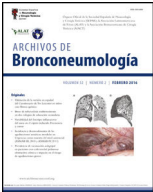




ARCHIVOS DE
Bronconeumología

www.archbronconeumol.org



Clinical Letter

The A209= Synonymous CFTR Variant Conceals Deep Intronic Variant: Disclosing its Phenotypic Spectrum

To the Director,

We present three cases of two unrelated families with cystic fibrosis (CF) confirmed diagnosis, high chloride concentrations, mild clinical phenotype and the synonymous CFTR A209= variant linked to a deep intronic variant affecting splicing, and in compound heterozygosity with another CFTR-causing variant. These findings underscore the need for deeper investigation of the synonymous variant, as its benign appearance might obscure its pathogenic role linked to the intronic variant.

Two maternal cousins (cases 1 and 2) and an unrelated male infant (case 3) were diagnosed with CF (Table 1). Case 1 initially presented with failure to thrive, later developing transient *Pseudomonas aeruginosa* colonization but has remained asymptomatic since. Case 2, diagnosed through neonatal screening, showed a mild phenotype without complications. Case 3, also diagnosed via neonatal screening, experienced a single episode of hyponatremic dehydration due to a medication error, with no further symptoms observed. All three cases demonstrate a mild clinical course, contrasting with the severity often associated with CF.

Genetic analysis of the CFTR gene was performed using genomic DNA extracted from blood samples. Targeted enrichment of exonic and flanking intronic regions was followed by massive parallel sequencing (NextSeq platform) to detect coding and intronic variants. To confirm the presence of the c.2989-313A>T variant,

PCR amplification of exon 18 was performed, followed by Sanger sequencing (AB3500 Genetic Analyzer, Applied Biosystems). Variant analysis was conducted in both index cases and their parents to assess segregation patterns and confirm the inheritance of the identified variants. The synonymous NM.000492.4:c.627A>G (p.Ala209=) variant was identified in linkage disequilibrium with the NM.000492.4:c.2989-313A>T and in compound heterozygosity with another pathogenic variant. mRNA analysis showed that the c.627A>G variant did not affect transcription.

The c.627A>G variant was first reported in 2001 by Le Maréchal et al.¹ as having no impact on the amino acid sequence in a CF patient. In 2019, Bergougnoux et al.² linked it to the deep intronic c.2989-313A>T variant, identifying it as a complex allele with a broad phenotypic spectrum ranging from mild chronic bronchitis to severe CF symptoms. In contrast, the three cases presented consistently exhibit a mild phenotype, with normal pulmonary function, no pancreatic insufficiency, and no significant health impacts, suggesting that the intronic variant may lead to milder clinical manifestations in these patients. Given their favorable clinical course, none of them have required CFTR modulators.

While Bergougnoux et al. provided data on symptom onset, they did not specify the exact age of diagnosis, making it difficult to assess whether these patients had access to early detection or long-term follow-up. In contrast, our patients were diagnosed within the first year of life due to newborn screening or early clinical suspicion, allowing for timely intervention and continuous monitoring, which may have influenced their milder presentation. Additionally, the treatment regimens received by the 2019 cohort were not explicitly documented, making it difficult to determine the impact

Table 1
Demographic, Genotyping and Clinical Data of the Three Reported Patients.

Case	Gender	DoB	1st Allele	2nd Allele	IRT (ng/mL)	Sweat Chloride (mmol/L)	Pulmonary Assessment	Pancreatic Insufficiency	Other Symptoms
1	F	2011	c.1000C>T (R334W)	c.627A>G (A209=)+c.2989-313A>T	ND	76 and 81	PFT normal CCTS normal	No	Failure to thrive (at 12 months) <i>Pseudomonas aeruginosa</i> colonization (at 15 months) Asymptomatic
2	M	2018	c.1624G>T (G542X)	c.627A>G (A209=)+c.2989-313A>T	110	69 and 106	ND	No	
3	M	2022	c.1624G>T (G542X)	c.627A>G (A209=)+c.2989-313A>T	55	69 and 79	PFT normal	No	Hyponatremic dehydration (medication error; 0.9% solution instead of 20%)

DoB: date of birth; TIR: immunoreactive trypsinogen; ND: not determined; NA: not applicable; PFT: pulmonary function test; CCTS: chest CT scan.

of clinical management and healthcare accessibility on disease progression. This raises the question of whether the milder phenotype observed in our cohort is primarily due to early diagnosis and proactive medical care or whether other genetic or environmental modifiers contribute to the observed differences.

