



Review

Animal Models of Chronic Obstructive Pulmonary Disease[☆]

Sandra Pérez-Rial, Álvaro Girón-Martínez, Germán Peces-Barba*

Laboratorio de Neumología, Instituto de Investigación Sanitaria-Fundación Jiménez Díaz-CIBERES-UAM, Madrid, Spain

ARTICLE INFO

Article history:

Received 20 March 2014

Accepted 25 June 2014

Available online 1 February 2015

Keywords:

Chronic obstructive pulmonary disease

Animal model

Smoking

Emphysema

Transgenic animals

Exacerbation

Autoimmune

Therapeutic assays

ABSTRACT

Animal models of disease have always been welcomed by the scientific community because they provide an approach to the investigation of certain aspects of the disease in question.

Animal models of COPD cannot reproduce the heterogeneity of the disease and usually only manage to represent the disease in its milder stages. Moreover, airflow obstruction, the variable that determines patient diagnosis, not always taken into account in the models. For this reason, models have focused on the development of emphysema, easily detectable by lung morphometry, and have disregarded other components of the disease, such as airway injury or associated vascular changes.

Continuous, long-term exposure to cigarette smoke is considered the main risk factor for this disease, justifying the fact that the cigarette smoke exposure model is the most widely used. Some variations on this basic model, related to exposure time, the association of other inducers or inhibitors, exacerbations or the use of transgenic animals to facilitate the identification of pathogenic pathways have been developed. Some variations or heterogeneity of this disease, then, can be reproduced and models can be designed for resolving researchers' questions on disease identification or treatment responses.

© 2014 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

Modelos animales de enfermedad pulmonar obstructiva crónica

RESUMEN

El desarrollo de modelos animales de una enfermedad ha sido siempre bien acogido por la comunidad científica porque permite realizar una aproximación a la investigación de determinados aspectos de la misma.

Los modelos animales de la EPOC no pueden llegar a reproducir la heterogeneidad de esta enfermedad y generalmente solo llegan a representar los estadios más leves de la misma. Además, la obstrucción al flujo aéreo, variable que determina el diagnóstico en un paciente, no siempre se tiene en cuenta en los modelos. Por este motivo, los modelos se han centrado en el desarrollo de enfisema, fácilmente detectable por morfometría pulmonar, sin prestar atención a otros componentes de la enfermedad, como la lesión de las vías aéreas o las alteraciones vasculares asociadas.

La exposición continua y prolongada al humo de tabaco se considera el principal factor de riesgo de esta enfermedad, lo que justifica que sea el modelo de exposición al humo de tabaco el más ampliamente utilizado. Sobre esta base de modelo podemos encontrar algunas variantes relacionadas con el tiempo de exposición, la asociación de otros inductores o inhibidores, las exacerbaciones o el uso de animales transgénicos que facilitan la identificación de las vías patogénicas. Es posible, por tanto, reproducir algunas variantes o heterogeneidades de esta enfermedad y diseñar uno u otro modelo que sea capaz de responder a una u otra pregunta de investigación, dirigida bien a una identificación patogénica y/o bien a una respuesta terapéutica.

© 2014 SEPAR. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Palabras clave:

Enfermedad pulmonar obstructiva crónica

Modelo animal

Tabaco

Enfisema

Transgénico

Exacerbación

Autoinmune

Ensayos terapéuticos

[☆] Please cite this article as: Pérez-Rial S, Girón-Martínez Á, Peces-Barba G. Modelos animales de enfermedad pulmonar obstructiva crónica. Arch Bronconeumol. 2015;51:121–127.

* Corresponding author.

E-mail address: gpeces@fjd.es (G. Peces-Barba).

Introduction

Chronic obstructive pulmonary disease (COPD) has a huge global impact, and clinicians must make use of all the tools available to tackle the many aspects of this disease. The development of animal models can help address problems such as under-diagnosis, frail exacerbator patients, and existing uncertainties about the development of one or other clinical form of the disease or its natural history (in which cases with minor and accelerated disease progression are mixed). Similarly, all new therapeutic trials are generally based on an earlier study in an animal model.

Smoking is the leading cause of COPD, but its ability to generate a permanent inflammatory response depends on the patient's susceptibility. For this reason, animal models of COPD developed by exposure to cigarette smoke are primarily chosen to study the pathogenic mechanisms of the disease and of susceptibility to development and progression. In these cases, the use of transgenic animals, in which a particular metabolic pathway is inhibited or activated, helps researchers understand the pathogenic pathways that exist in each case. Likewise, new approaches to the classification of COPD proposed by both the GOLD initiative¹ and Spanish COPD guidelines (GesEPOC)² place greater emphasis on exacerbations due to their effect on the severity of symptoms, progression of the obstruction, and mortality. For this reason, interest has grown in the study of exacerbations in models of COPD, and the results may help to improve knowledge of the mechanisms underlying the condition in the exacerbator patient.

When evaluating the results of studies in animal models of any disease, the limitation of having to extrapolate a conclusion as to what is potentially present in a patient must always be taken into consideration. However, they are an essential part of clinical research when used as "preclinical models", an increasingly widespread term that encompasses the notion of translation into clinical practice that must form the basis of any study design.

