erradica en la infección inicial, mientras que la reducción de su carga bacteriana es el objetivo terapéutico en la infección crónica y exacerbaciones. El cribado neonatal y la farmacocinética/farmacodinámica han cambiado el manejo del paciente con FQ. Se debe realizar un seguimiento microbiológico en los pacientes sin infección por *P. aeruginosa*. En la infección inicial se recomienda tratamiento inhalado (28 días) con colistina (0,5–2 MU/8 h), tobramicina (300 mg/12 h) o aztreonam (75 mg/8 h) con o sin ciprofloxacino oral (15–20 mg/kg/12 h, 2–3 semanas). En la infección crónica se recomienda solo vía inhalada en tratamiento continuo con colistina, o en ciclos *on-off* de 28 días con tobramicina o aztreonam. Durante las exacerbaciones leves-moderadas se recomienda tratamiento oral (ciprofloxacino, 2–3 semanas) y en las graves tratamiento intravenoso ( $\beta$ -lactámico asociado a un aminoglucósido o una fluoroquinolona). Estudios futuros sustentarán la rotación y nuevas combinaciones de antimicrobianos. Se deben establecer también medidas epidemiológicas que eviten nuevas infecciones y la transmisión cruzada de *P. aeruginosa*.

© 2014 SEPAR. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

### Introduction

The management and evolution of cystic fibrosis (CF) patients are directed by the presence of *Pseudomonas aeruginosa* (PA) bronchial colonization. In 2005, the Spanish Consensus Group for Antimicrobial Therapy in the CF Patient published a consensus document on the use of antibiotics to treat *P. aeruginosa* colonization in CF.<sup>1</sup> It was the first report to consider the pharmacokinetics (PK) and pharmacodynamics (PD) of treatment strategies and the dynamics of bacterial infection in CF. The publication of additional recommendations,<sup>2–6</sup> neonatal screening programs,<sup>7</sup> the authorization of new antibiotics for CF<sup>8,9</sup> and new scientific developments<sup>10–19</sup> have prompted us to update our earlier report. Recommendations have been graded according to their level of evidence<sup>20</sup> based on studies published in PubMed and weighted according to clinical experience (Table 1).

### Clinical Evaluation

PA can be detected early by frequent bacterial culture of respiratory secretions.<sup>21–24</sup> This is particularly useful in children screening positive for CF, as these patients are usually asymptomatic (III-A). Other useful strategies include monitoring patients for PA antibodies (III-B),<sup>25,26</sup> polymerase chain reaction (III-B),<sup>27</sup> and study of sputum for volatile organic compounds (III-C).<sup>28</sup>

The prevalence of PA colonization increases with age, and can be as high as 80% in patients aged >18 years.<sup>29–36</sup> Persistently high bacterial load and the change from a non-mucoid to a mucoid morphotype correlate with more exacerbations, pulmonary decline and mortality.<sup>29,32,36</sup> Exacerbations (Table 2) are characterized by changes in existing symptoms and the appearance of new symptoms.<sup>35,37,38</sup>

Lung function can be normal in patients with primary PA infection. Initially, changes are observed in hyperinflation markers, such as residual volume (RV), RV/total lung capacity, and mesoexpiratory flows. Lung clearance index, a measure of inert gas washout using multiple-breath tests, detects minimal abnormalities, and is higher in patients with PA infection. Test results within normal limits have an excellent negative predictive value (93%) for ruling out PA infection (III-B).<sup>33,39–41</sup>

Reduced forced expiratory volume in 1 s (FEV1) shows progression of the infection and is a good measure of the severity of exacerbations (III-A).<sup>42</sup> Chest computed tomography (HRCT) provides early detection of airway decline and is useful in

estimating severity (III-A).<sup>43–46</sup> Lung infection is associated with increased air trapping and greater structural damage, including bronchiectasis.<sup>47</sup> HRTC scores are worse in PA colonization<sup>29,48,49</sup> and during exacerbations.<sup>50</sup>

### Patterns of Infection and Microbiological Evaluation

Pulmonary infection in CF can be classified into 3 phases<sup>1</sup>:

- *Initial colonization (primary colonization)*. Detection of the first positive PA culture in which non-mucoid, antibiotic-sensitive strains are usually isolated. A negative culture after the first positive culture can indicate an aborted initial colonization, cryptic colonization, or eradication of PA following antibiotic therapy. PA is considered eradicated when negative results are obtained from at least 2 cultures: 1 taken 1–2 weeks following completion of treatment and another 2–4 weeks later.<sup>51</sup>
- *Intermittent colonization*. This is indicated by intermittent positive and negative results from consecutive cultures after an initial infection. It can indicate permanent low-grade colonization, sample heterogeneity (different origins), or apparently transient eradication with colonization persisting in the sinuses, causing intermittent reinfection.<sup>3,52</sup> Different strains are isolated, including mucoid morphotypes.
- *Chronic infection*. Common in CR patients with advanced disease. Mucoid colonies with a variety of other morphotypes are found as a result of specialization and adaptation of PA to the endobronchial environment. The causes of exacerbations are still unclear, but they usually coincide with increases in bacterial load or the emergence of antigenic variants.

Table 3 shows the patterns of infection defined in 2005<sup>1</sup> and updated according to the Leeds criteria and correlated with clinical parameters (II-A).<sup>53,54</sup>

### Basic Antimicrobial Treatment Strategies

Treatment of PA infection is essential for controlling progression of respiratory disease. PA is rarely eradicated once chronic infection has set in, so the best treatment strategy is early prevention (I-A). Inhaled antibiotics are the treatment of choice in PA colonization in view of the particular niche in which it thrives, its tendency to form biofilms, and the inability of some oral or intravenous antibiotics to reach effective concentrations in the respiratory mucosa.<sup>9,18,55–58</sup> This strategy can also help prevent the development of drug resistance, although it is important to

**Table 1**  
Levels of Clinical Evidence.<sup>20</sup>

Quality of evidence		Strength of recommendation <sup>a</sup>	
Grade	Definition	Category	Definition
I	Evidence obtained from at least one randomized controlled trial	A	Good quality evidence for use
II	Evidence obtained from non-randomized controlled trials or from cohort or case-control analytic studies, preferably from more than one center	B	Moderate quality evidence for use
III	Recommendation made by experts based on clinical experience or case studies	C	Poor quality evidence for use
		D	Moderate quality evidence against use

<sup>a</sup> Weighted according to clinical experience.

**Table 2**  
Criteria for Defining Exacerbations in Cystic Fibrosis Patients.<sup>36,38,39</sup>

Signs and symptoms	Radiological, functional and/or laboratory criteria
<ul style="list-style-type: none"> <li>• Changes in cough intensity and/or characteristics</li> <li>• Changes in sputum (increase in volume, purulence and/or consistency)</li> <li>• Increase in or appearance of dyspnea and/or reduced exercise tolerance</li> <li>• Anorexia, asthenia and/or weight loss</li> <li>• Chest pain</li> <li>• Fever <math>\geq 38^{\circ}\text{C}</math> more than once over the previous week</li> <li>• Increased breathing rate</li> <li>• Changes in auscultation</li> <li>• Hemoptysis or other complications</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\text{FEV}_1 \geq 10\%</math> lower than baseline values over the previous 3 months</li> <li>• <math>\text{O}_2</math> saturation <math>\geq 10\%</math> lower than baseline values over the previous 3 months</li> <li>• Increase in inflammatory markers (ESR, C-reactive protein, etc.)</li> <li>• Increase in air trapping or appearance of radiologic evidence of infiltrates</li> <li>• Changes in the density of microbial colonization or acquisition of new bacteria</li> <li>• Changes or increase in levels of antibodies for <i>P. aeruginosa</i></li> </ul>

**Table 3**  
Microbiological Patterns and Criteria in Pulmonary *Pseudomonas aeruginosa* Colonization/infection in Cystic Fibrosis Patients.<sup>1,53,54</sup>

Stages	Definition	Microbiological criteria	Comments
I: Initial infection (primary or pioneer colonization)	Detection of the first positive <i>P. aeruginosa</i> culture in the bronchial tree. No clinical symptoms or specific immunological response, aside from possible inflammation.	First positive <i>P. aeruginosa</i> culture	A positive culture following 1 year of negative cultures after finishing treatment is considered a new initial colonization. The strains are usually non-mucoid, with little diversity in morphotypes or antimicrobial susceptibilities. There is usually no specific immune response to <i>P. aeruginosa</i> .
II. Intermittent colonization	Intermittent presence of positive and negative <i>P. aeruginosa</i> cultures in consecutive samples after initial colonization. There may be no clinical signs, but there is usually some kind of immune response	$\leq 50\%$ of cultures positive for <i>P. aeruginosa</i> over the previous 12 months	Mucoid colonies and other colonial morphotypes may appear. There may be a specific immune response to <i>P. aeruginosa</i> .
III: Chronic colonization	Persistent positive <i>P. aeruginosa</i> cultures with no new clinical signs of infection, but positive immune response	$>50\%$ of cultures positive for <i>P. aeruginosa</i> over the previous 12 months	Mucoid colonies and other colonial morphotypes usually appear. This is the common pattern in advanced disease stages. Patients exhibit an immune response consistent with <i>P. aeruginosa</i> colonization.
IV: Exacerbation	Onset or increase in respiratory symptoms during chronic colonization	Cultures positive for <i>P. aeruginosa</i>	Bacterial load may increase. In patients with no microbiological studies, increased antibodies in two consecutive blood samples can be used as a diagnostic criterion.