The c.627A>G variant, while considered benign, gains significance due to its consistent linkage disequilibrium with the pathogenic intronic c.2989-313A>T variant. This deep intronic variant generates a pseudo-exon, leading to a premature stop codon and potentially truncated CFTR protein,² as seen in all three cases. Its co-occurrence with the synonymous variant highlights the need for thorough genetic investigation, as routine gene analysis often excludes intronic regions.

Although mRNA analysis is a direct method for characterizing splicing variants,³ its limitations in accessibility and sample stability make it less practical.⁴ For detecting the c.2989-313A>T variant linked to c.627A>G, cost-effective methods like Sanger sequencing are sufficient and more feasible for routine diagnostics.

Statement of Ethics

The authors have no ethical conflicts to disclose.

Contribution of Each Author

FMB: Conception and study design, data acquisition, data analysis, manuscript drafting, and final manuscript approval.

GGR: Data analysis, manuscript drafting, and final manuscript approval.

CPM: Writing – review and editing, and final manuscript approval.

OMM: Writing – review and editing, and final manuscript approval.

ACC: Writing – review and editing, and final manuscript approval.

All authors have contributed substantially to obtaining the results and preparing the manuscript in accordance with the ICMJE criteria.

Artificial Intelligence Involvement

No part of this manuscript has been produced with the help of artificial intelligence software or tools.

Funding of the Research

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

The authors have not received any funding specifically for this correspondence.

Conflicts of Interest

The authors declare not to have any conflicts of interest that may be considered to influence directly or indirectly the content of the manuscript.

The authors have no conflicts of interest to declare.

References

1. Le Maréchal C, Audrézet MP, Quéré I, Raguénès O, Langonné S, Férec C. Complete and rapid scanning of the cystic fibrosis transmembrane conductance regulator (CFTR) gene by denaturing high-performance liquid chromatography (D-HPLC): major implications for genetic counselling. *Hum Genet.* 2001;108(4):290–8, <http://dx.doi.org/10.1007/s004390100490>.
2. Bergougnoux A, Délétang K, Pommier A, Varilh J, Houriez F, Altieri JP, et al. Functional characterization and phenotypic spectrum of three recurrent disease-causing deep intronic variants of the CFTR gene. *J Cyst Fibros.* 2019;18(4):468–75, <http://dx.doi.org/10.1016/j.jcf.2018.10.012>.
3. Costa C, Pruliere-Escabasse V, de Becdelievre A, Gameiro C, Golmard L, Guittard C, et al. A recurrent deep-intronic splicing CF mutation emphasizes the importance of mRNA studies in clinical practice. *J Cyst Fibros.* 2011;10(6):479–82, <http://dx.doi.org/10.1016/j.jcf.2011.06.011>.
4. Will K, Dörk T, Stuhmann M, von der Hard H, Ellemunter H, Tümmler B, et al. Transcript analysis of CFTR nonsense mutations in lymphocytes and nasal epithelial cells from cystic fibrosis patients. *Hum Mutat.* 1995;5(3):210–20, <http://dx.doi.org/10.1002/humu.1380050305>.

Francisco Martínez Bugallo^{a,*}, Gema García de la Rosa^a,
Carol Prieto-Morín^a, Orlando Mesa-Medina^b,
Alicia Callejón-Callejón^b

^a Human Genetic Unit, Department of Clinical Analysis Laboratory, University Hospital Nuestra Señora de Candelaria, Santa Cruz de Tenerife, Spain

^b Pediatric Pulmonology Unit, Department of Pediatrics, University Hospital Nuestra Señora de Candelaria, Santa Cruz de Tenerife, Spain

*Corresponding author.

E-mail address: fmarbug@gobiernodecanarias.org
(F. Martínez Bugallo).