Models of Cigarette Smoke-Induced COPD

Models of cigarette smoke-induced COPD are those that best reflect the inflammatory and pathogenic mechanisms of the disease and, consequently, those that are potentially better suited to testing new therapies. Exposure to cigarette smoke has been applied in numerous animal species, such as dogs, guinea pigs, rabbits, rats, and mice. Of these, guinea pigs and mice have proven to be most susceptible to the development of COPD through prolonged exposure.³ There are two general procedures for administering cigarette smoke: the so-called "nose only" method, where the smoke is channeled directly into the animal's nose, and "whole body" administration (Fig. 1), where the animal is placed in a chamber filled with a controlled concentration of smoke to

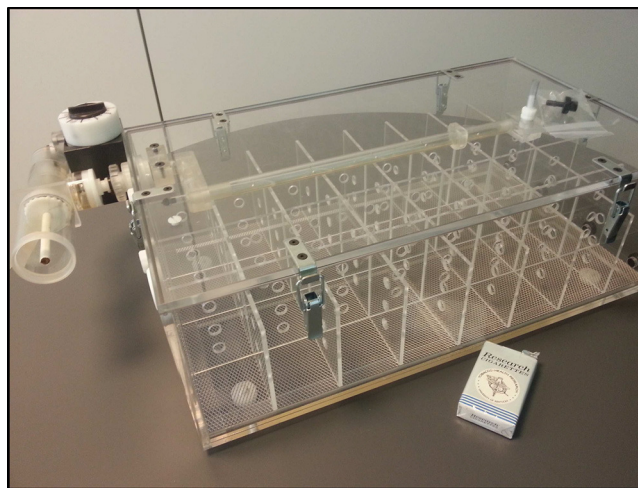


Fig. 1. Whole body tobacco exposure system.

ensure complete exposure to stable, non-toxic carboxyhemoglobin levels.^{4,5} Conceptually different, both methods have been widely used, and have shown similar findings as regards the presence of inflammatory cell populations, cytokine levels, changes in lung remodeling, and therapeutic response⁶ (Fig. 2).

In approximately 90% of patients, COPD is caused by smoking an average of at least 10 pack-years, and they develop a disease that can take different clinical forms, with different levels of progression and severity.⁷ Animal models of COPD, guinea pig or murine are usually established over a 6-month exposure period,⁸ although major inflammatory and morphometric changes can already be detected after the second month.⁹ They do not usually reach the stage equivalent to severe COPD in a patient, but they can develop many of the characteristics typical of this disease, such as chronic inflammation with increased neutrophil and macrophage counts, presence of CD4 and CD8T lymphocytes, mucus hypersecretion, changes in lung function, emphysema, and vascular and airway remodeling.¹⁰

The murine model of cigarette smoke exposure is the most widely used, due to its low cost and easy management, well-mapped genome, the availability of many transgenic variants, a wide range of specific antibodies for laboratory use, and a large number of strains with differing susceptibilities to cigarette smoke. Strain-dependent susceptibility for developing COPD is well identified in the murine model,^{11–14} and the pulmonary morphometric pattern of COPD can be generated when mice are exposed to cigarette smoke for at least 3 to 6 months, with typical inflammatory cells, inflammatory mediators and functional changes characteristic of the disease.¹⁵

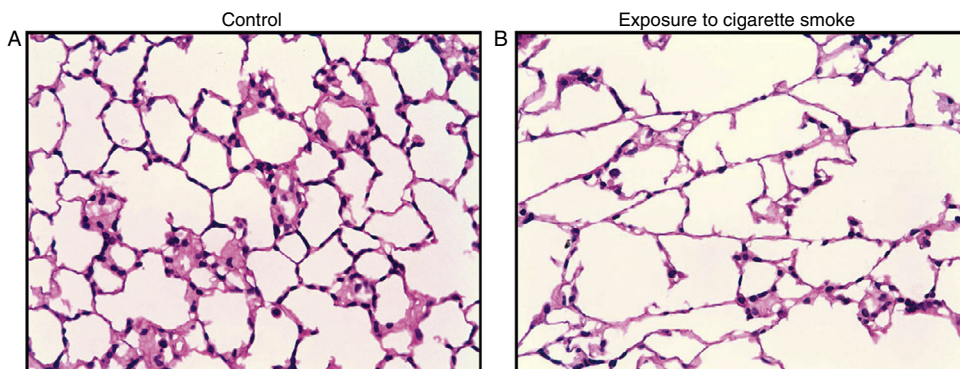


Fig. 2. Histological sections (H&E) of the lungs of mice exposed to ambient air (A) and mice exposed to tobacco smoke for 6 months (B), showing emphysema.

Table 1
Examples of Cigarette Smoke-Induced COPD.

Animal model	Methodology	Finding	Reference
Serpin B1 KO mice	Chronic exposure	Not associated with severe emphysema	83
NZWLac/J, AJ, SJL, C57BL/6, and AKR mice	Chronic exposure	The degree of susceptibility to developing lung injury is strain-dependent	12
NZWLac/J and AKR mice	Chronic exposure	Egr-1 (proinflammatory marker) greatly increased in the susceptible AKR strain, but far less so in the NZWLac/J strain	84
C57BL/6 (CD8 ^{-/-}) mice	Chronic exposure	CD8+ T lymphocytes are essential for the development of emphysema	85
C57BL/6 mice	Acute exposure	Increased markers of DNA damage by oxidative stress (8-OHdG and 4-HNE)	86
ICR (Nrf2 ^{-/-}) mice	Chronic exposure	More susceptible to developing emphysema, with increased antioxidant enzyme expression	78
Guinea pig	Chronic exposure	Increase in vascular remodeling and arterial pulmonary pressure	87
C57BL/6 and "pallid" mice	Chronic exposure	The α 1-antitrypsin levels determine the emphysematous profile	88
Guinea pig	Chronic exposure	Role of MMPs in the development of emphysema and airway remodeling	89
C57BL/6 mice	Acute exposure (direct and indirect)	The intensity of the inflammatory response depends on the composition of the cigarette smoke	90
C57BL/6 mice (sGC α 1 ^{-/-})	Acute and chronic exposure	Reduction in sGC expression contributing to airflow limitation	91
Mice	Sub-chronic exposure	Anti-inflammatory effect of PPAR- γ ligands	92
Mice	Chronic exposure	The neutralization of CXCL13 partially blocks the inflammatory response and alveolar wall destruction	93
Mice	Chronic exposure	Blocking T lymphocytes may be effective as therapy for COPD	94
C57BL/6 and BALB/cj mice	Chronic exposure	Cigarette smoke triggers an antigen-dependent response in which CD4+ and CD8+ lymphocytes participate	95
C57BL/6 and 129S2/SvHsd mice	Acute exposure	Implication of the pro-inflammatory monocytes in susceptibility to developing lung injury	5

Guinea pigs are also a good choice for generating models of COPD, as these animals are very susceptible to developing the disease after only a few months of exposure.¹⁶ In 1990, Wright and Churg published one of the first studies in guinea pigs exposed to cigarette smoke.¹⁷ In this case, after 12 months of exposure the guinea pigs developed emphysema and presented lung function changes very similar to those found in smokers with COPD. The main difficulty in the use of this species comes from the limited availability of specific antibodies. Rats, the species of rodent closest to mice and guinea pigs, are rarely used as models of COPD because they are more resistant to developing changes due to cigarette smoke exposure.¹⁸ Table 1 summarizes some relevant findings described in this model.