**Table 4**Recommended Dose and Route of Administration of Antibiotics to Treat Exacerbations Due to *Pseudomonas aeruginosa* Colonization in Cystic Fibrosis Patients.

Group or family of antimicrobials		Route of administration	Dosing schedule in:	
			Children (<50 kg)	Adults (>50 kg)
Penicillins	Piperacillin/tazobactam	IV	100 mg/kg/6 h	2–4 <sup>a</sup> g/6–8 h
Cephalosporins	Ceftazidime	IV	50–70 mg/kg/8 h	2 g/8 h
	Cefepime	IV	50 mg/kg/8 h	2 g/8 h
Other $\beta$ -lactams	Aztreonam	IV or IM	50 mg/kg/8 h	1–2 g/6–8 h
	Imipenem	IV or IM	15–25 mg/kg/6 h	1 g/6–8 h
	Meropenem	IV	20–40 mg/kg/8 h	2 g/8 h
	Doripenem	IV	Not recommended	0.5–1 g/8 h <sup>b</sup>
Aminoglycosides	Gentamicin	IV or IM	10–15 mg/kg/24 h	10–15 mg/kg/24 h
	Tobramycin	IV or IM	5–10 mg/kg/24 h	5–10 mg/kg/24 h
	Amikacin	IV or IM	30–35 mg/kg/24 h	30–35 mg/kg/24 h
Quinolones	Ciprofloxacin	Oral	15–20 mg/kg/12 h	0.75 g/12 h
		IV	15–20 mg/kg/12 h	0.4 g/12 h
	Levofloxacin	Oral	ND	0.5 g/12 h
Others	Colistin	IV		0.75 g/24 h
		IV or IM	20 000 IU/kg/8 h	2 000 000 IU/8 h

IV: intravenous; IM: intramuscular; ND: not defined.

<sup>a</sup> Expressed as piperacillin.<sup>b</sup> 4 h infusion.

monitor for the emergence of opportunistic infection from other bacteria.

Standard preparations specifically designed for this route of administration are recommended, as these are less likely to cause local adverse effects (mainly bronchospasm) (III-A).

The blood pharmacokinetics of antibiotics, meanwhile, differ from those of non-CF patients,<sup>17,61–63</sup> and for this reason systemically administered doses (for example, during exacerbations) are not the same as those given to non-CF patients (Table 4), and have an impact on the antibiotic's PK/PD index of efficacy.<sup>58–60,64–72</sup>

#### Antimicrobial Prophylaxis and Vaccination

There is no evidence to support the effectiveness of administering antibiotics before initial PA colonization, nor of PA vaccination, so these strategies are not recommended (I-A).<sup>73,74</sup>

#### Primary Infection

Early treatment of primary PA colonization slows the development of chronic colonization (I-A). Treatment should be started as soon as the bacterium is isolated, as the eradication rates are very high (63%–100%).<sup>42,75–81</sup> There are various eradication protocols, none of which have been shown to be clearly superior (I-A).<sup>3,55,82</sup> Differences in design, in the definition of eradication, and in antibiotic treatment, make it difficult to compare studies published to date.<sup>79</sup> Primary colonization is usually asymptomatic; however, if it debuts as an exacerbation, standard treatment regimens are recommended (III-A).

Twice-daily colistimethate sodium, taken together with oral ciprofloxacin for periods ranging from 3 weeks to 3 months,<sup>83,84</sup> was the first antibiotic to have a significant effect on chronic infection. In Spain, this drug is marketed under the brand names GES<sup>®</sup> (Genéricos Españoles Laboratorio) and Promixin<sup>®</sup> (Praxis Pharmaceutical).

Inhaled tobramycin (300 mg/5 ml/12 h) (TNS, TOBI<sup>®</sup>, Novartis) is also a safe and effective therapy for eradication of PA, and can even be used in children aged from 6 months to 6 years.<sup>78,80,85,86</sup> The ELITE<sup>80</sup> study compared the safety and efficacy of TNS (300 mg/5 ml) for 28 and 56 days following the first PA-positive culture, finding both strategies to be similar. Over 90% of patients tested negative for PA after 1 month of treatment, and most (69%) remained infection-free at 27 months follow-up. These

and other studies suggest that short-duration (1–3 months) inhaled antimicrobials should be the treatment of choice (II-A).<sup>79,84–86</sup>

In the EPIC<sup>85</sup> study, patients (aged from 1 to 12 years) were randomized to different treatment regimens for 18 months: combined TNS and oral ciprofloxacin cyclically every 3 months or placebo, or combined TNS plus oral ciprofloxacin or placebo only when 3-monthly sputum culture was positive for PA. TNS (300 mg/5 ml) was given in 28 days on/28 days off cycles for 18 months, and ciprofloxacin or placebo was administered for 14 days. No significant differences were observed between groups, suggesting that the addition of oral ciprofloxacin does not improve the efficacy of inhaled TNS.

Taccetti et al. in a study comparing TNS (300 mg/5 ml or 300 mg/4 ml) with inhaled colistin (2 MU/12 h) for 28 days, both in combination with oral ciprofloxacin, found that neither regimen was clearly superior.<sup>79</sup>

Recently, the ALPINE study of aztreonam lysine for inhalation (AZLI, Cayston<sup>®</sup>, Gilead), 75 mg/8 h for 28 days, to treat primary colonization in patients aged between 3 months and 18 years showed that 89.1% of patients were negative for PA at the end of treatment, and that 75.2% and 58.2% remained free of infection at 28 days and 6 months, respectively, after the end of treatment.<sup>87</sup>

In the light of these studies, inhaled antibiotic treatment with colistimethate sodium (0.5–2 MU, 2 or 3 $\times$ day) should be given for 1 month (I-A), or TNS (300 mg/12 h) for 28 days (I-A), or AZLI (75 mg/8 h) (II-A), either as monotherapy or in combination with oral ciprofloxacin (15–20 mg/kg/12 h) (maximum 2 g/day), for 2–3 weeks (Table 5). Based on the analysis of microbiological tests, combination therapy with ciprofloxacin improves antibiotic action.<sup>89</sup>

A follow-up culture should be taken 1–2 weeks after the end of the eradication therapy.<sup>52,88,90–92</sup> If the culture is negative, TNS or AZLI should be prolonged for between 1 and 3 on/off (28-day) cycles (maximum 3–6 months), or 3–6 months uninterrupted colistimethate (II-A). If the culture is positive, the original regimen should be repeated, or a new regimen (ciprofloxacin in combination with an inhaled antibiotic not previously used) should be ordered (III-A). Cultures should be repeated at the end of the second cycle. If positive, the chronic infection protocol should be applied (I-A), with at least 1 follow-up culture every 3 months (III-A).

There is no conclusive data defining the number of times eradication cycles should be repeated following initial treatment failure. If PA persists after the second eradication attempt, an alternative strategy, such as combination inhaled and intravenous antibiotics

**Table 5**  
Antimicrobial Treatment for *Pseudomonas aeruginosa* Bronchial Colonization in Cystic Fibrosis Patients According to Their Clinical Status. (Inhaled Antimicrobials Are Listed in Chronological Order of Their Use in Spain, not in Order of Recommendation.).

Clinical status	Treatment	Comments
<i>First positive P. aeruginosa culture (primary colonization)</i>		
No symptoms	Inhaled colistin for 1 month, 0.5–2 million U, 2–3×day (I-A)±Oral ciprofloxacin 15–20 mg/kg twice daily 2–3 weeks or Inhaled tobramycin for 28 days, 300 mg, twice daily (I-A)±Oral ciprofloxacin 15–20 mg/kg twice daily 2–3 weeks or Inhaled aztreonam for 28 days, 75 mg, 3 x day (II-A) <sup>a</sup> ±Oral ciprofloxacin 15–20 mg/kg twice daily 2–3 weeks	Take culture 1–2 weeks after end of treatment: • <i>if negative</i> : continue with the same colistin regimen (maximum 3–6 months) or order 1–3 on/off cycles of tobramycin or aztreonam (maximum 6 months) (II-A). • <i>if positive</i> : repeat the same treatment or change to another combination (ciprofloxacin+an inhaled antibiotic not used in the first cycle) (III-A). Repeat culture 1–2 weeks after end of treatment. If still positive, apply protocol for chronic infection (I-A). Eradication protocols studied to date vary greatly and include both oral and inhaled antibiotics. If at least 2 of these strategies fail, combined inhaled+IV antibiotics have been suggested (I-B). Always differentiate between primary colonization in children screening positive for cystic fibrosis with little lung damage and colonization in adults or in recent <i>P. aeruginosa</i> recolonization. A shorter course of treatment (1–3 months) may suffice in children with primary colonization.
With symptoms	Follow recommendation for chronic colonization with exacerbation.	
<i>Chronic P. aeruginosa colonization</i>		
Stable	Continuous inhaled colistin therapy, 0.5–2 million U, 2–3×day (III-A) <sup>b,c</sup> or 28-day on/off cycles of inhaled tobramycin, 300 mg, twice daily (I-A) On/off cycles of inhaled tobramycin dry powder, 112 mg, twice daily (I-A) <sup>d</sup> or 28-day on/off cycles of inhaled aztreonam, 75 mg, 3×day (I-A) <sup>e</sup>	Maintain inhalation regimen while the benefits outweigh the risks. For moderate–severe pulmonary infection or poor response to on/off cycles: • use continuous inhaled antibiotics, alternating/rotating without rest periods or with shorter on/off periods (less than 28 days) (III-B). • combine with an oral or IV antipseudomonal antibiotic <sup>f</sup> according to sensitivity, as needed, or in cycles (every 3–6 months) (III-B).
Exacerbation <sup>g</sup>	<i>Mild exacerbation</i> : Antipseudomonal antibiotic therapy, 2–3 weeks <i>Severe exacerbation</i> : IV ceftazidime 50–70 mg/kg/8 h or IV cefepime 50 mg/kg/8 h 2–3 weeks	Consider continuing inhaled schedule if previously ordered (III-C) Prolong IV treatment if no improvement or severe infection (3–4 weeks) (III-A) If there is no response to treatment, or in the case of multi-drug resistance, adjust combined IV therapy to the sensitivity profile using antipseudomonal penicillins (piperacillin/tazobactam), monobactams (aztreonam) or a combination of a carbapenem (imipenem, meropenem or doripenem) and an aminoglycoside or fluoroquinolone.
	+ IV tobramycin 5–10 mg/kg/24 h or IV amikacin 30–35 mg/kg/24 h 2–3 weeks	