Models of COPD due to chronic cigarette smoke exposure continue to be limited insofar as they are unable to reproduce some of the characteristics of this complex, heterogeneous disease. So far, attempts to develop known clinical phenotypes, such as the exacerbator or accelerated progression types, or forms with bacterial colonization, for example, have failed. Some studies, however, have attempted to address these questions by combining agents. Exposure to toxic and irritant gases such as nitrogen dioxide, ozone or sulfur dioxide causes more severe lung damage than cigarette smoke.^{19–22}

Models of COPD Exacerbations

Exacerbations are a characteristic of COPD that, if repeated, determine the poor clinical course of the patient, as they are associated with greater disease progression, poorer quality of life and higher risk of mortality. The availability of animal models of exacerbation gives researchers the chance to study associated pathogenic mechanisms and detect possible associated biological markers.

Most infectious COPD exacerbations are viral in origin (75%), while the remainder are bacterial. Studies in *in vivo* models have demonstrated the effect of viral infection on mice previously exposed, both short- and long-term, to cigarette smoke. Inflammation of the lung is more severe if the viral infection affects an

animal previously exposed to cigarette smoke, and also accelerates emphysema progression and the severity of airway damage.^{23,24}

The most commonly isolated bacterium in COPD exacerbations is nontypeable *Haemophilus influenzae* (NTHI). For this reason, the results obtained in models of this infection in healthy rats²⁵ vs. those previously exposed to cigarette smoke are particularly interesting. After C57BL/6 mice had been exposed to cigarette smoke for 8 weeks, NTHI infection caused a more severe inflammatory response and greater lung damage than in previously healthy animals.^{26,27}

Bacterial lipopolysaccharides (LPS) have been used alone, in long-term administration,²⁸ or in combination with short periods of exposure to cigarette smoke²⁹ to develop models of emphysema. However, single massive insult can cause an inflammatory response that is accompanied by fever, mucus hypersecretion and bronchoconstriction, which reproduces symptoms of an exacerbation³⁰ seen on computed tomography.³¹

Models of Severe COPD by Combining Induction Agents

In the more severe stages of COPD a clear breakdown of the lung "maintenance program" occurs that can inevitably lead to emphysema and pulmonary hypertension. There are various models of "frail" (very severe) COPD pathology, such as the combination of cigarette smoke exposure and vascular endothelial growth factor (VEGF) inhibitor.³² Exposure to cigarette smoke causes a significant decrease in VEGF and VEGF receptor-2 (VEGFR-2) expression in animal models of emphysema. Furthermore, treatment with the VEGF receptor blocker SU5416 induces alveolar cell apoptosis, capillary retraction, and alveolar space enlargement.³³ For this reason, emphysema presents as a VEGF deficiency that compromises the survival of the endothelial cells and consequently the lung's maintenance program. Other results can be achieved by combining cigarette smoke exposure and hypoxia induction. This model can lead to pulmonary hypertension, a condition present only in advanced severe COPD.³⁴

Table 2
Strains of Natural Mutant Mice That Develop Emphysema.

Mouse	Mutation	Pulmonary phenotype	Reference
Blotchy (<i>Blo</i>)	Abnormal translation in the Menkes gene on the X chromosome	Air space enlargement Disorganization of the elastic fibers	35,36 37,38
Tight skin (<i>Tsk+/-</i>)	Duplication of fibrillin-1	Abnormal air space development, with development of panlobular emphysema	39
Beige (<i>Bg</i>)	Deletion in <i>Lyst</i>	Normal at birth, but with abnormal neonatal alveolization	40,41
Pallid (<i>Pa</i>)	Syntaxin-3	Develop mild, late onset emphysema	42
Osteopetrotic (<i>Op</i>)	Macrophage colony-stimulating factor deficiency	Reduced number of alveolar macrophages and emphysema	

Transgenic Models of COPD

Before the advent of targeted genetic engineering, some mutant strains of C57BL/6 mice that spontaneously developed emphysema had appeared (Table 2). These were the “blotchy” mice that have an abnormal translation of the *Menkes gene* on the X chromosome,³⁵ causing defects in lung connective tissue proteins, which affects the structure and function of the lungs, causing emphysema³⁶; “Tight Skin” mice, with a mutation in *fibrillin-1*, one of the key components of the microfibrils in the lung extracellular matrix, which causes oxidative stress and cell death, injury cascades central to the development of emphysema^{37,38}; “Beige” mice, in which the lungs appear normal at birth but, due to the deletion of *Lyst*, do not form alveoli normally during development³⁹; “Pallid” mice,⁴⁰ with a mutation that affects syntaxin-13 (a cell membrane protein), resulting in the gradual and progressive development of emphysema⁴¹; and more recently, “Osteopetrotic” mice, which are macrophage colony-stimulating factor-deficient and eventually develop emphysema.⁴² One of the major technological breakthroughs of the last few decades has been the development of transgenic animals. These are animals in which a gene that does not form part of their genome, and which will sequence a certain pathway of interest, is inserted by intranuclear injection in the early embryonic phases.^{13,15,43–45} One of the first applications of transgenic technology to COPD was the constitutive overexpression of human collagenase-1 (MMP-1) in mice, which causes emphysema⁴⁶ by degradation of type III collagen in the alveolar walls.⁴⁷ The constitutive expression of transgenes, however, does not distinguish the lung’s own development process. To overcome this, the transgene expression construct was developed. Thus, overexpression of IL-13,⁴⁸ a cytokine produced by T-helper type 2 (Th2) lymphocytes, or overexpression of IFN- γ ,⁴⁹ the main product of T-helper type 1 (Th1) lymphocytes, are two important examples of inducible conditional transgenes. In the case of IL-13 transgenic mice (“Dutch”), this leads to MMP-9 and MMP-12-dependent emphysema in adult mice.⁵⁰ In these animals, IL-13 is overexpressed only when they are exposed to tetracycline, thereby allowing investigators to activate overexpression after the lung is fully developed. MMP-9-mediated activation of TGF- β appears to be responsible for collagen

remodeling in this model. However, in IFN- γ transgenic mice (“British”), the inflammatory component appears to be more subtle, with prominent apoptosis but no associated airways disease. These are only two examples that demonstrate the complexity of inflammatory networks, and how unexpected findings in animal models have led to the search for new mediators in human disease. Other studies show how TNF- α induction in the adult lung facilitates the formation of lymphoid tissue and emphysema, providing a model for research into the pathogenic effects of TNF- α in the lung,⁵¹ or how prothymosin- α (ProT- α) expression contributes to the pathogenesis of emphysema by increasing acetylation of histones and expression of NF- κ B-dependent MMP-2 and MMP-9, especially after cigarette smoke exposure.⁵²

An alternative to the transgenic model is the “knockout” model (Table 3), in which the expression of a certain gene is inhibited, thereby enabling the function of proteins dependent on this gene to be determined. Sometimes, gene inhibition protects against the development of emphysema, as in the case of inhibition of MMP-12 expression, which impairs alveolar macrophage recruitment and thereby protects against the development of emphysema.⁵³ Another example is the absence of neutrophil elastase (NE), which also appears to protect against the development of smoke-induced emphysema.⁵⁴ In both cases, the direct role of these proteins in emphysema has been demonstrated, highlighting the interdependence of the proteinases and inflammatory cells that mediate lung destruction in response to cigarette smoke. At other times, the deletion interferes with alveogenesis. In this case, platelet-derived growth factor A (PDGF-A)-deficient mice develop emphysema due to loss of myofibroblasts and the associated elastin fiber deposits.⁵⁵ Double knockout mice for fibroblast growth factor receptors 3 and 4 (FGFR-3 and -4) have abnormal alveolar formation and septation,⁵⁶ while elastin-deficient mice have fewer dilated distal air sacs and arrested airway development.⁵⁷ In other cases, deletion of certain genes causes alveolar space enlargement. This is true of integrin α V β 6-deficient mice, in which TGF- β activation in alveolar air spaces does not occur, leading to development of MMP-12-dependent emphysema.⁵⁸ Other examples are knockout mice for pulmonary surfactant protein D (SP-D), which present macrophage activation, production of MMPs and air space enlargement,⁵⁹ or

Table 3
Examples of “Knockout” Models of Emphysematous Mice.

Gene	Pulmonary phenotype	Reference
Macrophage elastase (MMP-12)	Complete protection against the development of emphysema after exposure to cigarette smoke	53
Neutrophil elastase (NE)	Protection against cigarette smoke-induced emphysema	54
Elastin	Fewer dilated distal air sacs and arrested airway development	57
Platelet-derived growth factor A (PDGF-A)	Lack of tropoelastin and failure in alveolar septation	55
Fibroblast growth factor receptor (FGFR-3 and -4)	Abnormal alveolar formation and septation	56
Integrin α V β 6	Spontaneous development of MMP-12-dependent emphysema	58
Pulmonary surfactant protein D (SP-D)	Macrophage activation, production of MMPs and subsequent emphysema	59
Tissue inhibitor of metalloprotease-3 (TIMP-3)	Air space enlargement with gradual development of emphysema	60

tissue inhibitor of metalloproteinase-3 (TIMP-3) deficiency, which appears to combine air space enlargement with the gradual development of emphysema.⁶⁰

To overcome the cross-species barrier in the murine model, certain murine genes can be eliminated and human genes inserted ("knocked in") under the control of murine promoters. Emphysema-prone mice, in which the murine alpha-1 antitrypsin (A1AT) genes have been removed and replaced by normal or deficient human A1AT genes, have also been developed.⁶¹

Autoimmune Models of COPD

Pulmonary inflammation in severe COPD involves a large number of activated Th1T lymphocytes, B lymphocytes and CD8 lymphocytes, which persist for years, even after smoking cessation; this is consistent with a self-perpetuating process, which is one of the characteristics of autoimmune diseases. This chain of events suggests that the adaptive immune response in COPD, together with its persistence after smoking cessation, could be due to a response to autoantigens. Initially, this was merely a hypothesis,^{62–64} but since then new evidence, including the development of the first animal model of autoimmune emphysema,^{66–68} would seem to confirm the suggestion.⁶⁵ The presence of anti-elastin autoantibodies⁶⁹ and other autoantigens^{70,71} has been correlated with emphysema severity, and induction of autoantibodies against lung matrix proteins has been shown to increase the smoke-induced immune response in mice previously immunized with a mixture of lung extracellular matrix proteins.⁷²

Models for Therapeutic Trials in COPD

Current treatments do little to inhibit chronic inflammation, do not reverse COPD pathology, and do not modify the factors that initiate and lead to disease progression in the long term. It is clear, therefore, that new therapies that can prevent COPD induction and progression must be developed, and this is only possible through animal models that accurately reflect the physiopathology of the disease. Many anti-COPD drugs in clinical development have been identified from studies in animal models. Various inhibitors of inflammatory mediators are being developed for the treatment of COPD, although to date the results of tests using LTB₄, TNF- α , IL-1, IL-8, and EGF inhibitors have been disappointing.⁷³ Studies in animals exposed to cigarette smoke and treated with synthetic neutrophil elastase inhibitors have shown their potential anti-inflammatory activity.⁷⁴ Similarly, findings in animal models of cigarette smoke-induced airway inflammation support the potential therapeutic usefulness of kinase inhibitors (p38 MAPK and PI3K) in COPD.⁷⁵ The antioxidant enzyme Gpx-1 protects against lung inflammation and cigarette smoke-induced emphysema in mice,⁷⁶ and a Gpx mimetic also reduced lung inflammation when administered both prophylactically and therapeutically.^{76,77} In other studies, deletion of the Nrf2 antioxidant stress response gene led to increased lung inflammation and emphysema in mice exposed to cigarette smoke,⁷⁸ and an Nrf2 activator is currently undergoing clinical trials for COPD.⁷³