IV: intravenous.

<sup>a</sup> In patients aged over 3 months and under 18 years.

<sup>b</sup> The level of evidence is based on the literature and not on wide clinical experience with the drug. Therefore, although the level of evidence differs from that of new drugs, it has a high grade of recommendation.

<sup>c</sup> 3×day in case of aggressive clinical course.

<sup>d</sup> Sold in Spain without health authority reimbursement.

<sup>e</sup> In patients aged 6 years or older.

<sup>f</sup> Treat as in exacerbation.

<sup>g</sup> Use this regimen in primary colonization with symptoms of exacerbation.

(I-B) or the chronic infection protocol (I-A), could be used.<sup>3</sup> The appearance of a positive culture after 1 year of negative cultures should be treated as a new initial colonization event (III-A).

### Chronic Infection

Long-term antibiotic therapy has been shown to be effective against chronic PA colonization (I-A),<sup>92</sup> inhalation being the route of choice (I-A).<sup>10,93</sup> Comparative studies in inhaled antibiotics are limited insofar as they do not usually include patients not previously treated with the study drugs.

Colistimethate sodium with no antibiotic-free rest periods has been shown to be effective (III-A).<sup>2,94</sup> The usual dose in adults is 0.5–2 MU, 2–3×day, and in children 1 MU, 2–3×day. However, this dose can be halved with some nebulizers. In a trial comparing TNS vs colistimethate sodium, most patients had previously received colistimethate. In both groups, the PA colony count fell significantly, although only the TNS group showed significant improvement in lung function.<sup>95</sup>

Colistimethate in dry powder (Colobreathe<sup>®</sup>, Forest Laboratories) (125 mg/12 h) is indicated in patients aged over 6 years with chronic PA colonization,<sup>72</sup> and has been shown to be safe and equally effective as TNS. TNS (300 mg/5 ml/12 h) in 28-day on/off cycles significantly improves FEV1, and reduces PA density in sputum, the number of hospitalizations, need for intravenous antibiotics and number of exacerbations (I-A),<sup>57,96</sup> even in patients with mild-moderate lung infection.<sup>96</sup> It also improves quality of life. A recent study has suggested that it can reduce mortality in patients with chronic infection.<sup>97</sup> Tobramycin is also available in a new, more concentrated nebulized formulation (Bramitob<sup>®</sup>, Chiesi) with the same posology (300 mg/4 ml), thereby effectively reducing nebulization time.<sup>98</sup>

A dry powder formulation of tobramycin has recently been approved (TIP<sup>™</sup>, TOBI<sup>®</sup> Podhaler<sup>®</sup>, Novartis). The efficacy and tolerability of the powder form are similar to that of TNS, although a higher incidence of coughing has been reported.<sup>99,100</sup> It is available in 112 mg doses (4/28 mg capsules/12 h), in 28-day on/off cycles.

AZLI (75 mg/8 h in 28-day on/off cycles) has recently been introduced for patients aged 6 years or more (I-A).<sup>58,101</sup> AZLI was

**Table 6**  
Epidemiological Control Strategies in Different Settings for CF Patients With *Pseudomonas aeruginosa* Colonization.

Strategy	Home	Outpatient	Hospital
Follow the general infection control protocols in place in each hospital, and provide an infection control handbook adapted to both the hospital and CF patients.	–	III-A	III-A
Ensure patients, family members and staff follow appropriate rules of hygiene (hand-washing).	III-A	III-A	III-A
Maintain a minimum distance of 2 meters between patients	III-A	III-A	III-A
Do not share equipment used for functional and medical tests between patients (spirometers, stethoscopes, pulse oximeters, etc.)	–	III-B	III-B
Clean surfaces and equipment after seeing each patient	–	III-A	III-A
Use infection control measures (masks, disposable gowns, etc.) when examining patients with multi-drug resistant or highly contagious infection, exacerbations or respiratory symptoms (coughing, sneezing, etc.)	III-A <sup>a</sup>	III-A	III-A
Segregate patients according to source (for example <i>P. aeruginosa</i> , <i>S. aureus</i> , etc.) and type of infection (intermittent, chronic).	–	III-A <sup>a</sup>	III-A <sup>a</sup>
Segregate patients infected by multi-drug resistant or highly contagious strains of <i>P. aeruginosa</i>	III-A	III-A	III-A
Patients should be attended by specific medical and nursing staff according to the type of infection and the presence of multi-drug resistant or highly contagious pathogens.	–	III-C	III-C
Establish protocols for physiotherapists, such as hand-washing, use of gloves, disposable gowns, etc.	III-B <sup>b</sup>	III-B	III-B
Provide each patient with toys, books, etc. for personal use only	III-C <sup>c</sup>	III-C	III-C
Use copper surfaces.	–	III-C <sup>a</sup>	III-C <sup>a</sup>
Provide patients with information on the severity of their infection and how to prevent transmission.	III-C	III-C	III-C
Inform school staff and recommend classroom segregation for patients infected by multi-drug resistant or highly contagious strains	III-C	–	–
Do not attend summer camps, meetings or go on group holidays with other cystic fibrosis patients.	III-B	–	–
Siblings with CF should sleep, use their nebulizers and receive physiotherapy in separate rooms.	III-C	–	–
Clean and disinfect nebulizers according to the manufacturer's instructions.	III-A	III-A	III-A
Do not use saunas, jacuzzis or spas.	III-B	–	–

<sup>a</sup> Evaluate the cost-effectiveness of this recommendation for each setting.

<sup>b</sup> Home care.

<sup>c</sup> Siblings with CF.

shown to be superior to TNS (300 mg/5 ml) for a number of different parameters.<sup>102</sup> Treatment was followed by an open-label extension period in which patients received 3 courses of AZLI. Improvements were comparable to those obtained during the initial 6-month trial.

In conclusion, several different therapeutic options for treating chronic bronchial infection are available, including continuous inhaled colistimethate sodium, or 28-day on/off cycles of TNS or AZLI (Table 5). Evidence that most of the benefits obtained during the treatment phase are not maintained during rest periods<sup>57,58,102</sup> has led to the development of other regimens, including continuous inhaled antibiotics, alternating or rotating regimens, or even shortened cycles (for example, 14-day on/off cycles) (III-A).<sup>103</sup> Combinations have been shown to be effective both in vitro and in animal models<sup>104</sup> and are currently at the clinical trial stage; it is hoped they will limit the development of drug resistance.

Inhaled antibiotics should preferably be administered using the same nebulizers as tested in the clinical trials (III-A). The most common side effect is bronchospasm, so a fast-acting bronchodilator should be administered prior to inhalation. Patients receiving ototoxic or nephrotoxic inhaled antibiotics should be monitored once a year for kidney and auditory function.

### Exacerbations

Exacerbations increase the risk of mortality and are associated with a decline in respiratory function and quality of life. Despite its importance, no definition of exacerbation in CF has yet been agreed.<sup>105,106</sup> It is generally accepted that a patient presenting 2 or more of the signs and symptoms indicated in Table 2 should receive antimicrobial treatment (III-A).<sup>1,105,107</sup> Symptoms can be less evident in infants and preschool children.