New COPD drugs that can reduce the rate of pulmonary destruction and airflow limitation, and even arrest or reverse the underlying processes have yet to be discovered. In this regard, some evidence suggests that retinoic acid significantly slows elastase-induced emphysema in rats,⁷⁹ and this has sparked interest in the retinoids⁸⁰ and other growth factors as potential therapeutic agents,⁸¹ although how these can be used has yet to be confirmed. Studies published by our group show that liver growth factor (LGF) has pulmonary anti-fibrotic activity, and can improve lung function

and partially revert the deposit of matrix proteins following CdCl₂ administration in a model of fibrosis in rats.⁸²

In conclusion, studies in animal models of COPD continue to contribute important information. These valuable tools further our understanding of the pathogenic aspects of the disease and help develop therapeutic clinical trials. The inherent heterogeneity of the disease can also be reflected in animal models developed by using different combinations or doses of induction agents. This is why it is important to choose the model according to whether the research is focused on pathogenesis, diagnosis or treatment.

Conflict of Interests

The authors declare that they have no conflict of interests.

References

- Lopez-Giraldo A, Rodriguez-Roisin R, Agusti A. Chronic obstructive pulmonary disease: the golden decade. Implications for the diagnosis, prevention and treatment of chronic obstructive pulmonary disease. *Med Clin (Barc)*. 2014. <http://dx.doi.org/10.1016/j.medcli.2014.03.009>. pii:S0025-7753(14)00226-7 [Epub ahead of print].
- Miravittles M, Soler-Cataluna JJ, Calle M, Molina J, Almagro P, Quintano JA, et al. Spanish guideline for COPD (GesEPOC). *Arch Bronconeumol*. 2014;50:1–16.
- Peces-Barba G, Heili S. Modelos animales de EPOC. *Arch Bronconeumol*. 2007;43:30–7.
- Perez-Rial S, Del Puerto-Navado L, Gonzalez-Mangado N, Peces-Barba G. Early detection of susceptibility to acute lung inflammation by molecular imaging in mice exposed to cigarette smoke. *Mol Imaging*. 2011;10:398–405.
- Perez-Rial S, del Puerto-Navado L, Terron-Exposito R, Giron-Martinez A, Gonzalez-Mangado N, Peces-Barba G. Role of recently migrated monocytes in cigarette smoke-induced lung inflammation in different strain of mice. *PLOS ONE*. 2013;8:e72975.
- Leberl M, Kratzer A, Taraseviciene-Stewart L. Tobacco smoke induced COPD/emphysema in the animal model-are we all on the same page? *Front Physiol*. 2013;4:91.
- Miravittles M, Soler-Cataluna JJ, Calle M, Molina J, Almagro P, Antonio Quintano J, et al. Spanish Guideline for COPD (GesEPOC). *Arch Bronconeumol*. 2014;50:1–16.
- Wright JL, Cosio M, Churg A. Animal models of chronic obstructive pulmonary disease. *Am J Physiol Lung Cell Mol Physiol*. 2008;295:L1–15.
- van der Strate BW, Postma DS, Brandsma CA, Melgert BN, Luinge MA, Geerlings M, et al. Cigarette smoke-induced emphysema: a role for the B cell? *Am J Respir Crit Care Med*. 2006;173:751–8.
- Vlahos R, Bozinovski S. Recent advances in pre-clinical mouse models of COPD. *Clin Sci (Lond)*. 2014;126:253–65.
- March TH, Barr EB, Finch GL, Hahn FF, Hobbs CH, Menache MG, et al. Cigarette smoke exposure produces more evidence of emphysema in B6C3F1 mice than in F344 rats. *Toxicol Sci*. 1999;51:289–99.
- Guerassimov A, Hoshino Y, Takubo Y, Turcotte A, Yamamoto M, Ghezzi H, et al. The development of emphysema in cigarette smoke-exposed mice is strain dependent. *Am J Respir Crit Care Med*. 2004;170:974–80.
- Brusselle GG, Bracke KR, Maes T, D'Hulst AI, Moerloose KB, Joos GF, et al. Murine models of COPD. *Pulm Pharmacol Ther*. 2006;19:155–65.
- Bartalesi B, Cavarra E, Fineschi S, Lucattelli M, Lungchi B, Martorana PA, et al. Different lung responses to cigarette smoke in two strains of mice sensitive to oxidants. *Eur Respir J*. 2005;25:15–22.
- Shapiro SD. Animal models for chronic obstructive pulmonary disease: age of klotho and marlboro mice. *Am J Respir Cell Mol Biol*. 2000;22:4–7.
- Wright JL, Churg A. A model of tobacco smoke-induced airflow obstruction in the guinea pig. *Chest*. 2002;121:188S–91S.
- Wright JL, Churg A. Cigarette smoke causes physiologic and morphologic changes of emphysema in the guinea pig. *Am Rev Respir Dis*. 1990;142:1422–8.
- Stevenson CS, Docx C, Webster R, Battram C, Hynx D, Giddings J, et al. Comprehensive gene expression profiling of rat lung reveals distinct acute and chronic responses to cigarette smoke inhalation. *Am J Physiol Lung Cell Mol Physiol*. 2007;293:L1183–93.
- Farone A, Huang S, Paulauskis J, Kobzik L. Airway neutrophilia and chemokine mRNA expression in sulfur dioxide-induced bronchitis. *Am J Respir Cell Mol Biol*. 1995;12:345–50.
- Shore S, Kobzik L, Long NC, Skornik W, Van Staden CJ, Boulet L, et al. Increased airway responsiveness to inhaled methacholine in a rat model of chronic bronchitis. *Am J Respir Crit Care Med*. 1995;151:1931–8.
- Chitano P, Hosselet JJ, Mapp CE, Fabbri LM. Effect of oxidant air pollutants on the respiratory system: insights from experimental animal research. *Eur Respir J*. 1995;8:1357–71.
- Wegmann M, Renz H, Herz U. Long-term NO₂ exposure induces pulmonary inflammation and progressive development of airflow obstruction in C57BL/6