Although the choice of empirical treatment is usually based on a sensitivity analysis of the most recent respiratory sample, in vitro sensitivity does not always correlate with response to treatment (III-A). Traditionally, 2 antimicrobials with different mechanisms of action have been used to enhance antibiotic action and restrict the selection of resistant mutants. Guidelines recommend, albeit on the basis of insufficient evidence, the use of dual therapy in severe exacerbations and in patients with more severe pulmonary infection<sup>1,4</sup> (III-C). In mild to moderate exacerbations, treatment should start with oral ciprofloxacin (15–20 mg/kg/12 h, 2–3 weeks) (III-B).<sup>106</sup> In the case of severe exacerbations, or when oral therapy has failed, a combination of an antipseudomonal beta-lactam (piperacillin/tazobactam, ceftazidime, cefepime, aztreonam, imipenem, meropenem or doripenem) and an aminoglycoside (usually tobramycin) or a fluoroquinolone is usually recommended (II-B).<sup>105,106</sup> These drugs should be used in high doses due to their limited ability to penetrate respiratory secretions, and because the renal clearance of some antimicrobials, particularly aminoglycosides, is particularly rapid in CF patients.<sup>60,64,108</sup>

Intravenous colistimethate sodium has also been shown to be effective (III-A)<sup>2,108</sup> although kidney function must be monitored. This antibiotic is usually reserved for multi-drug resistant strains, or when the usual treatment options have failed. There is insufficient evidence to recommend withdrawal or continuation of inhaled antibiotics in patients receiving the same antibiotic intravenously,<sup>4</sup> although the risk of increased toxicity must be considered (III-C). Exacerbations should not be treated solely with inhaled antibiotics (III-D).

There is insufficient evidence to recommend continuous infusion of beta-lactams; therefore, intermittent cycles should be the

**Table 7**  
Keys to Cost-effective Treatment of *Pseudomonas aeruginosa* Colonization in Cystic Fibrosis Patients.

Treatment	Benefits
Aggressive treatment during primary colonization	Prevents chronic infection and limits the high prevalence of chronic <i>P. aeruginosa</i> colonization Reduces the costs associated with chronic infection
Suppressive treatment for chronic infection	Slows progressive pulmonary decline and improves quality of life
Effective treatment for exacerbations	Slows progressive pulmonary decline
Reduces the number of hospitalizations by replacing hospital treatment with other strategies such as home care	Improves quality of life and reduces the risks associated with hospitalization
Change the antibiotic regimen (extended or continuous infusion, dose increase, shorter time between doses, etc.)	Therapy can be adjusted according to PK/PD criteria and therapeutic efficacy
Ensure treatment compliance in patients with <i>P. aeruginosa</i> colonization	Slows progressive pulmonary decline
Measure quality of life	Evaluates treatment given

norm (II-C).<sup>4</sup> In the case of children, once-daily aminoglycosides should be given (B-II) to reduce the risk of kidney damage.<sup>109</sup> There is no evidence to define the dosage regimen in adults. When receiving these drugs, patients must be monitored for serum levels and kidney function (III-A). Patients receiving frequent cycles of aminoglycosides should be monitored for auditory function (III-A).

Although there is insufficient evidence to determine the duration of exacerbation therapy,<sup>110</sup> it should be continued until symptoms resolve and lung function returns to previous levels (III-C). This usually occurs within 2 weeks,<sup>111</sup> except in the case of multi-drug resistant PA or severe pulmonary infection, where prolonged treatment is required.

Home intravenous therapy improves quality of life and can be more cost-effective (III-A); it should be chosen on the basis of the patient's circumstances and the resources available.<sup>112</sup> Chest physiotherapy should be stepped up during exacerbations, and existing chronic therapies should be maintained (III-A).<sup>4</sup>

## Inflammation

The typical initial intense inflammatory response of the airways in CF patients, directed toward limiting infection, is ultimately harmful to the patient.<sup>113</sup> Early administration of anti-inflammatory agents could limit the effects of this reaction and slow pulmonary decline.<sup>114</sup> The strategies used in this case are:

- **Macrolides.** Macrolides modulate cytokine production, inhibit bacterial protein synthesis, attenuate bacterial virulence factors and impair biofilm formation.<sup>115,116</sup> Oral azithromycin is the drug of choice, although studies are needed on its long-term effect, above all in children diagnosed by neonatal screening.<sup>31</sup> It should be used in patients aged over 6 years with PA colonization, and should be given once daily (10 mg/kg in patients weighing <40 kg or 500 mg in patients weighing >40 kg) 3 times/week for at least 4–6 months (I-B).<sup>6,92,117,118</sup> It should not be given in the presence of nontuberculous mycobacteria, as macrolides are part of the treatment of this disease. If nontuberculous mycobacteria are detected during macrolide therapy, administration should be discontinued to avoid induced resistance. All patients should undergo a QT study at the start of treatment, with at least once yearly follow-up studies (III-B).
- **Ibuprofen.** High-dose ibuprofen improves chest radiograph scores and nutritional status, reduces the number of hospitalizations, and significantly improves FEV<sub>1</sub>.<sup>118–121</sup> Although associated with side effects,<sup>121,122</sup> ibuprofen is indicated in children >6 years not receiving nephrotoxic drugs<sup>120</sup> and with an FEV<sub>1</sub>>60%<sup>92</sup> (I-C). There is no evidence for subjects <6 years.<sup>92</sup>
- **Systemic corticosteroids.** These control the action of inflammatory bronchial cells.<sup>123</sup> Long-term (4 years) prednisone every other day (1–2 mg/kg) slows pulmonary decline and improves, albeit temporarily, spirometry values,<sup>123</sup> particularly in children with

mild pulmonary disease.<sup>114,124</sup> It is, however, associated with significant side effects, and the benefits have only been shown to outweigh the risks in allergic bronchopulmonary aspergillosis, intractable bronchospasm, severe small airways disease and terminal disease. Non-enteric-coated tablets are preferable (I-C).<sup>92,114</sup>

- **Inhaled corticosteroids.** No prospective, randomized, placebo-controlled studies have shown these to be beneficial in CF patients, and as such, they should not be routinely used.<sup>92,125</sup> They are only recommended in patients with documented bronchial hyperresponsiveness (I-C). They inhibit the inflammatory action of bronchial epithelial cells and slow FEV<sub>1</sub> decline, although they are associated with side effects.<sup>126–128</sup>

## Epidemiological Control

The importance of delaying the appearance of PA colonization calls for the adoption of strategies to prevent primary infection and disease transmission among CF patients (III-A). The source of PA is largely unknown, although some environmental reservoirs have been identified, particularly warm, damp places. PA is not found in the ocean, as high salt levels would inhibit growth of the bacterium. Showers and swimming pools containing chlorinated water are usually safe, although hydrotherapy pools and jacuzzis can harbor bacteria. PA is often found in hospitals, particularly in ICUs, although the risk of infection is low if appropriate precautions are taken. Nebulizers used by patients with chronic colonization do not usually harbor PA<sup>129</sup> although contaminated healthcare equipment can cause cross-infection. PA is usually transmitted via direct contact, droplets or fomites. Non-mucoid PA can survive for 24 h on inanimate surfaces, and the mucoid strain can survive for 48 h, and even as long as 8 days, in sputum left on a dry surface.<sup>130</sup> To prevent droplet transmission, guidelines recommend maintaining a distance of at least 2 meters between patients, and the use of masks,<sup>19,131</sup> although routine use is not recommended in all guidelines<sup>132</sup> (III-C).

There is evidence of cross-infection in summer camps, meetings, doctors' surgeries and hospitals; therefore, preventive measures should be taken in these settings (Table 6).<sup>129,133–136</sup> New PA colonization, above all multi-drug resistant and highly contagious strains, should be monitored (III-A). PA genotyping should be performed to document transmission and study sources of infections (III-C).

## Pharmacoeconomics

CF is associated with a high consumption of health resources: the longer the survival rate, the greater the cost of the disease.<sup>137</sup> A number of studies have focused on the pharmacoeconomics of CF, evaluating the severity of infection, the long-term nature of the disease, the cost of laboratory studies, healthcare including

hospitalization, and the different therapies required.<sup>137–142</sup> In Spain, direct costs are especially: drugs (45%), visits to the doctor (19%), hospitalization (17%) and laboratory and other studies (12%).<sup>142</sup> Overall, PA infection increases these costs by 40%–55%,<sup>137,141</sup> the highest being associated with chronic patients and women.<sup>141</sup> Given the circumstances, economic analyses of the cost-effectiveness, cost-benefit and cost-usefulness of PA therapies are particularly important (Table 7), although these can be affected by the market price of different over-the-counter drugs.

## Future Outlook

Treatment of PA infection is based on clinical evidence and experience with a small number of antimicrobial drugs. New formulations and antibiotics will soon be available in Spain (colistin dry powder, liposomal amikacin, levofloxacin solution and ciprofloxacin dry powder). These could pave the way for new treatment regimens, including combination therapy, rotation and alternating cycles. The overall aim is to reduce the number and frequency of exacerbations, prevent the development of drug resistant strains, slow the rate of lung function decline, and reduce the need for oral antibiotics during rest periods in on/off cycles. Researchers are currently studying nebulizers that can facilitate administration of these drugs and improve treatment adherence. Other lines of research include the use of anti-inflammatory drugs and their effect on infection, the search for biomarkers, and the study of the cost-benefit ratio of different therapeutic alternatives. Finally, therapies designed to enhance the action of CFTR<sup>143</sup> may change the way in which antimicrobial drugs are used to treat PA colonization in CF patients.

## Funding

The research carried out by R. Cantón, A. Oliver, L. Martínez-Martínez into cystic fibrosis and *P. aeruginosa* was co-financed by the Instituto de Salud Carlos III (Spanish Ministry of the Economy and Competitiveness) (ref. PI12/00734 and PI12/00103) and the European Development Regional Fund “A Way to Achieve Europe,” through the Red Española de Investigación en Patología Infecciosa (REIPI RD12/0015).

## Conflict of Interests

R. Cantón has taken part in training programs sponsored by AstraZeneca, Gilead, Novartis and Praxis Pharmaceutical and has conducted research projects for Astra-Zeneca; L. Maize has taken part in advisory committees and training programs sponsored by Chiesi, Gilead, Novartis, Praxis and Vertex; A. Scribing has taken part in training activities sponsored by Chiesi, Novartis and Praxis and has been a member of Gilead advisory committees; C. Oliveira has taken part in advisory committees and training programs sponsored by Gilead, Novartis, Praxis and Vertex; A. Oliver has conducted research projects sponsored by Gilead; O. Ascension has taken part in training programs sponsored by Chiesi, Gilead, Novartis and Praxis and in advisory committees and research projects funded by GSK and Novartis; S. Gartner has taken part in training programs sponsored by Chiesi, Gilead, Novartis, Praxis and Vertex; E. Roma has taken part in training programs sponsored by Gilead, Novartis and Praxis; E. Quintana-Gallegos has taken part in advisory committees and training programs sponsored by Gilead, Novartis and Praxis; R. Groin has taken part in training programs sponsored by Chiesi, Gilead, Novartis and Praxis; M.I. Barrio has taken part in training programs sponsored by Chiesi, Gilead, Novartis, Praxis and Vertex; M.D. Pastor has taken part in training programs sponsored by Gilead; C. Prados has taken part in training

programs sponsored by Gilead, Novartis and Praxis; J. Barber has taken part in training programs sponsored by AstraZeneca and Gilead; L. Martínez-Martínez has taken part in training programs and research projects sponsored by Astra-Zeneca, GSK, Janssen-Cilag, MSD and Pfizer; C. Vázquez has taken part in training programs sponsored by Gilead, Novartis, Praxis and Vertex; J. de Garcia has taken part in advisory committees and training programs sponsored by Chiesi, Gilead, Novartis and Praxis; A. Sole has taken part in training programs sponsored by Gilead, Novartis, Vertex and Praxis. A. Salved, M.T. Martinez-Martinez, J.J. Castón and J.L. Poveda have no conflicts of interest.

## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.arbr.2014.09.018](https://doi.org/10.1016/j.arbr.2014.09.018).

## References

- Cantón R, Cobos N, de Gracia J, Baquero F, Honorato J, Gartner S, et al., Spanish Consensus Group for Antimicrobial Therapy in the Cystic Fibrosis Patient. Antimicrobial therapy for pulmonary pathogenic colonisation and infection by *Pseudomonas aeruginosa* in cystic fibrosis patients. *Clin Microbiol Infect*. 2005;11:690–703.
- UK Cystic Fibrosis Trust. Antibiotic treatment for cystic fibrosis. Report of the UK Cystic Fibrosis Trust Antibiotic Working Group; 2009. Available from: <https://www.cysticfibrosis.org.uk/media/82010/CD.Antibiotic.treatment.for.CF.May.09.pdf>
- Döring G, Flume P, Heijerman H, Elborn JS, for the Consensus Study Group. Treatment of lung infection in patients with cystic fibrosis: current and future strategies. *J Cyst Fibros*. 2012;11:461–79.
- Flume PA, Mogayzel PJ Jr, Robinson KA, Goss CH, Rosenblatt RL, Kuhn RJ, et al., Clinical Practice Guidelines for Pulmonary Therapies Committee. Cystic fibrosis pulmonary guidelines: treatment of pulmonary exacerbations. *Am J Respir Crit Care Med*. 2009;180:802–8.
- Barrio Gómez de Agüero MI, García Hernández G, Gartner S, y Grupo de Trabajo de Fibrosis Quística. Protocolo de diagnóstico y seguimiento de los pacientes con fibrosis quística. *An Pediatr (Barc)*. 2009;71:250–64.
- Mogayzel PJ Jr, Naureckas ET, Robinson KA, Mueller G, Hadjilidiadis D, Hoag JB, et al., Pulmonary Clinical Practice Guidelines Committee. Cystic fibrosis pulmonary guidelines. Chronic medications for maintenance of lung health. *Am J Respir Crit Care Med*. 2013;187:680–9.
- Gartner S, Cobos N. Cribado neonatal para la fibrosis quística. *An Pediatr (Barc)*. 2009;71:481–2.
- Plosker GL. Aztreonam lysine for inhalation solution in cystic fibrosis. *Drugs*. 2010;70:1843–55.
- Máiz L, Girón RM, Olveira C, Quintana E, Lamas A, Pastor D, et al. Inhaled antibiotics for the treatment of chronic bronchopulmonary *Pseudomonas aeruginosa* infection in cystic fibrosis: systematic review of randomized controlled trials. *Expert Opin Pharmacother*. 2013;14:1135–49.
- Ryan G, Singh M, Dwan K. Inhaled antibiotics for long-term therapy in cystic fibrosis. *Cochrane Database Syst Rev*. 2011;3:CD001021.
- Hauser AR, Jain M, Bar-Meir M, McColley SA. Clinical significance of microbial infection and adaptation in cystic fibrosis. *Clin Microbiol Rev*. 2011;24:29–70.
- Ciofu O, Hansen CR, Høiby N. Respiratory bacterial infections in cystic fibrosis. *Curr Opin Pulm Med*. 2013;19:251–8.
- Folkesson A, Jelsbak L, Yang L, Johansen HK, Ciofu O, Høiby N, et al. Adaptation of *Pseudomonas aeruginosa* to the cystic fibrosis airway: an evolutionary perspective. *Nat Rev Microbiol*. 2012;10:841–51.
- Burns JL, Rolain JM. Culture-based diagnostic microbiology in cystic fibrosis: can we simplify the complexity? *J Cyst Fibros*. 2014;13:1–9.
- Oliver A, Alarcón T, Caballero E, Cantón R. Diagnóstico microbiológico de la colonización-infección broncopulmonar en el paciente con fibrosis quística. *Enferm Infecc Microbiol Clin*. 2009;27:89–94.
- Prescott WA, Nagel JL. Extended-interval once-daily dosing of aminoglycosides in adult and pediatric patients with cystic fibrosis. *Pharmacotherapy*. 2010;30:95–108.
- Stockmann C, Sherwin CM, Zobel JT, Young DC, Waters CD, Spigarelli MG, et al. Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: III. Fluoroquinolones. *Pediatr Pulmonol*. 2013;48:211–20.
- Young DC, Zobel JT, Waters CD, Ampofo K, Stockmann C, Sherwin CM, et al. Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: IV. Colistimethate sodium. *Pediatr Pulmonol*. 2013;48:1–7.
- Saiman L. Infection prevention and control in cystic fibrosis. *Curr Opin Infect Dis*. 2011;24:390–5.
- Gross PA, Barrett TL, Dellinger P, Krause PJ, Martone WJ, McGowan JE Jr, et al. Purpose of quality standards for Infectious Diseases Infectious Diseases Society of America. *Clin Infect Dis*. 1994;18:421.