- mice: a mouse model for chronic obstructive pulmonary disease? *Pathobiology*. 2002;70:284–6.
23. Gualano RC, Hansen MJ, Vlahos R, Jones JE, Park-Jones RA, Deliyannis G, et al. Cigarette smoke worsens lung inflammation and impairs resolution of influenza infection in mice. *Respir Res*. 2008;9:53.
 24. Kang MJ, Lee CG, Lee JY, Dela Cruz CS, Chen ZJ, Enelow R, et al. Cigarette smoke selectively enhances viral PAMP- and virus-induced pulmonary innate immune and remodeling responses in mice. *J Clin Invest*. 2008;118:2771–84.
 25. Morey P, Viadas C, Euba B, Hood DW, Barberan M, Gil C, et al. Relative contributions of lipooligosaccharide inner and outer core modifications to nontypeable *Haemophilus influenzae* pathogenesis. *Infect Immun*. 2014;81:4100–11.
 26. Gaschler GJ, Skrtic M, Zavitz CC, Lindahl M, Onnervik PO, Murphy TF, et al. Bacteria challenge in smoke-exposed mice exacerbates inflammation and skews the inflammatory profile. *Am J Respir Crit Care Med*. 2009;179:666–75.
 27. Gaschler GJ, Zavitz CC, Bauer CM, Stampfli MR. Mechanisms of clearance of nontypeable *Haemophilus influenzae* from cigarette smoke-exposed mouse lungs. *Eur Respir J*. 2010;36:1131–42.
 28. Stolk J, Rudolphus A, Davies P, Osinga D, Dijkman JH, Agarwal L, et al. Induction of emphysema and bronchial mucus cell hyperplasia by intratracheal instillation of lipopolysaccharide in the hamster. *J Pathol*. 1992;167:349–56.
 29. Nie YC, Wu H, Li PB, Luo YL, Zhang CC, Shen JG, et al. Characteristic comparison of three rat models induced by cigarette smoke or combined with LPS: to establish a suitable model for study of airway mucus hypersecretion in chronic obstructive pulmonary disease. *Pulm Pharmacol Ther*. 2012;25:349–56.
 30. Spond J, Billah MM, Chapman RW, Egan RW, Hey JA, House A, et al. The role of neutrophils in LPS-induced changes in pulmonary function in conscious rats. *Pulm Pharmacol Ther*. 2004;17:133–40.
 31. Kobayashi S, Fujinawa R, Ota F, Angata T, Ueno M, Maeno T, et al. A single dose of lipopolysaccharide into mice with emphysema mimics human chronic obstructive pulmonary disease exacerbation as assessed by micro-computed tomography. *Am J Respir Cell Mol Biol*. 2014;49:971–7.
 32. Edirisinghe I, Yang SR, Yao H, Rajendrasozhan S, Caito S, Adenuga D, et al. VEGFR-2 inhibition augments cigarette smoke-induced oxidative stress and inflammatory responses leading to endothelial dysfunction. *FASEB J*. 2008;22:2297–310.
 33. Kasahara Y, Tudor RM, Taraseviciene-Stewart L, Le Cras TD, Abman S, Hirth PK, et al. Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. *J Clin Invest*. 2000;106:1311–9.
 34. Olea E, Ferrer E, Prieto-Lloret J, Gonzalez-Martin C, Vega-Agapito V, Gonzalez-Obeso E, et al. Effects of cigarette smoke and chronic hypoxia on airways remodeling and resistance. Clinical significance. *Respir Physiol Neurobiol*. 2014;179:305–13.
 35. Mercer JF, Grimes A, Ambrosini L, Lockhart P, Paynter JA, Dierick H, et al. Mutations in the murine homologue of the Menkes gene in dappled and blotchy mice. *Nat Genet*. 1994;6:374–8.
 36. Fisk DE, Kuhn C. Emphysema-like changes in the lungs of the blotchy mouse. *Am Rev Respir Dis*. 1976;113:787–97.
 37. Kieley CM, Raghunath M, Siracusa LD, Sherratt MJ, Peters R, Shuttleworth CA, et al. The Tight skin mouse: demonstration of mutant fibrillin-1 production and assembly into abnormal microfibrils. *J Cell Biol*. 1998;140:1159–66.
 38. Podowski M, Calvi CL, Cheadle C, Tudor RM, Biswal S, Neptune ER. Complex integration of matrix, oxidative stress, and apoptosis in genetic emphysema. *Am J Pathol*. 2009;175:84–96.
 39. Starcher B, Williams I. The beige mouse: role of neutrophil elastase in the development of pulmonary emphysema. *Exp Lung Res*. 1989;15:785–800.
 40. Martorana PA, Brand T, Gardi C, van Even P, de Santi MM, Calzoni P, et al. The pallid mouse. A model of genetic alpha 1-antitrypsin deficiency. *Lab Invest*. 1993;68:233–41.
 41. Huang L, Kuo YM, Gitschier J. The pallid gene encodes a novel, syntaxin 13-interacting protein involved in platelet storage pool deficiency. *Nat Genet*. 1999;23:329–32.
 42. Shibata Y, Zsengeller Z, Otake K, Palaniyar N, Trapnell BC. Alveolar macrophage deficiency in osteopetrotic mice deficient in macrophage colony-stimulating factor is spontaneously corrected with age and associated with matrix metalloproteinase expression and emphysema. *Blood*. 2001;98:2845–52.
 43. Mahadeva R, Shapiro SD. Chronic obstructive pulmonary disease * 3: experimental animal models of pulmonary emphysema. *Thorax*. 2002;57:908–14.
 44. Tudor RM, McGrath S, Neptune E. The pathobiological mechanisms of emphysema models: what do they have in common? *Pulm Pharmacol Ther*. 2003;16:67–78.
 45. Vlahos R, Bozinovski S, Gualano RC, Ernst M, Anderson GP. Modelling COPD in mice. *Pulm Pharmacol Ther*. 2006;19:12–7.
 46. D'Armiento J, Dalal SS, Okada Y, Berg RA, Chada K. Collagenase expression in the lungs of transgenic mice causes pulmonary emphysema. *Cell*. 1992;71:955–61.
 47. Shiomi T, Okada Y, Foronjy R, Schiltz J, Jaenish R, Krane S, et al. Emphysematous changes are caused by degradation of type III collagen in transgenic mice expressing MMP-1. *Exp Lung Res*. 2003;29:1–15.
 48. Zheng T, Zhu Z, Wang Z, Homer RJ, Ma B, Riese RJ Jr, et al. Inducible targeting of IL-13 to the adult lung causes matrix metalloproteinase- and cathepsin-dependent emphysema. *J Clin Invest*. 2000;106:1081–93.
 49. Wang Z, Zheng T, Zhu Z, Homer RJ, Riese RJ, Chapman HA Jr, et al. Interferon gamma induction of pulmonary emphysema in the adult murine lung. *J Exp Med*. 2000;192:1587–600.