21. Troxler RB, Hoover WC, Britton LJ, Gerwin AM, Rowe SM. Clearance of initial mucoid *Pseudomonas aeruginosa* in patients with CF. *Pediatr Pulmonol*. 2012;47:1113–22.
22. Dalboge CS, Pressler T, Hoiby N, Nielsen JG, Krogh Johansen H. A cohort study of the Copenhagen CF-Centre eradication strategy against *Staphylococcus aureus*. *J Cyst Fibros*. 2013;12:42–8.
23. Sly PD, Gangel CL, Chen L, Ware RS, Ranganathan S, Mott LS, et al. AREST CF Investigators. Risk factors for bronchiectasis in children with cystic fibrosis. *N Engl J Med*. 2013;368:1963–70.
24. Staffer P, Davies JC, Balfour-Lynn IM, Rosenthal M, Bush A. Bronchoscopy in cystic fibrosis infants diagnosed by newborn screening. *Pediatr Pulmonol*. 2011;46:696–700.
25. Ratjen F, Walter H, Haug M, Meisner C, Grassemann H, Döring G. Diagnostic value of serum antibodies in early *Pseudomonas aeruginosa* infection in cystic fibrosis patients. *Pediatr Pulmonol*. 2007;42:249–55.
26. Anstead M, Heltshe SL, Khan U, Barbieri JT, Langkamp M, Döring G, et al. *Pseudomonas aeruginosa* serology and risk for re-isolation in the EPIC trial. *J Cyst Fibros*. 2013;12:147–53.
27. Deschagh P, van Daele S, de Baets F, Vanechoutte M. PCR and the detection of *Pseudomonas aeruginosa* in respiratory samples of CF patients. A literature review. *J Cyst Fibros*. 2011;10:293–7.
28. Goeminne PC, Vandendriessche T, van Eldere J, Nicolai BM, Hertog ML, Dupont LJ. Detection of *Pseudomonas aeruginosa* in sputum headspace through volatile organic compound analysis. *Respir Res*. 2012;13:87.
29. Rosenfeld M, Emerson J, McNamara S, Joubbran K, Retsch-Bogart G, Graff GR, et al. EPIC Study Group Participating Clinical Sites. Baseline characteristics and factors associated with nutritional and pulmonary status at enrollment in the cystic fibrosis EPIC observational cohort. *Pediatr Pulmonol*. 2010;45:934–44.
30. Cystic Fibrosis Foundation. Cystic Fibrosis Foundation Patient Registry 2011. Annual Data Report. Bethesda, MD: Cystic Fibrosis Foundation; 2013.
31. McKay KO, Cooper PJ, van Asperen PP. Segregation of children with CF diagnosed via newborn screening and acquisition of *Pseudomonas aeruginosa*. *J Cyst Fibros*. 2009;8:400–4.
32. Emerson J, Rosenfeld M, McNamara S, Ramsey B, Gibson RL. *Pseudomonas aeruginosa* and other predictors of mortality and morbidity in young children with cystic fibrosis. *Pediatr Pulmonol*. 2002;34:91–100.
33. Ren CL, Rosenfeld M, Mayer OH, Davis SD, Kloster M, Castile RG, et al. Analysis of the associations between lung function and clinical features in preschool children with cystic fibrosis. *Pediatr Pulmonol*. 2012;47:574–81.
34. Taylor-Robinson D, Whitehead M, Diderichsen F, Olesen HV, Pressler T, Smyth RL, et al. Understanding the natural progression in %FEV1 decline in patients with cystic fibrosis: a longitudinal study. *Thorax*. 2013;68:294–5.
35. Vendrell M, de Gracia J, Oliveira C, Martínez MA, Girón R, Máz L, et al. Diagnosis and treatment of bronchiectasis. *Arch Bronconeumol*. 2008;44:629–40.
36. Li Z, Kosorok MR, Farrell PM, Lavova A, West SE, Green CG, et al. Longitudinal development of mucoid *Pseudomonas aeruginosa* infection and lung disease progression in children with cystic fibrosis. *JAMA*. 2005;293:581–8.
37. Ferkol T, Rosenfeld M, Milla CE. Cystic fibrosis pulmonary exacerbations. *J Pediatr*. 2006;148:259–64.
38. Regelmann WE, Schechter MS, Wagener JS, Morgan WJ, Pasta DJ, Elkin EP, et al. Pulmonary exacerbations in cystic fibrosis: young children with characteristic signs and symptoms. *Pediatr Pulmonol*. 2013;48:649–57.
39. Belesis Y, Dixon B, Hawkins G, Pereira J, Peat J, MacDonald R, et al. Early cystic fibrosis lung disease detected by bronchoalveolar lavage and lung clearance index. *Am J Respir Crit Care Med*. 2012;185:862–73.
40. Aurora P. Multiple-breath inert gas washout test and early cystic fibrosis lung disease. *Thorax*. 2010;65:373–4.
41. Owens CM, Aurora P, Stanojevic S, Bush A, Wade A, Oliver C, et al. Lung clearance index and HRCT are complementary markers of lung abnormalities in young children with CF. *Thorax*. 2011;66:481–8.
42. Taccetti G, Campana S, Festini F, Mascherini M, Döring G. Early eradication therapy against *Pseudomonas aeruginosa* in cystic fibrosis patients. *Eur Respir J*. 2005;26:1–4.
43. Linnane B, Robinson P, Ranganathan S, Stick S, Murray C. Role of high-resolution computed tomography in the detection of early CF lung disease. *Paediatr Respir Rev*. 2008;9:168–74.
44. Young C, Owens C. 'To CT or not to CT? That is the question': outcome surrogates for surveillance in childhood cystic fibrosis. *Thorax*. 2012;67:471–2.
45. Tiddens HA, de Jong PA. Update on the application of chest computed tomography scanning to cystic fibrosis. *Curr Opin Pulm Med*. 2006;12:433–43.
46. De Jong PA, Tiddens HA, Lequin MH, Robinson TE, Brody AS. Estimation of the radiation dose from CT in cystic fibrosis. *Chest*. 2008;133:1289–91.
47. Mott LS, Park J, Murray CP, Gangel CL, de Klerk NH, Robinson PJ, et al. Progression of early structural lung disease in young children with cystic fibrosis assessed using computed tomography. *Thorax*. 2012;67:509–16.
48. Robinson TE, Leung AN, Chen X, Moss RB, Emond MJ. Cystic fibrosis HRCT scores correlate strongly with *Pseudomonas* infection. *Pediatr Pulmonol*. 2009;44:1107–17.
49. Folescu TW, Marques Ede A, Boechat MC, Daltro P, Higa LY, Cohen RW. High-resolution computed tomography scores in cystic fibrosis patients colonized with *Pseudomonas aeruginosa* or *Staphylococcus aureus*. *J Bras Pneumol*. 2012;38:41–9.
50. Loeve M, Gerbrands K, Hop WC, Rosenfeld M, Hartmann IC, Tiddens HA. Bronchiectasis and pulmonary exacerbations in children and young adults with cystic fibrosis. *Chest*. 2011;140:178–85.
51. European Medicines Agency. Report of the workshop on endpoints for cystic fibrosis clinical trials. 2012. Available from: [http://www.ema.europa.eu/docs/en\\_GB/document\\_library/Report/2012/12/WC500036159.pdf](http://www.ema.europa.eu/docs/en_GB/document_library/Report/2012/12/WC500036159.pdf)
52. Aanaes K. Bacterial sinusitis as a focus for initial lung colonisation and chronic lung infection in patients with cystic fibrosis. *J Cyst Fibros*. 2013;12:1–20.
53. Lee TWR, Brownlee KG, Conway SP, Denton M, Littlewood JM. Evaluation of a new definition for chronic *Pseudomonas aeruginosa* infection in cystic fibrosis patients. *J Cyst Fibros*. 2003;2:29–34.
54. Proesmans M, Balinska-Miskiewicz W, Dupont L, Bossuyt X, Verhaegen J, Hoiby N, et al. Evaluating the 'Leeds criteria' for *Pseudomonas aeruginosa* infection in a cystic fibrosis centre. *Eur Respir J*. 2006;27:937–43.
55. Langton Hewer SC, Smyth AR. Antibiotic strategies for eradicating *Pseudomonas aeruginosa* in people with cystic fibrosis. *Cochrane Database Syst Rev*. 2009:CD004197.
56. Littlewood KJ, Higashi K, Jansen JP, Capkun-Niggli G, Balp MM, Doering G, et al. A network meta-analysis of the efficacy of inhaled antibiotics for chronic *Pseudomonas* infections in cystic fibrosis. *J Cyst Fibros*. 2012;11:419–26.
57. Ramsey BW, Pepe MS, Quan JM, Otto KL, Montgomery AB, Williams-Warren J, et al. Interim administration of inhaled tobramycin in patients with cystic fibrosis. Cystic Fibrosis Inhaled Tobramycin Study Group. *N Engl J Med*. 1999;340:23–30.
58. McCoy KS, Quittner AL, Oermann CM, Gibson RL, Retsch-Bogart GZ, Montgomery AB. Inhaled aztreonam lysine for chronic airway *Pseudomonas aeruginosa* in cystic fibrosis. *Am J Respir Crit Care Med*. 2008;178:921–8.
59. Touw DJ, Vinks AA, Mouton JW, Horrevorts AM. Pharmacokinetic optimisation of antibacterial treatment in patients with cystic fibrosis Current practice and suggestions for future directions. *Clin Pharmacokinet*. 1998;35:437–59.
60. Zobell JT, Young DC, Waters CD, Stockmann C, Ampofo K, Sherwin CM, et al. Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: I. Aztreonam and carbapenems. *Pediatr Pulmonol*. 2012;47:1147–58.
61. Zobell JT, Young DC, Waters CD, Ampofo K, Stockmann C, Sherwin CM, et al. Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: VI. Executive summary. *Pediatr Pulmonol*. 2013;48:525–37.
62. Touw DJ. Clinical pharmacokinetics of antimicrobial drugs in cystic fibrosis. *Pharm World Sci*. 1998;20:149–60.
63. Rey E, Treluyer JM, Pons G. Drug disposition in cystic fibrosis. *Clin Pharmacokinet*. 1998;35:313–29.
64. Young DC, Zobell JT, Stockmann C, Waters CD, Ampofo K, Sherwin CM, et al. Optimization of anti-pseudomonal antibiotics for cystic fibrosis pulmonary exacerbations: V. Aminoglycosides. *Pediatr Pulmonol*. 2013;48:1047–61.
65. Butterfield JM, Lodise TP, Beegle S, Rosen J, Farkas J, Pai MP. Pharmacokinetics and pharmacodynamics of once-daily administration of intravenous tobramycin in adult patients with cystic fibrosis hospitalized for an acute pulmonary exacerbation. *Antimicrob Agents Chemother*. 2013;57:5175–7.
66. Geller DE, Pitlick WH, Nardella PA, Tracewell WG, Ramsey BW. Pharmacokinetics and bioavailability of aerosolized tobramycin in cystic fibrosis. *Chest*. 2002;122:219–26.
67. Macià MD, Pérez JL, Molin S, Oliver A. Dynamics of mutator and antibiotic-resistant populations in a pharmacokinetic/pharmacodynamic model of *Pseudomonas aeruginosa* biofilm treatment. *Antimicrob Agents Chemother*. 2011;55:5230–7.
68. Manduru M, Mihm LB, White RL, Friedrich LV, Flume PA, Bosso JA. *In vitro* pharmacodynamics of ceftazidime against *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. *Antimicrob Agents Chemother*. 1997;41:2053–6.
69. Ratjen F, Rietschel E, Kasel D, Schwiertz R, Starke K, Beier H, et al. Pharmacokinetics of inhaled colistin in patients with cystic fibrosis. *J Antimicrob Chemother*. 2006;57:306–11.
70. Li J, Coulthard K, Milne R, Nation RL, Conway S, Peckham D, et al. Steady-state pharmacokinetics of intravenous colistin methanesulphonate in patients with cystic fibrosis. *J Antimicrob Chemother*. 2003;52:987–92.
71. Dudhani RV, Turnidge JD, Coulthard K, Milne RW, Rayner CR, Li J, et al. Elucidation of the pharmacokinetic/pharmacodynamic determinant of colistin activity against *Pseudomonas aeruginosa* in murine thigh and lung infection models. *Antimicrob Agents Chemother*. 2010;54:1117–24.
72. Schuster A, Halilburn C, Döring G, Goldman MH, for the Freedom Study Group. Safety, efficacy and convenience of colistimethate sodium dry powder for inhalation (Colobreathe DPI) in patients with cystic fibrosis: a randomised study. *Thorax*. 2013;68:344–50.
73. Trämper-Stranders GA, Wolfs TF, van Haren Noman S, van Aalderen WM, Nagelkerke AF, Nuijsink M, et al. Controlled trial of cycled antibiotic prophylaxis to prevent initial *Pseudomonas aeruginosa* infection in children with cystic fibrosis. *Thorax*. 2010;65:915–20.
74. Johansen HK, Göttsche PC. Vaccines for preventing infection with *Pseudomonas aeruginosa* in cystic fibrosis. *Cochrane Database Syst Rev*. 2013;6:CD001399.
75. Vázquez C, Elorz J, Baranda F, Sojo A, Pijoan JI. Early treatment of bronchial colonization with *Pseudomonas aeruginosa* in cystic fibrosis. Twelve-year experience. *J Cyst Fibros*. 2002;1 Suppl. 1:51.
76. Vázquez C, Municio M, Corera M, Gaztelurrutia L, Sojo A, Vitoria JC. Early treatment of *Pseudomonas aeruginosa* colonization in cystic fibrosis. *Acta Paediatr*. 1993;82:308–9.
77. Wiesemann HG, Steinkamp G, Ratjen F, Bauernfeind A, Przyklenk B, Döring G, et al. Placebo-controlled, double-blind, randomized study of aerosolized tobramycin for early treatment of *Pseudomonas aeruginosa* colonization in cystic fibrosis. *Pediatr Pulmonol*. 1998;25:88–92.