 50. Elias JA, Kang MJ, Crothers K, Homer R, Lee CG. State of the art. Mechanistic heterogeneity in chronic obstructive pulmonary disease: insights from transgenic mice. *Proc Am Thorac Soc*. 2006;3:494–8.
 51. Vuilleminot BR, Rodriguez JF, Hoyle GW. Lymphoid tissue and emphysema in the lungs of transgenic mice inducibly expressing tumor necrosis factor-alpha. *Am J Respir Cell Mol Biol*. 2004;30:438–48.
 52. Su BH, Tseng YL, Shieh GS, Chen YC, Shiang YC, Wu P, et al. Prothymosin alpha overexpression contributes to the development of pulmonary emphysema. *Nat Commun*. 2013;4:1906.
 53. Hautamaki RD, Kobayashi DK, Senior RM, Shapiro SD. Requirement for macrophage elastase for cigarette smoke-induced emphysema in mice. *Science*. 1997;277:2002–4.
 54. Shapiro SD, Goldstein NM, Houghton AM, Kobayashi DK, Kelley D, Belaouaj A. Neutrophil elastase contributes to cigarette smoke-induced emphysema in mice. *Am J Pathol*. 2003;163:2329–35.
 55. Bostrom H, Willetts K, Pekny M, Leveen P, Lindahl P, Hedstrand H, et al. PDGF-A signaling is a critical event in lung alveolar myofibroblast development and alveogenesis. *Cell*. 1996;85:863–73.
 56. Weinstein M, Xu X, Ohyama K, Deng CX. FGFR-3 and FGFR-4 function cooperatively to direct alveogenesis in the murine lung. *Development*. 1998;125:3615–23.
 57. Wendel DP, Taylor DG, Albertine KH, Keating MT, Li DY. Impaired distal airway development in mice lacking elastin. *Am J Respir Cell Mol Biol*. 2000;23:320–6.
 58. Morris DG, Huang X, Kaminski N, Wang Y, Shapiro SD, Dolganov G, et al. Loss of integrin alpha(v)beta6-mediated TGF-beta activation causes Mmp12-dependent emphysema. *Nature*. 2003;422:169–73.
 59. Wert SE, Yoshida M, LeVine AM, Ikegami M, Jones T, Ross GF, et al. Increased metalloproteinase activity, oxidant production, and emphysema in surfactant protein D gene-inactivated mice. *Proc Natl Acad Sci U S A*. 2000;97:5972–7.
 60. Leco KJ, Waterhouse P, Sanchez OH, Gowing KL, Poole AR, Wakeham A, et al. Spontaneous air space enlargement in the lungs of mice lacking tissue inhibitor of metalloproteinases-3 (TIMP-3). *J Clin Invest*. 2001;108:817–29.
 61. Curiel D, Brantly M, Curiel E, Stier L, Crystal RG. Alpha 1-antitrypsin deficiency caused by the alpha 1-antitrypsin Nullmattawa gene. An insertion mutation rendering the alpha 1-antitrypsin gene incapable of producing alpha 1-antitrypsin. *J Clin Invest*. 1989;83:1144–52.
 62. Agusti A, MacNee W, Donaldson K, Cosio M. Hypothesis: does COPD have an autoimmune component? *Thorax*. 2003;58:832–4.
 63. Cosio MG, Saetta M, Agusti A. Immunologic aspects of chronic obstructive pulmonary disease. *N Engl J Med*. 2009;360:2445–54.
 64. Cosio BG, Agusti A. Autoimmunity in chronic obstructive pulmonary disease (COPD). *Arch Bronconeumol*. 2005;41:10–4.
 65. Kheradmand F, Shan M, Xu C, Corry DB. Autoimmunity in chronic obstructive pulmonary disease: clinical and experimental evidence. *Expert Rev Clin Immunol*. 2014;8:285–92.
 66. Taraseviciene-Stewart L, Scerbavicius R, Choe KH, Moore M, Sullivan A, Nicolls MR, et al. An animal model of autoimmune emphysema. *Am J Respir Crit Care Med*. 2005;171:734–42.
 67. Taraseviciene-Stewart L, Burns N, Kraskauskas D, Nicolls MR, Tudor RM, Voelkel NF. Mechanisms of autoimmune emphysema. *Proc Am Thorac Soc*. 2006;3:486–7.
 68. Taraseviciene-Stewart L, Douglas IS, Nana-Sinkam PS, Lee JD, Tudor RM, Nicolls MR, et al. Is alveolar destruction and emphysema in chronic obstructive pulmonary disease an immune disease? *Proc Am Thorac Soc*. 2006;3:687–90.
 69. Lee SH, Goswami S, Grudo A, Song LZ, Bandi V, Goodnight-White S, et al. Anti-elastin autoimmunity in tobacco smoking-induced emphysema. *Nat Med*. 2007;13:567–9.
 70. Kuo YB, Chang CA, Wu YK, Hsieh MJ, Tsai CH, Chen KT, et al. Identification and clinical association of anti-cytokeratin 18 autoantibody in COPD. *Immunol Lett*. 2010;128:131–6.
 71. Feghali-Bostwick CA, Gadgil AS, Otterbein LE, Pilewski JM, Stoner MW, Cszizmadia E, et al. Autoantibodies in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2008;177:156–63.
 72. Brandsma CA, Timens W, Geerlings M, Jekel H, Postma DS, Hylkema MN, et al. Induction of autoantibodies against lung matrix proteins and smoke-induced inflammation in mice. *BMC Pulm Med*. 2014;10:64.
 73. Barnes PJ. New anti-inflammatory targets for chronic obstructive pulmonary disease. *Nat Rev Drug Discov*. 2013;12:543–59.
 74. Stevens T, Ekholm K, Granse M, Lindahl M, Kozma V, Jungar C, et al. AZD9668: pharmacological characterization of a novel oral inhibitor of neutrophil elastase. *J Pharmacol Exp Ther*. 2014;339:313–20.
 75. Medicherla S, Fitzgerald MF, Spicer D, Woodman P, Ma JY, Kapoun AM, et al. p38alpha-selective mitogen-activated protein kinase inhibitor SD-282 reduces inflammation in a subchronic model of tobacco smoke-induced airway inflammation. *J Pharmacol Exp Ther*. 2008;324:921–9.
 76. Duong C, Seow HJ, Bozinovski S, Crack PJ, Anderson GP, Vlahos R. Glutathione peroxidase-1 protects against cigarette smoke-induced lung inflammation in mice. *Am J Physiol Lung Cell Mol Physiol*. 2014;299:L425–33.
 77. Vlahos R, Bozinovski S. Glutathione peroxidase-1 as a novel therapeutic target for COPD. *Redox Rep*. 2014;18:142–9.
 78. Rangasamy T, Cho CY, Thimmulappa RK, Zhen L, Srisuma SS, Kensler TW, et al. Genetic ablation of Nrf2 enhances susceptibility to cigarette smoke-induced emphysema in mice. *J Clin Invest*. 2004;114:1248–59.
 79. Massaro GD, Massaro D. Retinoic acid treatment partially rescues failed septation in rats and in mice. *Am J Physiol Lung Cell Mol Physiol*. 2000;278:L955–60.
 80. McGowan SE. Contributions of retinoids to the generation and repair of the pulmonary alveolus. *Chest*. 2002;121:2065–85.