78. Gibson RL, Emerson J, McNamara S, Burns JL, Rosenfeld M, Yunker A, et al. Significant microbiological effect of inhaled tobramycin in young children with cystic fibrosis. *Am J Respir Crit Care Med.* 2003;167:841–9.
79. Taccetti G, Bianchini E, Cariani L, Buzzetti R, Costantini D, Trevisan F, et al. Early antibiotic treatment for *Pseudomonas aeruginosa* eradication in patients with cystic fibrosis: a randomised multicentre study comparing two different protocols. *Thorax.* 2012;67:853–9.
80. Ratjen F, Munck A, Kho P, Angyalosi G, ELITE Study Group. Treatment of early *Pseudomonas aeruginosa* infection in patients with cystic fibrosis: the ELITE trial. *Thorax.* 2010;65:286–91.
81. Stick S, Tiddens H, Aurora P, Gustafsson P, Ranganathan S, Robinson P, et al. Early intervention studies in infants and preschool children with cystic fibrosis: are we ready? *Eur Respir J.* 2013;42:527–38.
82. Schelstraete P, Haerynck F, van Daele S, Deseyne S, de Baets F. Eradication therapy for *Pseudomonas aeruginosa* colonization episodes in cystic fibrosis patients not chronically colonized by *P. aeruginosa*. *J Cyst Fibros.* 2013;12:1–8.
83. Frederiksen B, Koch C, Hoiby N. Antibiotic treatment of initial colonization with *Pseudomonas aeruginosa* postpones chronic infection and prevents deterioration of pulmonary function in cystic fibrosis. *Ped Pulmonol.* 1997;23:330–5.
84. Hansen CR, Pressler T, Høiby N. Early aggressive eradication therapy for intermittent *Pseudomonas aeruginosa* airway colonization in cystic fibrosis patients: 15 years experience. *J Cyst Fibros.* 2008;7:523–30.
85. Treggiari MM, Retsch-Bogart G, Mayer-Hamblett N, Khan U, Kulich M, Kronmal R, et al. Comparative efficacy and safety of 4 randomized regimens to treat early *Pseudomonas aeruginosa* infection in children with cystic fibrosis. *Arch Pediatr Adolesc Med.* 2011;165:847–56.
86. Gibson RL, Emerson J, Mayer-Hamblett N, Burns JL, McNamara S, Accurso FJ, et al. Duration of treatment effect after tobramycin solution for inhalation in young children with cystic fibrosis. *Pediatr Pulmonol.* 2007;42:610–23.
87. Tiddens HAW, de Boeck K, Clancy JP, Fayon M, Arets HGM, Bresnik M, et al., for the ALPINE Study Investigators. Open-label study of inhaled aztreonam for *Pseudomonas* eradication in children with cystic fibrosis: the ALPINE Study. *J Cyst Fibros.* 2015;14:111–9.
88. Proesmans M, Vermeulen F, Boulanger L, Verhaegen J, de Boeck K. Comparison of two treatment regimens for eradication of *Pseudomonas aeruginosa* infection in children with cystic fibrosis. *J Cyst Fibros.* 2013;12:29–34.
89. Fauvart M, de Groote VN, Michiels J. Role of persister cells in chronic infections: clinical relevance and perspectives on anti-persister therapies. *J Med Microbiol.* 2011;60:699–709.
90. Levy H, Kalish LA, Cannon CL, García KC, Gerard C, Goldmann D, et al. Predictors of mucoid *Pseudomonas* colonization in cystic fibrosis patients. *Pediatr Pulmonol.* 2008;43:463–71.
91. Tramper-Stranders GA, van der Ent CK, Molin S, Yang L, Hansen SK, Rau MH, et al. Initial *Pseudomonas aeruginosa* infection in patients with cystic fibrosis: characteristics of eradicated and persistent isolates. *Clin Microbiol Infect.* 2012;18:567–74.
92. Flume PA, O'Sullivan BP, Robinson KA, Goss CH, Mogayzel PJ Jr, Willey-Courand DB, et al. Cystic fibrosis pulmonary guidelines: chronic medications for maintenance of lung health. *Am J Respir Crit Care Med.* 2007;176:957–69.
93. Heijerman H, Westerman E, Conway S, Touw D, Döring G, Consensus Working Group. Inhaled medication and inhalation devices for lung disease in patients with cystic fibrosis: a European consensus. *J Cyst Fibros.* 2009;8:295–315.
94. Jensen T, Pedersen SS, Garne S, Heilmann C, Høiby N, Koch C. Colistin inhalation therapy in cystic fibrosis patients with chronic *Pseudomonas aeruginosa* lung infection. *J Antimicrob Chemother.* 1987;19:831–8.
95. Hodson ME, Gallagher CG, Govan JR. A randomised clinical trial of nebulised tobramycin or colistin in cystic fibrosis. *Eur Respir J.* 2002;20:658–64.
96. Murphy TD, Anbar RD, Lester LA, Nasr SZ, Nickerson B, van Devanter DR, et al. Treatment with tobramycin solution for inhalation reduces hospitalizations in young CF subjects with mild lung disease. *Pediatr Pulmonol.* 2004;38:314–20.
97. Sawicki GS, Signorovitch JE, Zhang J, Latremouille-Viau D, von Wartburg M, Wu EQ, et al. Reduced mortality in cystic fibrosis patients treated with tobramycin inhalation solution. *Pediatr Pulmonol.* 2012;47:44–52.
98. Chuchalin A, Csizsér E, Gyurkovics K, Bartnicka MT, Sands D, Kapranov N, et al. A formulation of aerosolized tobramycin (Bramitob) in the treatment of patients with cystic fibrosis and *Pseudomonas aeruginosa* infection: a double-blind, placebo-controlled, multicenter study. *Paediatr Drugs.* 2007;9:21–31.
99. Konstan MW, Geller DE, Minić P, Brockhaus F, Zhang J, Angyalosi G. Tobramycin inhalation powder for *P. aeruginosa* infection in cystic fibrosis: the EVOLVE trial. *Pediatr Pulmonol.* 2011;46:230–8.
100. Konstan MW, Flume PA, Kappler M, Chiron R, Higgins M, Brockhaus F, et al. Safety, efficacy and convenience of tobramycin inhalation powder in cystic fibrosis patients: the EAGER trial. *J Cyst Fibros.* 2011;10:54–61.
101. Oermann CM, Retsch-Bogart GZ, Quittner AL, Gibson RL, McCoy KS, Montgomery AB, et al. An 18-month study of the safety and efficacy of repeated courses of inhaled aztreonam lysine in cystic fibrosis. *Pediatr Pulmonol.* 2010;45:1121–34.
102. Assael BM, Pressler T, Bilton D, Fayon M, Fischer R, Chiron R, et al. Inhaled aztreonam lysine vs inhaled tobramycin in cystic fibrosis: a comparative efficacy trial. *J Cyst Fibros.* 2012;12:130–40.
103. Lo D, van Devanter DR, Flume P, Smyth A. Aerosolized antibiotic therapy for chronic cystic fibrosis airway infections: continuous or intermittent? *Respir Med.* 2011;105 Suppl. 2:S9–17.
104. Herrmann G, Yang L, Wu H, Song Z, Wang H, Høiby N, et al. Colistin-tobramycin combinations are superior to monotherapy concerning the killing of biofilm *Pseudomonas aeruginosa*. *J Infect Dis.* 2010;202:1585–92.
105. Bilton D, Canny G, Conway S, Dumcius S, Hjelte L, Proesmans M, et al. Pulmonary exacerbation: towards a definition for use in clinical trials. Report from the EuroCareCF Working Group on outcome parameters in clinical trials. *J Cyst Fibros.* 2011;10 Suppl. 2:S79–81.
106. Kraynack NC, Gothard MD, Falletta LM, McBride JT. Approach to treating cystic fibrosis pulmonary exacerbations varies widely across US CF care centers. *Pediatr Pulmonol.* 2011;46:870–81.
107. Sanders DB, Hoffman LR, Emerson J, Gibson RL, Rosenfeld M, Redding GJ, et al. Return of FEV1 after pulmonary exacerbation in children with cystic fibrosis. *Pediatr Pulmonol.* 2010;45:127–34.
108. Conway SP, Pond MN, Watson A, Etherington C, Robey HL, Goldman MH. Intravenous colistin sulphomethate in acute respiratory exacerbations in adult patients with cystic fibrosis. *Thorax.* 1997;52:987–93.
109. Smyth AR, Bhatt J. Once-daily versus multiple-daily dosing with intravenous aminoglycosides for cystic fibrosis. *Cochrane Database Syst Rev.* 2014;2:CD002009.
110. Plummer A, Wildman M. Duration of intravenous antibiotic therapy in people with cystic fibrosis. *Cochrane Database Syst Rev.* 2013:CD006682.
111. Collaco JM, Green DM, Cutting GR, Naughton KM, Mogayzel PJ Jr. Location and duration of treatment of cystic fibrosis respiratory exacerbations do not affect outcomes. *Am J Respir Crit Care Med.* 2010;182:1137–43.
112. Balaguer A, González de Dios J. Home versus hospital intravenous antibiotic therapy for cystic fibrosis. *Cochrane Database Syst Rev.* 2012:CD001917.
113. Chmiel JF, Konstan MW. Inflammation and anti-inflammatory therapies for cystic fibrosis. *Clin Chest Med.* 2007;28:331.
114. Pressler T. Targeting airway inflammation in cystic fibrosis in children: past, present, and future. *Paediatr Drugs.* 2011;13:141–7.
115. Saiman L, Marshall BC, Mayer-Hamblett N, Burns JL, Quittner AL, Cibene DA, et al. Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA.* 2003;290:1749–56.
116. Altenburg J, de Graaff CS, van der Werf TS, Boersma WG. Immunomodulatory effects of macrolide antibiotics—Part 1: Biological mechanisms. *Respiration.* 2011;81:67–74.
117. Wilms EB, Touw DJ, Heijerman HG, van der Ent CK. Azithromycin maintenance therapy in patients with cystic fibrosis: a dose advice based on a review of pharmacokinetics, efficacy, and side effects. *Pediatr Pulmonol.* 2012;47:658–65.
118. Royal Brompton Hospital. Clinical Guidelines: care of Children with Cystic Fibrosis. 5th ed; 2011. Available from: <http://www.rbht.nhs.uk/healthprofessionals/clinical-departments/paediatrics/childrencf/>
119. Konstan MW, Byard PJ, Hoppel CL, Davis PB. Effect of high-dose ibuprofen in patients with cystic fibrosis. *N Engl J Med.* 1995;323:848–54.
120. Lands LC, Milner R, Cantin AM, Manson D, Corey M. High-dose ibuprofen in cystic fibrosis: Canadian safety and effectiveness trial. *J Pediatr.* 2007;151:249–54.
121. Konstan MW, Schluchter MD, Xue W, Davis PB. Clinical use of ibuprofen is associated with slower FEV1 decline in children with cystic fibrosis. *Am J Respir Crit Care Med.* 2007;176:1084–9.
122. Konstan MW. Ibuprofen therapy for cystic fibrosis lung disease: revisited. *Curr Opin Pulm Med.* 2008;14:567.
123. Cheng K, Ashby D, Smyth RL. Oral steroids for long-term use in cystic fibrosis. *Cochrane Database Syst Rev.* 2011;10:CD000407.
124. Kieninger E, Regamey N. Targeting inflammation in cystic fibrosis. *Respiration.* 2010;79:189–90.
125. Balfour-Lynn IM, Welch K. Inhaled corticosteroids for cystic fibrosis. *Cochrane Database Syst Rev.* 2012;11:CD001915.
126. Rebeyrol C, Saint-Criq V, Guillot L, Riffault L, Corvol H, Chadelat K, et al. Glucocorticoids reduce inflammation in cystic fibrosis bronchial epithelial cells. *Cell Signal.* 2012;24:1093–9.
127. De Boeck K, Vermeulen F, Wanyama S, Thomas M, Members of the Belgian CF Registry. Inhaled corticosteroids and lower lung function decline in young children with CF. *Eur Respir J.* 2011;37:1091–5.
128. Ren CL, Pasta DJ, Rasouliyan L, Wagener JS, Konstan MW, Morgan WJ, et al. Relationship between inhaled corticosteroid therapy and rate of lung function decline in children with CF. *J Pediatr.* 2008;153:746–51.
129. UK Cystic Fibrosis Trust. Infection Control Group *Pseudomonas aeruginosa* infection in people with cystic fibrosis. Suggestions for prevention and infection control. 2nd ed; 2004. Available from: <http://www.cftrust.org.uk/aboutcf/publications/consensusdoc/>
130. O'Malley CA. Infection control in cystic fibrosis: cohorting, cross-contamination, and the respiratory therapist. *Respir Care.* 2009;54:641–57.
131. Saiman L, Siegel JD, LiPuma JJ, Brown RF, Bryson EA, Chambers MJ, et al. Infection prevention and control guideline for cystic fibrosis: 2013 update. *Infect Control Hosp Epidemiol.* 2014;35 Suppl. 1:S1–67.
132. Zhou J, Garber E, Saiman L. Survey of infection control policies for patients with cystic fibrosis in the United States. *Am J Infect Control.* 2008;36:220–2.
133. Siegel JD, Rhinehart E, Jackson M, Chiarello L, Health Care Infection Control Practices Advisory Committee. Guideline for isolation precautions: preventing transmission of infectious agents in health care settings. *Am J Infect Control.* 2007;35 Suppl. 2:S65–164.
134. Zuckerman JB, Zuaro DE, Prato BS, Ruoff KL, Sawicki RW, Quinton HB, et al. Bacterial contamination of cystic fibrosis clinics. *J Cyst Fibros.* 2009;8:186–92.

135. Casey AL, Adams D, Karpanen TJ, Lambert PA, Cookson BD, Nightingale P, et al. Role of copper in reducing hospital environment contamination. *J Hosp Infect.* 2010;74:72–7.
136. Mikolay A, Huggett S, Tikana L, Grass G, Braun J, Nies DH. Survival of bacteria on metallic copper surfaces in a hospital trial. *Appl Microbiol Biotechnol.* 2010;87:1875–9.
137. Sansgiry SS, Joish VN, Boklage S, Goyal RK, Chopra P, Sethi S. Economic burden of *Pseudomonas aeruginosa* infection in patients with cystic fibrosis. *J Med Econom.* 2012;15:219–24.
138. Schreyögg J, Hollmeyer H, Bluemel M, Staab D, Busse R. Hospitalisation costs of cystic fibrosis. *Pharmacoeconomics.* 2006;24:999–1009.
139. Braccini G, Festini F, Boni V, Neri AS, Galici V, Campana S, et al. The costs of treatment of early and chronic *Pseudomonas aeruginosa* infection in cystic fibrosis patients. *J Chemother.* 2009;21:188–92.
140. Heimeshoff M, Hollmeyer H, Schreyo J, Tiemann O, Staab D. Cost of illness of cystic fibrosis in Germany. Results from a Large Cystic Fibrosis Centre. *Pharmacoeconomics.* 2012;30:763–77.
141. DeWitt EM, Grussemeyer CA, Friedman JY, Dinan MA, Lin L, Schulman KA, et al. Resource use, costs and utility estimates for patients with cystic fibrosis with mild impairment in lung function: analysis of data collected alongside a 48-week multicenter clinical trial. *Value Health.* 2012;15:277–83.
142. López Bastida J, Linertová R, Serrano Aguilar P, Hens Pérez M, Posada de la Paz M, Oliva Moreno J. Los costes socioeconómicos y la calidad de vida relacionada con la salud en pacientes con enfermedades raras en España. Proyecto del IMSERSO N.º 167/10; 2012, March. Available from: [http://www.fqmadrid.org/Noticias/costes\\_socioeconomicos/Costes\\_socioeconomicos.ER.2012.pdf](http://www.fqmadrid.org/Noticias/costes_socioeconomicos/Costes_socioeconomicos.ER.2012.pdf)
143. Ramsey BW, Davies J, McElvaney NG, Tullis E, Bell SC, Dřevínek P, et al. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med.* 2011;365:1663–72.