81. Muyl JP, Muyl V, Kotnala S, Kumar D, Bhardwaj H. Therapeutic potential of growth factors in pulmonary emphysematous condition. *Lung*. 2013;191:147–63.
82. Martínez-Galan L, del Puerto-Nevado L, Pérez-Rial S, Díaz-Gil JJ, González-Mangado N, Peces-Barba G. Liver growth factor improves pulmonary fibrosis secondary to cadmium administration in rats. *Arch Bronconeumol*. 2010;46:20–6.
83. Cremona TP, Tschanz SA, von Garnier C, Benarafa C. SerpinB1 deficiency is not associated with increased susceptibility to pulmonary emphysema in mice. *Am J Physiol Lung Cell Mol Physiol*. 2013;305:L981–9.
84. Reynolds PR, Cosio MG, Hoidal JR. Cigarette smoke-induced Egr-1 upregulates proinflammatory cytokines in pulmonary epithelial cells. *Am J Respir Cell Mol Biol*. 2006;35:314–9.
85. Maeno T, Houghton AM, Quintero PA, Grumelli S, Owen CA, Shapiro SD. CD8+ T Cells are required for inflammation and destruction in cigarette smoke-induced emphysema in mice. *J Immunol*. 2007;178:8090–6.
86. Aoshiha K, Yokohori N, Nagai A. Alveolar wall apoptosis causes lung destruction and emphysematous changes. *Am J Respir Cell Mol Biol*. 2003;28:555–62.
87. Wright JL, Tai H, Churg A. Vasoactive mediators and pulmonary hypertension after cigarette smoke exposure in the guinea pig. *J Appl Physiol*. 2006;100:672–8.
88. Takubo Y, Guerassimov A, Ghezzi H, Triantafillopoulos A, Bates JH, Hoidal JR, et al. Alpha1-antitrypsin determines the pattern of emphysema and function in tobacco smoke-exposed mice: parallels with human disease. *Am J Respir Crit Care Med*. 2002;166:1596–603.
89. Churg A, Wang R, Wang X, Onnervik PO, Thim K, Wright JL. Effect of an MMP-9/MMP-12 inhibitor on smoke-induced emphysema and airway remodelling in guinea pigs. *Thorax*. 2007;62:706–13.
90. John G, Kohse K, Orasche J, Reda A, Schnelle-Kreis J, Zimmermann R, et al. The composition of cigarette smoke determines inflammatory cell recruitment to the lung in COPD mouse models. *Clin Sci*. 2014;126:207–21.
91. Glynos C, Dupont LL, Vassilakopoulos T, Papapetropoulos A, Brouckaert P, Giannis A, et al. The role of soluble guanylyl cyclase in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2013;188:789–99.
92. Lea S, Plumb J, Metcalfe H, Spicer D, Woodman P, Fox JC, et al. The effect of peroxisome proliferator-activated receptor-gamma ligands on in vitro and in vivo models of COPD. *Eur Respir J*. 2014;43:409–20.
93. Bracke KR, Verhamme FM, Seys LJ, Bantsimba-Malanda C, Cunoosamy DM, Herbst R, et al. Role of CXCL13 in cigarette smoke-induced lymphoid follicle formation and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2013;188:343–55.
94. Podolin PL, Foley JP, Carpenter DC, Bolognese BJ, Logan GA, Long E 3rd, et al. T cell depletion protects against alveolar destruction due to chronic cigarette smoke exposure in mice. *Am J Physiol Lung Cell Mol Physiol*. 2013;304:L312–23.
95. Eppert BL, Wortham BW, Flury JL, Borchers MT. Functional characterization of T cell populations in a mouse model of chronic obstructive pulmonary disease. *J Immunol*. 2013;190:1331–40.